

microRNA调控肿瘤形成相关的因素及信号通路

黄玲巍^{1,2} 张震宇^{1,2} 王家敏^{1,2,3} 乔自林^{1,2,3} 阿依木古丽·阿不都热依木^{1,2} 杨迪^{1,2,4*}

(¹西北民族大学, 生物医学研究中心, 甘肃省动物细胞技术创新中心, 兰州 730030; ²西北民族大学, 生物医学研究中心, 细胞基质疫苗关键技术与产业化教育部工程研究中心, 兰州 730030; ³西北民族大学, 生物医学研究中心, 生物工程与技术国家民委重点实验室, 兰州 730030; ⁴西北民族大学, 实验教学部, 兰州 730030)

摘要 miRNA(microRNA)作为单链非编码RNA, 通过结合靶mRNA的非翻译区而抑制蛋白质编码基因的表达。已有研究表明, miRNA可通过ceRNA、外泌体、环境等因素调控肿瘤的形成。绝大多数miRNA对肿瘤细胞的增殖、凋亡与自噬、细胞周期、细胞的迁移和侵袭以及能量代谢具有调节作用, 故miRNA可作为一种新颖的生物标志物, 并已成为肿瘤治疗中极具吸引力的靶点。同时, 该文回顾了miRNA调控肿瘤形成相关的信号通路, 并深入讨论了JAK/STAT3、Wnt/ β -catenin、PI3K/AKT三种常见的信号通路, 旨在为了解肿瘤发生发展的机制提供良好的基础, 为发现新的肿瘤治疗途径和肿瘤治疗靶点奠定基础。

关键词 miRNA; 肿瘤形成; 信号通路

Factors and Signaling Pathways Related to microRNA Regulation of Tumor Formation

HUANG Lingwei^{1,2}, ZHANG Zhenyu^{1,2}, WANG Jiamin^{1,2,3}, QIAO Zilin^{1,2,3}, AYIMUGULI Abudureyimu^{1,2}, YANG Di^{1,2,4*}

(¹Gansu Tech Innovation Center of Animal Cell, Biomedical Research Center, Northwest Minzu University, Lanzhou 730030, China; ²Engineering Research Center for Key Technologies and Industrialization of Cell-Based Vaccines, Ministry of Education, Biomedical Research Center, Northwest Minzu University, Lanzhou 730030, China; ³Key Laboratory of Bioengineering and Technology State Ethnic Affairs Commission, Biomedical Research Center, Northwest Minzu University, Lanzhou 730030, China; ⁴Department of Experiment & Teaching, Northwest Minzu University, Lanzhou 730030, China)

Abstract miRNA (microRNA) acts as a single-stranded non-coding RNA that inhibits the expression of protein coding genes through binding to the untranslated regions of target mRNA. Previous research has established that miRNA can regulate tumor formation through some factors such as ceRNA, exosomes and environment. The majority of miRNA exhibits regulatory effects on essential processes of tumor cells, including proliferation, apoptosis and autophagy, cell cycle, cell migration and invasion, energy metabolism and so on. Thus, miRNAs has emerged as a promising biomarker for targeted-tumor therapy. This article undertakes an in-depth review of the signaling pathways linked with miRNA regulation of tumor formation, with a particular focus on the interplay among JAK/STAT3, Wnt/ β -catenin and PI3K/AKT pathways. By providing a sound basis for understanding the mechanisms involved in tumor genesis and development, this article aims to lay the groundwork for discovering new tumor treatment pathways and cancer therapy targets.

Keywords miRNA; tumor formation; signaling pathways

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*通讯作者。Tel: 0931-2938313, E-mail: xbmzyd@163.com

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*Corresponding author. Tel: +86-931-2938313, E-mail: xbmzyd@163.com

微小RNA(microRNA, miRNA)作为一种内源性且相对保守的长度仅为21~25个核苷酸的单链非编码RNA,在胚胎发生、细胞分化、血管生成、神经发育以及肿瘤发生等多种细胞生理事件中发挥重要作用^[1-2]。研究发现,体细胞突变以及肿瘤细胞表观基因组的改变会解除细胞内信号通路的调控,最终导致肿瘤细胞的异常生长、生存和侵袭。miRNA可通过表观遗传基因调节多种生物学过程(包括调控细胞增殖、迁移、侵袭、糖酵解、细胞凋亡、肿瘤干细胞样表型、化疗耐药和上皮-间充质转化等)从而控制肿瘤的发生和发展^[3]。miRNA已被证明可以抑制重要的癌症相关基因的表达,并有望在癌症的诊断和治疗中发挥作用^[4],已有研究表明miRNA功能丧失可影响癌症的进展,因此,miRNA替代疗法已成为治疗恶性肿瘤的有希望的治疗策略^[5]。为了更加清晰地阐明miRNA调控肿瘤形成相关的信号通路,经过查阅相关文献,本文对miRNA调控肿瘤发生的研究现状和涉及的主要信号通路进行阐述,为发现新的肿瘤治疗途径以及癌症治疗靶点提供理论依据。

1 miRNA参与调控肿瘤形成

1.1 竞争内源性RNA调控

miRNA可与编码蛋白质的mRNA的3'-非翻译区(3'-UTR)结合,在转录后抑制蛋白质编码基因表达,从而对进化产生影响,因此其在动物、植物、微生物中发挥重要的基因调控作用^[6]。miRNA的功能主要由第2到7位的核苷酸组成的种子区决定,miRNA可与mRNA靶标的3'-UTR中的一个或多个互补序列结合^[7]。miRNA与其宿主基因共享启动子或独立转录,可通过调节与宿主生物学功能相关的靶基因,与宿主基因发挥协同或拮抗作用^[8];miRNA能够同时抑制多个靶基因,被认为是肿瘤发生过程的关键介质^[9],亦有学者认为健康细胞分泌的miRNA可向附近的癌细胞发送生长抑制信号^[10]。随着有关非编码RNA的研究的飞速发展,科学家们发现作用于肿瘤细胞的非编码RNA并非由单一RNA组成的。

竞争内源性RNA(competing endogenous RNA, ceRNA)是一类转录本,可以在转录后竞争性地调节miRNA,并可以通过ceRNA-miRNA-mRNA轴参与广泛的生物过程^[11]。ceRNA调控机制以lncRNA-miRNA-mRNA和circRNA-miRNA-mRNA调控网络为主,相关研究已经证实了ceRNA通过调控miRNA在肿瘤

发生发展中发挥作用^[12]。在表1中,作者通过查阅文献总结分析了近三年(2021年~2023年)在人类肿瘤疾病的研究中受ceRNA网络调控的miRNA,并对miRNA的靶基因的作用途径及影响作用作简要说明。

1.2 外泌体调控

外泌体参与了体内肿瘤转移、血管生成、免疫逃逸、代谢和耐药的过程,miRNA通过外泌体转移到受体细胞,此过程构成了生理细胞通讯的一种重要形式。受外泌体调控的非编码RNA可以作为内源性调节因子,直接与癌症的发生和进展相关^[22]。例如,在胃癌患者血清外泌体中发现miR-122-5p表达下调抑制胃癌细胞增殖和转移,且miR-122-5p在体内抑制肿瘤生长,该过程与miR-122-5p靶基因*GIT1*有关^[23]。表2中整理了近年来作为肿瘤标志物的外泌体miRNA,并对其靶基因及影响作用作了简要说明。从表2中已收集到的文献中可以推断,外泌体大多来源于血浆或血清,外泌体中的miRNA通过靶基因调控肿瘤的形式多样,并且转运到癌细胞中的miRNA会对肿瘤微环境产生影响。

1.3 环境因素调控

研究表明,环境因素会增加实体肿瘤血管生成、生长因子和基因的不稳定性,并促进肿瘤侵袭和转移^[36]。缺氧能够降低癌细胞中核糖核酸酶3和Dicer酶的表达量,抑制AGO2(Argonaute 2)的磷酸化,干扰Dicer与AGO2的结合,抑制miRNA从前体到成熟体的加工^[37]。研究发现非小细胞肺癌(non-small cell lung cancer, NSCLC)肿瘤组织中*Bcl-w*表达与缺氧有关,miR-519d-3p可通过调节*Bcl-w*和缺氧诱导因子(hypoxia-inducible factor 1-alpha, *HIF-1a*)的表达抑制缺氧诱导的肿瘤发生,可作为NSCLC的诊断和靶向治疗标志物^[38]。缺氧时,骨髓细胞通过下调干扰因子调节因子4(interferon regulatory factor 4, *IRF4*)来获得抗凋亡能力,此时miR-210表达水平上调,这与18S RNA碱基甲基转移酶DIMT1直接下调有关^[39]。在未成熟胶质母细胞瘤细胞缺氧时,miR-210表达同样上调^[40]。综上所述,缺氧会抑制miRNA的加工进程,从而影响细胞的正常生理过程,引发癌变并阻碍相关癌症的诊断。

2 miRNA对肿瘤细胞的影响

2.1 影响细胞增殖

*Glypicans*作为生长因子的辅助受体影响细胞

表1 受ceRNA网络调控的miRNA(2021年~2023年)
Table 1 miRNA regulated by ceRNA networks (2021-2023)

| ceRNA类型 ceRNA types | miRNA类型 miRNA types | 疾病 Diseases | 靶基因/途径 Target gene/pathway | 影响作用 Influence | 参考文献 Reference |
|------------------------|--|-----------------------------------|-------------------------------|---|-------------------|
| MALAT1 | miR-26a/26b | Breast cancer | <i>ST8SIA4</i> | It regulates the tumorigenicity of breast cancer cells | [13] |
| LncRNA ABHD11-AS1 | Many miRNAs | Gastric cancer and thyroid cancer | PI3K/AKT | Clinical biomarker | [14] |
| circ-0005105 | miR-20a-3p | Pancreatic ductal adenocarcinoma | <i>COL11A1</i> | Reduce tumorigenicity and metastasis of pancreatic duct adenocyte | [15] |
| circ-ABCB10 | miR-588 | Squamous carcinoma of larynx | <i>CXCR4</i> | Involved in malignant progression of laryngeal squamous cells | [16] |
| lncRNA AGAP2-AS1 | miR-3064-5p | Cervical cancer | <i>SIRT1</i> | Promote cell proliferation | [17] |
| circ_0001402 | miR-625-5p | Squamous cell carcinoma of skin | <i>KPNA4</i> | It affects the tumorigenicity of squamous cell carcinoma of skin | [18] |
| LncRNA HULC | miR-372 | Hepatocellular carcinoma | <i>PRKACB</i> | Influence tumor growth | [19] |
| LncRNA RPPH1 | miR-122 | Hepatocellular carcinoma | Wnt/ β -catenin | Provide therapeutic targets for patients with liver cancer | [20] |
| lncRNAs, mRNAs | hsa-miR-17, hsa-miR-93, hsa-miR-150, hsa-miR-25, hsamiR-125b | Prostatic cancer | <i>PCA3, H19, RND3, ITGB8</i> | Diagnostic biomarkers for prostate cancer | [21] |

表2 作为潜在肿瘤标志物的外泌体miRNA
Table 2 Exosome miRNA as potential tumor marker

| 外泌体来源 Exosome source | miRNA类型 miRNA types | 疾病 Diseases | 靶基因/途径 Target gene/pathway | 影响作用 Influence | 参考文献 References |
|-------------------------|--|---------------------------|-------------------------------|--|--------------------|
| Plasma | miR-125a-5p/miR-141-5p | Prostate cancer | PI3K/AKT/mTOR | Indirectly predict the possibility of tumor development | [24] |
| Plasma | miR-21, miR-181a-5p | Thyroid carcinoma | <i>OTUD6B-AS1</i> | Differentiation of different thyroid cancers | [25] |
| Serum | miR-25, miR-130b, miR-425 | Colorectal cancer | PTEN/PI3K/AKT | Promote metastasis of colorectal cancer | [26] |
| Serum | miR-1247-3p | Liver cancer | <i>B4GALT3</i> | Promote lung metastasis of liver cancer | [27] |
| Serum | miR-139-5p | Bladder cancer | <i>PRCI</i> | Inhibition of tumor formation | [28] |
| Serum | miR-7977 | Lung adenocarcinoma | Unknown | Promote A549 cell proliferation and invasion, and inhibit cell apoptosis | [29] |
| Serum | miR-1468-5p | Cervical cancer | <i>HMBOX1</i> & JAK2/STAT3 | Promote tumor immune escape | [30] |
| Wide range of sources | miR-21 | Various cancers | Multiple pathways | Promote the development of cancer | [31] |
| Serum and tissue | miR-21-5p, miR-142-5p, miR-150-5p, miR-320a-3p, miR-4433b-5p | Breast cancer | Multiple pathways | Promote chemotherapy resistance | [32-33] |
| Plasma and macrophage | miR-223, miR-320d, miR-4479, miR-6763-5p | Epithelial ovarian cancer | PTEN/PI3K/AKT | Promote chemotherapy resistance | [34-35] |

增殖,是miR-509-3p在黑色素瘤中的作用靶点^[41]。*Glypican-3*是一种癌基因,在肝细胞癌(hepato cellular carcinoma, HCC)和肝母细胞瘤等肝脏恶性肿瘤中频繁上调,miR-4510通过直接靶向*Glypican-3*并调控Wnt/ β -catenin信号通路的转录活性来实现抑癌作用^[42]。miR-125a过表达可抑制人胚肾细胞系293T(human embryonic kidney cell 293T, HEK293T)细胞的增殖,miR-125a通过靶向*Glypican-4*基因影响细胞增殖,增加了miRNA作为抑制细胞增殖的治疗策略的可行性^[43]。*GPC5*(*Glypican-5*)是硫酸乙酰肝素蛋白多糖的一员,可抑制肿瘤细胞增殖,抑制miR-301b会使*GPC5*的表达上调从而抑制胶质瘤细胞的增殖和侵袭^[44]。由此可见,miRNA对肿瘤细胞的影响绝大多数是通过调控不同的生长因子实现的,其通过这种途径只是对肿瘤的增殖起到简单的外部介导作用,随着研究的不断深入,科研人员发现miRNA也可通过介导基因的转录来影响肿瘤的发生。

对基因转录水平的研究更清晰地阐明了miRNA如何通过不同的基因影响肿瘤的增殖,为更好地研究miRNA调控肿瘤形成的机制提供了理论依据。HCC细胞中miR-15a低表达可靶向上调转录因子4,抑制HCC细胞的增殖、迁移和侵袭,抑制体内HCC成瘤和转移从而延缓HCC的发生发展^[45]。在上皮性卵巢癌细胞(epithelial ovarian cancer cell, EOCC)中发现抑制miR-200a表达可使上皮-间充质转化相关转录因子*ZEB2*表达上调,影响细胞成瘤,miR-200a可能是EOCC预后评估和治疗干预的潜在标志物^[46]。过表达miR-339通过靶向调控*ZNF689*基因的表达,显著抑制胃癌细胞的增殖、侵袭和迁移^[47]。表观遗传抑制因子促进细胞增殖,miR-150通过直接和间接机制调控表观遗传抑制因子*NMT3A*和*DNMT3B*的水平。miR-150作为一种肿瘤抑制因子,通过提高肿瘤抑制因子的水平和降低细胞周期调节因子的水平,下调表观遗传抑制因子,抑制细胞增殖并诱导S期阻滞^[48]。

2.2 影响细胞凋亡与自噬

miRNA主要通过影响凋亡通路及自噬通路相关基因和蛋白质之间的相互作用,从而影响肿瘤的发生发展。肿瘤干细胞(cancer stem cells, CSCs)因其多能性而能够推动肿瘤的发生。ZHOU等^[49]从两个胰腺癌异种移植物中分离出两种不同表型的胰腺癌干细胞(pancreatic cancer stem cells, PCSCs),通

过全转录组测序发现479个mRNA和15个miRNA在PCSCs中特异性表达,其中miR-146b-3p下调最显著。miR-146b-3p在胰腺癌组织和细胞系MIA Paca-2(CSC^{high})中表达下调。恢复miR-146b-3p表达可抑制MIA Paca-2(CSC^{high})的细胞增殖,此过程通过增加细胞的G₁期、减少S期,诱导细胞凋亡,最终抑制细胞成瘤;进一步研究发现,miR-146b-3p可通过作用于DYRK2和GLI2从而激活Hedgehog通路并直接靶向MAP3K10。miR-7-5p通过下调丝裂铁蛋白从而降低铁水平来抑制抗辐射细胞铁死亡,并且miR-9和miR-137通过降低细胞内谷胱甘肽水平来促进铁死亡^[50]。在胶质母细胞瘤亚型的研究中发现miR-93的异常表达与细胞自噬有关,miR-93可影响胶质瘤干细胞(glioma stem cells, GSCs)表型以及耐药性,miR-93通过同时抑制多种自噬调节因子改变GSC自噬活性,揭示了miR-93在自噬调节中的关键作用^[51]。miR-124在胶质瘤组织中的表达降低,可导致细胞外基质金属蛋白酶诱导物的表达增加,从而抑制胶质瘤发展^[52]。在神经胶质瘤的研究中发现,调控细胞自噬的mRNA与AKT2相互作用,AKT2抑制miR-193a-3p表达后可抑制神经胶质瘤细胞增殖并促进细胞凋亡^[53]。

2.3 影响细胞周期

研究表明,细胞周期蛋白依赖性激酶1(cyclin-dependent kinase 1, CDK1)参与宫颈癌细胞周期改变,在宫颈癌细胞中miR-495-3p和miR-143-3p可共同靶向CDK1,导致其表达下调,抑制宫颈癌细胞增殖,促进细胞凋亡,在体内降低宫颈癌细胞的成瘤性^[54]。lncRNA EVADR在结直肠癌(colorectal cancer, CRC)中高度上调,在EVADR的全长cDNA(EVADR-v1)扩增过程中,发现了一个新的短变体(EVADR-v2)。生物信息学分析和双荧光素酶检测证实,EVADR可作为海绵与miR-7和miR-29b相互作用,EVADR-v1/-v2过表达后,CRC细胞的细胞周期和迁移情况发生改变,早期/晚期凋亡率降低,Bax/Bcl2值降低^[55]。GHEIDARI等^[56]发现miR-429在多形性胶质母细胞瘤(glioblastoma, GBM)中表达下调,并利用生物信息学工具预测了几个与ERBB信号通路相关的靶基因;在GBM细胞中用慢病毒载体过表达miR-429,结果发现miR-429通过ERBB途径直接靶向癌基因*MYC*、*BCL2*和*EGFR*,并显著降低靶基因的表达水平,诱导细胞周期停

滞。在肝癌放疗耐药组织和细胞中, miR-302a-3p的表达量显著降低, 细胞周期抑制剂可逆转miR-302a-3p下调对肝癌放疗敏感性的抑制作用; 进一步的研究表明 miR-302a-3p/MCL1轴可通过诱导G₀/G₁期停滞来增强肝癌细胞的放射敏感性^[57]。HASSAN等^[58]发现, 通过使用miR-122模拟物和miR-221抑制剂靶向*SENPI*和*ARF4*基因, 可使细胞周期蛋白D1、转化生长因子-β和β-连环蛋白基因的表达下调, 诱导肿瘤的凋亡和坏死。ZHOU等^[59]发现, *CDK1*在肝细胞癌组织和细胞中高表达, 沉默*CDK1*可调节肝癌细胞的细胞周期, 抑制DNA复制和细胞增殖。在肝癌细胞中, miR-195-5p可降低*CDK1*水平, 且抑制G₁期到S期的转变, 诱导DNA损伤反应, 抑制DNA复制和细胞增殖。综上所述, miRNA可以通过调控*CDK1*、细胞周期蛋白D1、细胞周期通路相关基因、G₀/G₁期停滞以及抑制G₁期到S期的转变来影响肿瘤细胞的细胞周期, 这可为癌症的临床治疗提供理论依据。

2.4 影响细胞迁移和侵袭

癌组织易侵袭和转移通常会导导致预后不良, 例如高级别卵巢浆液性癌是一种因癌细胞转移而预后不良的卵巢癌, 迫切需要新的治疗靶点, miRNA通过调节下游靶基因及相关通路, 提高或降低肿瘤细胞的迁移和侵袭能力。LIU等^[60]发现, 细胞质聚腺苷酸化元件结合蛋白3(cytoplasmic polyadenylation element binding protein 3, CPEB3)作为miR-301b-3p的靶蛋白与其表达水平呈负相关, 并在高级别卵巢浆液性癌组织和细胞系中下调, miR-301b-3p通过诱导表皮生长因子受体和下游转移相关蛋白p38及细胞外信号调节激酶1/2(extracellular signal-regulated kinase 1/2, ERK1/2)生成, 最终靶向CPEB3/EGFR轴并加速高级别卵巢浆液性癌的迁移和侵袭。无独有偶, 骨转移瘤在影响四肢骨骼的同时容易发生肺转移。骨肉瘤相关体外实验的结果表明, *RAB22A*作为癌基因家族的成员是miRNA-151a-3p的潜在靶基因, 受lncRNA SNHG3正调控, 但受miR-151a-3p负调控, miR-151a-3p敲除可以促进骨肉瘤细胞的侵袭和迁移, *RAB22A*的过度表达也可促进骨肉瘤细胞的侵袭和迁移^[61]。随着研究的深入, ZHANG等^[62]发现miR-338-3p过表达可通过靶向MAP3K2调控ERK1/2信号通路, 从而抑制人肺腺癌细胞的迁移、侵袭和增殖, 此研究加入miR-338-3p抑制剂后, *MAP3K2*、*p-*

ERK1/2、*MMP2*、*MMP3*、*MMP9*、细胞周期蛋白A2和细胞周期蛋白D1的表达水平升高, 导致人肺腺癌细胞的迁移和侵袭增加, 而加入miR-338-3p模拟物的结果与此相反。LIU等^[63]发现抑制miR-196-5p的表达可显著抑制胆管癌细胞的增殖和转移, 且*HAND1c*和β-catenin沉默均能逆转miR-196-5p上调带来的细胞异常增殖和迁移, 所以*HAND1/Wnt/β-catenin*信号通路的激活是miR-196-5p发挥作用的重要条件。

2.5 影响细胞能量代谢

一些miRNA对多个代谢基因进行正向和负向调节, 使肿瘤细胞能够在不利条件下存活, 并产生恶性肿瘤的相关特性, 如细胞黏附、迁移和侵袭增加, 三羧酸循环和头合成脂肪酸减弱、无氧糖酵解和自噬增加。一些信号通路因子、低氧诱导因子、磷脂酰肌醇-3激酶、蛋白激酶B、哺乳动物靶标雷帕霉素、磷酸酶以及胰岛素信号通路中的张力蛋白的同系物, 均受miRNA调控^[64]。QUIRICO等^[65]通过分析miR-455-3p、miR-122、miR-30a-5p、miR-203、miR-181d、miR-7、miR-489-3p、miR-155、miR-422a和miR-146b-5p后发现在癌症中, miRNA可控制糖酵解途径中葡萄糖的代谢。与此同时, SUN等^[66]发现通过PI3K/AKT/mTOR/HIF-1α途径下调miR-21的表达, 可抑制顺铂耐药的肺癌细胞株的糖代谢而促进癌细胞死亡。miR-125还可以和天然化合物结合来共同调节与癌症相关的线粒体功能障碍和能量代谢^[67], 由此可见, miRNA在癌细胞的代谢重编程中发挥着非常重要的作用, 并且可同时调控转录和转录后水平。

3 miRNA调控肿瘤形成相关的主要信号通路

恶性肿瘤的发生发展始于一系列遗传和表观遗传事件, 这导致多种信号通路发生改变, 靶向这些信号通路也被认为在癌症治疗中具有临床价值^[68]。作者通过查阅文献在表3中列举出了miRNA调控肿瘤形成相关的信号通路。如表3所述, 交叉分析可知miRNA调控肿瘤形成相关的主要通路为Wnt/β-catenin信号通路、JAK(Janus kinase)/STAT3(signal transducer and activator of transcription 3)信号通路、PI3K/AKT/mTOR信号通路。miRNA占基因的1%~3%, 但调节人类30%以上基因的表达, 可见miRNA通过调节基因的表达, 参与调控肿瘤形成相关的信号通路, 在有关肿瘤的研究

表3 miRNA影响肿瘤形成相关的信号通路

Table 3 miRNA affected signaling pathways related to tumor formation

| miRNA类型 miRNA types | 信号通路 Signaling pathway | 疾病 Disease | 影响作用 Influence | 参考文献 Reference |
|-------------------------|---|--|---|-------------------|
| Many miRNAs | PI3K/AKT/mTOR, Wnt/ β -catenin, JAK/STAT, MAPK | Breast cancer | Provide treatment options | [69] |
| Many miRNAs | Notch, TGF- β , Wnt, STAT3, AKT, EGFR | Glioma | Study the pathogenesis and regulation of drug resistance | [70] |
| Many miRNAs | Wnt/ β -catenin, PTEN/AKT/mTOR, TGF- β , KRAS, VEGFR, EGFR, p53 | Colorectal cancer | Control of cellular processes in colorectal cancer | [71] |
| Twenty-two kinds miRNAs | PI3K/AKT, Wnt, AMPK, MAPK | Fourteen types of cancer | Play a role in cancer inhibition | [72] |
| miR-411 | MAPK, PI3K/AKT/mTOR, p53, Ras, NF- κ B, Wnt/ β -catenin | Multiple cancers | Regulate cell proliferation, invasion, migration, apoptosis and colony, and the formation of cancer cells | [73] |
| miR-1275 | PI3K/AKT, ERK/JNK, MAPK, Wnt | Liver cancer, breast cancer, lung cancer, gastrointestinal cancer, urogenital tract cancer | It helps to study the development of many types of cancer | [74] |

中意义重大。

3.1 JAK/STAT3信号通路

miRNA通过直接或间接影响细胞因子信号转导抑制因子家族成员JAK、STAT3及调控其他细胞增殖、凋亡、迁移、侵袭和EMT相关基因的表达从而参与肿瘤形成^[75]。抗癌药物可通过miRNA来抑制JAK/STAT3信号通路, 目前正在骨肉瘤细胞^[76]和肺癌细胞^[77]中得到证明。JAK/STAT3信号参与细胞增殖、细胞周期等多种生物学过程^[78-79], 因此, miRNA可通过细胞内外的不同途径激活JAK/STAT3信号通路来影响细胞的增殖。STAT3在激活前存在于细胞质中, 在细胞因子和致癌因子的作用下, STAT3磷酸活化并在细胞核中过表达, 此过程促进细胞增殖和恶性转化, 阻碍细胞凋亡, 具有致癌作用^[80]。

miR-301a过表达会增强胰腺癌细胞的侵袭、血管生成和迁移能力, 抑制靶基因细胞因子信号抑制因子5(suppressor of cytokine signaling 5, SOCS5)的表达, 导致JAK/STAT3激活^[81]。卵巢肿瘤细胞外小泡富含miR-141-3p, 转染miR-141-3p后的小细胞外小泡使SOCS5的表达水平显著降低, 导致内皮细胞JAK/STAT3通路激活, miR-141-3p能够促进内皮细胞迁移和血管生成, 进而调控卵巢肿瘤发病机制^[82]。GSCs是胶质瘤发生和发展的关键, miR-30在原发性胶质瘤细胞的GSCs中过度表达, 而下调miR-30则能抑制JAK/STAT3通路, 降低GSCs的致癌性。miR-30通过靶向mRNA的3'-UTR来降低SOCS3的表达量^[83],

应用同样的原理, miR-30e-5p通过靶向泛素特异性肽酶22(ubiquitin specific peptidase 22, USP22)介导的JAK/STAT3信号, 抑制NSCLC肿瘤的发生^[84]。另外, NSCLC患者的miR-410表达水平较高, SOCS3表达水平较低, miR-410通过靶向抑制SOCS3的表达, 促进JAK/STAT3信号通路的激活和细胞增殖, 为未来肺癌的治疗提供了线索^[85]。miR-500a-3p水平在肝癌组织和细胞中显著升高, 上调miR-500a-3p可增强体外肿瘤干细胞的球体形成能力, 并增加其数量比例和肿瘤相关因子的表达水平, miR-500a-3p通过靶向JAK/STAT3信号通路的多种负性调节因子, 包括SOCS2、SOCS4和PTPN11, 促进肿瘤细胞增殖; STAT3信号通路激活, 使得miR-500a-3p下调并沉默细胞中的SOCS2、SOCS4和PTPN11的表达, 这个过程可逆转miR-500a-3p过表达引起的STAT3信号活性抑制和肿瘤细胞中癌症干细胞表型增加的现象^[86]。

另有研究发现, JAK1和JAK2的口服抑制剂通过抑制白细胞介素-6(interleukin 6, IL-6)受体依赖性JAK/STAT信号转导在胶质母细胞瘤血管生成中发挥重要作用, 抑制剂治疗诱导的miR-17a-3p和miR-20a过表达可能在下调JAK2、STAT3和PI3K蛋白中起主要作用^[87]。在宫颈腺癌中, miR-9启动子高甲基化后, 下调并抑制HeLa细胞的IL-6表达从而发挥抑癌作用, HeLa细胞去甲基化处理可增加成熟miR-9的表达量, 消除其对IL-6的抑制, 使IL-6/JAK/STAT3通路激活, 最终产生抑癌作用^[88]。miR-515-5p的过表达

抑制了肝癌细胞在体外和体内的迁移和侵袭, miR-515-5p可直接结合到IL-6的3'-UTR, 抑制JAK/STAT3信号通路的激活, IL-6的过表达则使JAK/STAT3信号通路激活^[89]。图1总结了miRNA参与JAK/STAT3信号通路调控的机制, 并通过调节该通路影响肿瘤细胞的增殖情况。

3.2 Wnt/ β -catenin信号通路

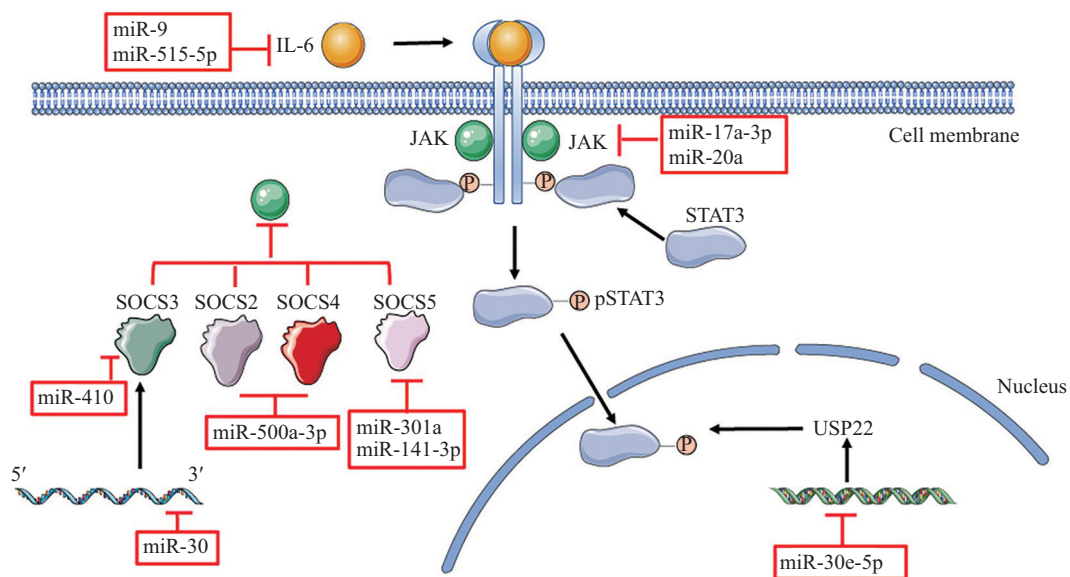
Wnt/ β -catenin参与人类的发育、生长和细胞内稳态维持^[90], Wnt通过 β -catenin激活靶基因的表达^[91]。 β -catenin是一种可在细胞间连接处与细胞黏附分子E-钙黏蛋白相互作用的蛋白质, 已被证明是肿瘤发生中Wnt信号通路的最重要的介导体之一^[92]。CSCs几乎在所有种类的癌症中都能被检测到, 具有自我更新和分化能力, 可增强肿瘤细胞的成瘤性。miRNA通过控制干细胞自我更新的级联信号, 即Wnt、Notch和Hedgehog通路, 影响CSCs的功能^[93]。

在Wnt/ β -catenin信号通路中, miRNA主要通过调控信号通路中的关键因子, 如以相关蛋白及基因为靶点, 影响肿瘤细胞的成瘤性。在研究直肠癌

干细胞(colorectal cancer stem cells, CRCSCs)自我更新的潜在机制过程中发现, miR-8063的表达明显下调, 异质核糖核蛋白A/B(heterogeneous nuclear ribonucleoprotein A/B, hnRNP A/B)的表达明显上调, 当hnRNP A/B过表达后, 直肠癌干细胞集落形成能力和成瘤性增强, Wnt/ β -catenin信号通路中关键蛋白Wnt3a、Wnt5a和 β -catenin的表达上调, 而miR-8063的表达缺失会削弱其对hnRNP A/B的抑制作用, miR-8063在CRCSCs中过表达后, hnRNP A/B、Wnt3a、Wnt5a和 β -catenin的表达下调, CSCs的自我更新能力减弱, miR-8063可激活Wnt/ β -catenin信号通路, 促进CRCSCs的自我更新^[94]。miR-664已被证明在皮肤鳞状细胞癌(squamous cell carcinoma, SCC)中含量升高, 可增强SCC细胞的迁移和侵袭能力, *ARMC8*(armadillo repeat containing 8)基因作为miR-664的直接作用靶点抑制Wnt/ β -catenin信号通路和cSCC中的上皮-间充质转化^[95]。

3.3 PI3K/AKT信号通路

PI3K/AKT通路是一条重要的细胞内信号转导



在细胞质内, miR-500a-3p负向调控SOCS3和SOCS4, miR-30和miR-410抑制SOCS3, miR-301a和miR-141-3p抑制SOCS5, 从而介导JAK/STAT3激活, 促进肿瘤发生发展; 在细胞核中, miR-30c-5p通过靶向调控USP22, 介导JAK/STAT3信号。与此相反, 在细胞外, miR-9和miR-515-5p通过抑制IL-6, 抑制JAK/STAT3途径激活而发挥抑癌作用。在细胞内胞质中, miR-17a-3p和miR-20a的上调可抑制JAK、STAT3和PI3K蛋白表达, 从而抑制JAK/STAT3途径激活。

In the cytoplasm, miR-500a-3p exerts a negative influence on SOCS3 and SOCS4, miR-30 and miR-410 mediate JAK/STAT3 activation by inhibiting SOCS3, as well as miR-301a and miR-141-3p mediate JAK/STAT3 activation by inhibiting SOCS5. MiR-30c-5p mediates the JAK/STAT3 signaling by targeting USP22 in the nucleus. Conversely, extracellular miR-9 and miR-515-5p have anti-cancer properties by inhibiting IL-6 and curbing the activation of the JAK/STAT3 pathway. Intracellular cytoplasm upregulation of miR-17a-3p and miR-20a inhibits JAK, STAT3, and PI3K proteins, thereby preventing JAK/STAT3 pathway activation.

图1 miRNA影响细胞增殖的机制

Fig.1 The mechanism of miRNA impacting cell proliferation

通路,调节细胞增殖、凋亡、代谢和血管生成等生物学过程,有研究表明,该信号通路的失控与不同类型人类恶性肿瘤的发生和发展有关^[96]。HASHEMI等^[97]研究发现,miRNA、lncRNA和circRNA等上游调控因子调节PI3K/AKT信号转导,刺激PI3K/AKT信号通路可促进前列腺癌细胞存活,减少细胞凋亡,PI3K/AKT信号通路可刺激EMT,促进前列腺癌细胞的转移,沉默PI3K/AKT信号会影响前列腺癌细胞的生长和转移。CHI等^[98]研究miR-194-5p在胰腺导管腺癌(pancreatic ductal adenocarcinoma, PDAC)进展中起关键作用的分子机制,发现miR-194-5p通过调控PVRL3/miR-194-5p/SOCS2轴激活PI3K/AKT信号通路,调控PDAC的增殖和转移。在PI3K/AKT信号通路中,miRNA主要通过信号转导参与基因的上游调控,进而影响肿瘤增殖通路,这与以上两种通路作用方式存在明显的区别,虽有较少文献讲解miRNA参与此条通路的相关知识,但miRNA在PI3K/AKT信号通路中发挥的作用具有深远的研究意义。

4 miRNA在临床医学中的应用

由于miRNA的稳定性,肿瘤进展过程中异常表达的基因和miRNA可作为肿瘤诊断和预后的生物标志物,在肿瘤诊断中可通过检测miRNA的甲基化水平来监控肿瘤变化。研究发现,在非小细胞肺癌及其组织学亚型中一些高甲基化miRNA存在甲基化水平的变化,在胰腺癌和鳞状细胞肺癌中,miR-124-1、miR-124-3、miR125B-1、miR129-2、miR137、miR1258和miR339的甲基化水平明显高于邻近组织学特征未变的肺组织样本,在鳞状细胞肺癌中,miR-124-2的甲基化水平也显著较高^[99]。

miRNA的比较重要的另外一种功能是在临床中可作为某一种或几种癌症的诊断标志物。HUANG等^[100]通过对全局miRNA表达进行分析,共鉴定出9个与胃肠癌预后有关的miRNA,其中,miR-194是唯一一个与胃癌、结直肠癌和肝癌的总生存期、疾病特异性生存期和无进展间期均显著相关的miRNA,可见miR-194是预测胃肠癌的最佳预后生物标志物。RUI等^[101]通过分析血清外切体中miRNA的比例和分布,发现miRNA是血清外切体中最重要的组成成分,3个来自血清外切体的miRNA(miR-122-5p、let-7d-5p和miR-4255p)对诊断肝癌具有重要价值,可成为肝癌诊断的生物标志

物。关于血清来源的miRNA let-7及其所调控的通路已成为耐药癌症的新治疗靶点,已经有较多文献报道。let-7家族调节包括KRAS基因在内的多种致癌基因,其还可作为头颈癌的预测性生物标志物,并已被证明在致癌作用中发挥关键作用^[102]。血清let-7-A1、miR-143和miR-145表达下调可能对埃及慢性丙型肝炎患者中肝细胞癌的发生发展有重要影响,可作为肝细胞癌诊断的生物标志物^[103]。let-7作为最早发现的miRNA,研究发现其和miR-21协同调控对肺癌的作用大于单一miRNA的作用。miR-21和let-7是肺癌中重要的差异表达miRNA,对于多种基因参与的肺癌来说,这两个miRNA的协同调控将为未来肺癌的治疗提供新的靶点和方向^[104]。WANG等^[105]研究表明血浆外切体miR-320d、miR-4479和miR-6763-5p可作为卵巢上皮性癌的诊断标志物,WROBLEWSKA等^[106]发现血清外切体源的hsa-miR-191-5p可作为一种新的葡萄膜黑色素瘤早期检测的生物标志物。ABO-ELELA等^[107]发现,miR-133a、miR-574-3p和miR-27a可作为早期诊断结直肠癌的较好的无创性标志物,具有较高的敏感性和特异性,有助于早期发现结直肠癌,且对于这种广泛传播的恶性肿瘤可通过手术进行干预,以改善结直肠癌患者的预后和提高患者的生存率。

除了在肿瘤诊断中可使用miRNA外,miRNA还显示出作为治疗靶点的潜力。在恶性胶质瘤治疗中,研究发现使用miRNA的3个主要考虑因素^[108]:(1)递送方法,尽管miRNA体积小,在血清中稳定,实现全身给药效率仍不佳。通过局部递送和基于纳米颗粒的递送,可以让miRNA跨越血脑屏障,到达肿瘤细胞,这种方法具有更高的稳定性和效率。(2)靶点的特异性,肿瘤特异性表面受体作为识别靶点,可确保miRNA传递到正确位置的特异性。(3)目标基因的特异性,miRNA靶向多个基因的能力是其治疗应用的限制因素,以miRNA为基础的药物主要障碍是以感兴趣的基因为靶点,以避免非特定靶点。所以需要更多的研究来提高基于miRNA的药物的特异性,以进行靶向治疗。

5 展望

在肿瘤发展早期检测miRNA可增加治疗成功的机会。circRNA-miRNA分子相互作用是癌症早期检测和预后中重要的研究领域,其不仅对提高患者

生存率至关重要, 而且对癌症预防也至关重要。根据miRNA的表达与肿瘤发生发展及治疗之间的关联, 可将miRNA确定为疾病诊断和预后的潜在生物标志物。miRNA已经被发现是一种新颖的、非常有前途的生物标志物, 也是新的治疗方法的有吸引力的工具和靶点。本文通过探讨miRNA参与调控肿瘤形成的方式以及对肿瘤细胞的影响, 阐明miRNA影响肿瘤发生的机制, 并对三条常见的JAK/STAT3信号通路、Wnt/ β -catenin信号通路和PI3K/AKT信号通路进行总结分析, 为miRNA后续研究提供帮助。

参考文献 (References)

- [1] ÇAKMAK H A, DEMIR M. MicroRNA and cardiovascular diseases [J]. *Balkan Med J*, 2020, 37(2): 60-71.
- [2] AMBROS V. The functions of animal microRNAs [J]. *Nature*, 2004, 431(7006): 350-5.
- [3] HAN X, GUO J, FAN Z. Interactions between m6A modification and miRNAs in malignant tumors [J]. *Cell Death Dis*, 2021, 12(6): 598.
- [4] ESQUELA-KERSCHER A, SLACK F J. Oncomirs-microRNAs with a role in cancer [J]. *Nat Rev Cancer*, 2006, 6(4): 259-69.
- [5] POZZO E, GIARRATANA N, SASSI G, et al. Upregulation of miR181a/miR212 improves myogenic commitment in murine fusion-negative rhabdomyosarcoma [J]. *Front Physiol*, 2021, 12: 701354.
- [6] BARTEL D P. MicroRNAs: target recognition and regulatory functions [J]. *Cell*, 2009, 136(2): 215-33.
- [7] DIECKMANN K P, RADTKE A, GECZI L, et al. Serum levels of microRNA-371a-3p (M371 test) as a new biomarker of testicular germ cell tumors: results of a prospective multicentric study [J]. *J Clin Oncol*, 2019, 37(16): 1412-23.
- [8] LIU B, SHYR Y, CAI J, et al. Interplay between miRNAs and host genes and their role in cancer [J]. *Brief Funct Genomics*, 2018, 18(4): 255-66.
- [9] JIA Y, ZHAO J, YANG J, et al. MiR-301 regulates the SIRT1/SOX2 pathway via CPEB1 in the breast cancer progression [J]. *Mol Ther Oncolytics*, 2021, 22: 13-26.
- [10] GROOT M, LEE H. Sorting mechanisms for microRNAs into extracellular vesicles and their associated diseases [J]. *Cells*, 2020, 9(4): 1044.
- [11] SHEN J, LIANG C, SU X, et al. Dysfunction and ceRNA network of the tumor suppressor miR-637 in cancer development and prognosis [J]. *Biomark Res*, 2022, 10(1): 72.
- [12] 石嘉琛, 阿依木古丽·阿不都热依木, 王家敏, 等. 竞争内源性RNA在肿瘤中的研究进展[J]. *生命科学*(SHI J C, AYIMUGULI A, WANG J M, et al. *Advances in the study of competing endogenous RNA in tumors* [J]. *Chinese Bulletin of Life Sciences*), 2020, 32(9): 929-36.
- [13] WANG N, CAO S, WANG X, et al. lncRNA MALAT1/miR-26a/26b/ST8SIA4 axis mediates cell invasion and migration in breast cancer cell lines [J]. *Oncol Rep*, 2021, 46(2): 181.
- [14] GOLLA U, SESHAM K, DALLAVALASA S, et al. ABHD11-AS1: an emerging long non-coding RNA (lncRNA) with clinical significance in human malignancies [J]. *Noncoding RNA*, 2022, 8(2): 21.
- [15] MA G, LI G, FAN W, et al. Circ-0005105 activates COL11A1 by targeting miR-20a-3p to promote pancreatic ductal adenocarcinoma progression [J]. *Cell Death Dis*, 2021, 12(7): 656.
- [16] ZHAO J, LI X, WANG M, et al. Circular RNA ABCB10 contributes to laryngeal squamous cell carcinoma (LSCC) progression by modulating the miR-588/CXCR4 axis [J]. *Aging*, 2021, 13(10): 14078-87.
- [17] LI M, WANG J, MA H, et al. Extracellular vesicles long non-coding RNA AGAP2-AS1 contributes to cervical cancer cell proliferation through regulating the miR-3064-5p/SIRT1 axis [J]. *Front Oncol*, 2021, 11: 684477.
- [18] LI M, LUO M, LIU P, et al. Circ_0001402 knockdown suppresses the chemoresistance and development of DDP-resistant cutaneous squamous cell carcinoma cells by functioning as a ceRNA for miR-625-5p [J]. *Exp Dermatol*, 2023, 32(4): 529-41.
- [19] LI F, ZHOU C, LI S, et al. Bioinformatic analysis of differentially expressed profiles of lncRNAs and miRNAs with their related ceRNA network in endometrial cancer [J]. *Medicine*, 2023, 102(3): e32573.
- [20] ZHOU J, SHI K, HUANG W, et al. lncRNA RPPH1 acts as a molecular sponge for miR-122 to regulate Wnt1/beta-catenin signaling in hepatocellular carcinoma [J]. *Int J Med Sci*, 2023, 20(1): 23-34.
- [21] TAHERI M, SAFARZADEH A, HUSSEN B M, et al. lncRNA/miRNA/mRNA network introduces novel biomarkers in prostate cancer [J]. *Cells*, 2022, 11(23): 3776.
- [22] WANG W, HAO L P, SONG H, et al. The potential roles of exosomal non-coding RNAs in hepatocellular carcinoma [J]. *Front Oncol*, 2022, 12: 790916.
- [23] JIAO Y, ZHANG L, LI J, et al. Exosomal miR-122-5p inhibits tumorigenicity of gastric cancer by downregulating GIT1 [J]. *Int J Biol Markers*, 2021, 36(1): 36-46.
- [24] LI W, DONG Y, WANG K, et al. Plasma exosomal miR-125a-5p and miR-141-5p as non-invasive biomarkers for prostate cancer [J]. *Neoplasma*, 2020, 67(6): 1314-8.
- [25] WANG Z, XIA F, FENG T, et al. OTUD6B-AS1 inhibits viability, migration, and invasion of thyroid carcinoma by targeting miR-183-5p and miR-21 [J]. *Front Endocrinol*, 2020, 11: 136.
- [26] WANG J, YUE B L, HUANG Y Z, et al. Exosomal RNAs: novel potential biomarkers for diseases-a review [J]. *Int J Mol Sci*, 2022, 23(5): 2461.
- [27] FANG T, LÜ H, LÜ G, et al. Tumor-derived exosomal miR-1247-3p induces cancer-associated fibroblast activation to foster lung metastasis of liver cancer [J]. *Nat Commun*, 2018, 9(1): 191.
- [28] JIA Y, DING X, ZHOU L, et al. Mesenchymal stem cells-derived exosomal microRNA-139-5p restrains tumorigenesis in bladder cancer by targeting PRC1 [J]. *Oncogene*, 2021, 40(2): 246-61.
- [29] CHEN L, CAO P, HUANG C, et al. Serum exosomal miR-7977 as a novel biomarker for lung adenocarcinoma [J]. *J Cell Biochem*, 2020, 121(5/6): 3382-91.
- [30] ZHOU C, WEI W F, MA J, et al. Cancer-secreted exosomal miR-1468-5p promotes tumor immune escape via the immunosuppressive reprogramming of lymphatic vessels [J]. *Mol Ther*, 2021, 29(4): 1512-28.

- [31] HASHEMI M, MIRDAMADI M S A, TALEBI Y, et al. Pre-clinical and clinical importance of miR-21 in human cancers: tumorigenesis, therapy response, delivery approaches and targeting agents [J]. *Pharmacol Res*, 2023, 187: 106568.
- [32] CARVALHO T M, BRASIL G O, JUCOSKI T S, et al. MicroRNAs miR-142-5p, miR-150-5p, miR-320a-3p, and miR-4433b-5p in serum and tissue: potential biomarkers in sporadic breast cancer [J]. *Front Genet*, 2022, 13: 865472.
- [33] LIU M, MO F, SONG X, et al. Exosomal hsa-miR-21-5p is a biomarker for breast cancer diagnosis [J]. *PeerJ*, 2021, 9: e12147.
- [34] WANG S W, SONG X G, WANG K Y, et al. Plasma exosomal miR-320d, miR-4479, and miR-6763-5p as diagnostic biomarkers in epithelial ovarian cancer [J]. *Front Oncol*, 2022, 12: 986343.
- [35] ZHU X L, SHEN H L, YIN X M, et al. Macrophages derived exosomes deliver miR-223 to epithelial ovarian cancer cells to elicit a chemoresistant phenotype [J]. *J Exp Clin Cancer Res*, 2019, 38(1): 81.
- [36] DIETERICH L C, BIKFALVI A. The tumor organismal environment: role in tumor development and cancer immunotherapy [J]. *Semin Cancer Biol*, 2020, 65: 197-206.
- [37] GOODALL G J, WICKRAMASINGHE V O. RNA in cancer [J]. *Nat Rev Cancer*, 2021, 21(1): 22-36.
- [38] CHOI J Y, SEOK H J, KIM R K, et al. MiR-519d-3p suppresses tumorigenicity and metastasis by inhibiting Bcl-w and HIF-1 α in NSCLC [J]. *Mol Ther Oncolytics*, 2021, 22: 368-79.
- [39] IKEDA S, KITADATE A, ABE F, et al. Hypoxia-inducible microRNA-210 regulates the DIMT1-IRF4 oncogenic axis in multiple myeloma [J]. *Cancer Sci*, 2017, 108(4): 641-52.
- [40] ROSENBERG T, THOMASSEN M, JENSEN S, et al. Acute hypoxia induces upregulation of microRNA-210 expression in glioblastoma spheroids [J]. *Cns Oncol*, 2015, 4(1): 25-35.
- [41] LI Y, LI M, SHATS I, et al. Glypican 6 is a putative biomarker for metastatic progression of cutaneous melanoma [J]. *PLoS One*, 2019, 14(6): e0218067.
- [42] CARTIER F, INDERSIE E, LESJEAN S, et al. New tumor suppressor microRNAs target glypican-3 in human liver cancer [J]. *Oncotarget*, 2017, 8(25): 41211-26.
- [43] FENG C, LI J, RUAN J, et al. MicroRNA-125a inhibits cell growth by targeting glypican-4 [J]. *Glycoconj J*, 2012, 29(7): 503-11.
- [44] HONG X, ZHANG Z, PAN L, et al. MicroRNA-301b promotes the proliferation and invasion of glioma cells through enhancing activation of Wnt/beta-catenin signaling via targeting Glypican-5 [J]. *Eur J Pharmacol*, 2019, 854: 39-47.
- [45] MA Y S, LIU J B, LIN L, et al. Exosomal microRNA-15a from mesenchymal stem cells impedes hepatocellular carcinoma progression via downregulation of SALL4 [J]. *Cell Death Discov*, 2021, 7(1): 224.
- [46] LI Y, FEI H, LIN Q, et al. ZEB2 facilitates peritoneal metastasis by regulating the invasiveness and tumorigenesis of cancer stem-like cells in high-grade serous ovarian cancers [J]. *Oncogene*, 2021, 40(32): 5131-41.
- [47] JIANG H, LIU Y, HU K, et al. MiRNA-339 targets and regulates ZNF689 to inhibit the proliferation and invasion of gastric cancer cells [J]. *Transl Cancer Res*, 2021, 10(7): 3516-26.
- [48] SELVAM M, BANDI V, PONNE S, et al. MicroRNA-150 targets major epigenetic repressors and inhibits cell proliferation [J]. *Exp Cell Res*, 2022, 415(1): 113110.
- [49] ZHOU M, GAO Y, WANG M, et al. MiR-146b-3p regulates proliferation of pancreatic cancer cells with stem cell-like properties by targeting MAP3K10 [J]. *J Cancer*, 2021, 12(12): 3726-40.
- [50] ZHANG X, WANG L, LI H, et al. Crosstalk between noncoding RNAs and ferroptosis: new dawn for overcoming cancer progression [J]. *Cell Death Dis*, 2020, 11(7): 580.
- [51] HUANG T, WAN X, ALVAREZ A A, et al. MiR93 (microRNA-93) regulates tumorigenicity and therapy response of glioblastoma by targeting autophagy [J]. *Autophagy*, 2019, 15(6): 1100-11.
- [52] SONG Y, BAI L, YAN F, et al. Inhibition of EMMPRIN by microRNA-124 suppresses the growth, invasion and tumorigenicity of gliomas [J]. *Exp Ther Med*, 2021, 22(3): 930.
- [53] CUI Y, WANG Q, LIN J, et al. MiRNA-193a-3p regulates the AKT2 pathway to inhibit the growth and promote the apoptosis of glioma cells by targeting ALKBH5 [J]. *Front Oncol*, 2021, 11: 600451.
- [54] TANG J, PAN H, WANG W, et al. MiR-495-3p and miR-143-3p co-target CDK1 to inhibit the development of cervical cancer [J]. *Clin Transl Oncol*, 2021, 23(11): 2323-34.
- [55] YARI M, SOLTANI B, GHAEMI Z, et al. EVADR ceRNA transcript variants upregulate WNT and PI3K signaling pathways in SW480 and HCT116 cells by sponging miR-7 and miR-29b [J]. *Biol Chem*, 2023, 404(1): 71-83.
- [56] GHEIDARI F, AREFIAN E, SAADATPOUR F, et al. The miR-429 suppresses proliferation and migration in glioblastoma cells and induces cell-cycle arrest and apoptosis via modulating several target genes of ERBB signaling pathway [J]. *Mol Biol Rep*, 2022, 49(12): 11855-66.
- [57] YANG Z, ZHANG M, ZHANG J, et al. MiR-302a-3p promotes radiotherapy sensitivity of hepatocellular carcinoma by regulating cell cycle via MCL1 [J]. *Comput Math Methods Med*, 2022, 2022: 1450098.
- [58] HASSAN M, ELZALLAT M, ABOUSHOUSA T, et al. MicroRNA-122 mimic/microRNA-221 inhibitor combination as a novel therapeutic tool against hepatocellular carcinoma [J]. *Non-coding RNA Res*, 2023, 8(1): 126-34.
- [59] ZHOU C, ZHU S, LI H. MiR-195-5p targets CDK1 to regulate new DNA synthesis and inhibit the proliferation of hepatocellular carcinoma cells [J]. *Appl Biochem Biotechnol*, 2023, 195(5): 3477-90.
- [60] LIU F, ZHANG G, LÜ S, et al. MiRNA-301b-3p accelerates migration and invasion of high-grade ovarian serous tumor via targeting CPEB3/EGFR axis [J]. *J Cell Biochem*, 2019, 120(8): 12618-27.
- [61] ZHENG S, JIANG F, GE D, et al. LncRNA SNHG3/miRNA-151a-3p/RAB22A axis regulates invasion and migration of osteosarcoma [J]. *Biomed Pharmacother*, 2019, 112: 108695.
- [62] ZHANG B, WANG D, WANG Y, et al. MiRNA-338-3p inhibits the migration, invasion and proliferation of human lung adenocarcinoma cells by targeting MAP3K2 [J]. *Aging*, 2022, 14(15): 6094-110.
- [63] LIU C, LI Y, ZHANG L, et al. MiRNA-196-5p promotes proliferation and migration in cholangiocarcinoma via HAND1/Wnt/ β -Catenin signaling pathway [J]. *J Oncol*, 2022, 2022: 4599676.

- [64] SUBRAMANIAM S, JEET V, CLEMENTS J A, et al. Emergence of microRNAs as key players in cancer cell metabolism [J]. *Clin Chem*, 2019, 65(9): 1090-101.
- [65] QUIRICO L, ORSO F, CUCINELLI S, et al. MiRNA-guided reprogramming of glucose and glutamine metabolism and its impact on cell adhesion/migration during solid tumor progression [J]. *Cell Mol Life Sci*, 2022, 79(4): 216.
- [66] SUN Y, LIU W, ZHAO Q, et al. Down-regulating the expression of miRNA-21 Inhibits the glucose metabolism of A549/DDP cells and promotes cell death through the PI3K/AKT/mTOR/HIF-1 α pathway [J]. *Front Oncol*, 2021, 11: 653596.
- [67] LO Y L, WANG C S, CHEN Y C, et al. Mitochondrion-directed nanoparticles loaded with a natural compound and a microRNA for promoting cancer cell death via the modulation of tumor metabolism and mitochondrial dynamics [J]. *Pharmaceutics*, 2020, 12(8): 756.
- [68] LIU X, ZHAO D. CKS1B promotes the progression of hepatocellular carcinoma by activating JAK/STAT3 signal pathway [J]. *Anim Cells Syst*, 2021, 25(4): 227-34.
- [69] ISMAIL A, EL-MAHDY H A, ABULSOUD A I, et al. Beneficial and detrimental aspects of miRNAs as chief players in breast cancer: a comprehensive review [J]. *Int J Biol Macromol*, 2023, 224: 1541-65.
- [70] NASROLAHI A, AZIZDOOST S, RADOSZKIEWICZ K, et al. Signaling pathways governing glioma cancer stem cells behavior [J]. *Cell Signal*, 2023, 101: 110493.
- [71] ELREBEHY M A, AL-SAEED S, GAMAL S, et al. MiRNAs as cornerstones in colorectal cancer pathogenesis and resistance to therapy: a spotlight on signaling pathways interplay-a review [J]. *Int J Biol Macromol*, 2022, 214: 583-600.
- [72] XIE Z, ZHONG C, SHEN J, et al. LINC00963: a potential cancer diagnostic and therapeutic target [J]. *Biomed Pharmacother*, 2022, 150: 113019.
- [73] ZOU M, SHEN J, WU Y, et al. Dysregulation of miR-411 in cancer: causative factor for pathogenesis, diagnosis and prognosis [J]. *Biomed Pharmacother*, 2022, 149: 112896.
- [74] CHONG Z, YEAP S, HO W, et al. Unveiling the tumour-regulatory roles of miR-1275 in cancer [J]. *Pathol Res Pract*, 2022, 230: 153745.
- [75] ZHANG L, LI J, WANG Q, et al. The relationship between microRNAs and the STAT3-related signaling pathway in cancer [J]. *Tumour Biol*, 2017, 39(7): 1010428317719869.
- [76] ZHU D, CHEN C, LIU X, et al. Osteosarcoma cell proliferation suppression via SHP-2-mediated inactivation of the JAK/STAT3 pathway by tubocapsenolide A [J]. *J Adv Res*, 2021, 34: 79-91.
- [77] SUN L, DING S, LUO Q, et al. Taxus wallichiana var. chinensis (Pilg.) floric aqueous extract suppresses the proliferation and metastasis in lung carcinoma via JAK/STAT3 signaling pathway [J]. *Front Pharmacol*, 2021, 12: 736442.
- [78] WANG H, FU Y. NR1D1 suppressed the growth of ovarian cancer by abrogating the JAK/STAT3 signaling pathway [J]. *BMC Cancer*, 2021, 21(1): 871.
- [79] DING H, YU X, YAN Z. Ailanthone suppresses the activity of human colorectal cancer cells through the STAT3 signaling pathway [J]. *Int J Mol Med*, 2022, 49(2): 21.
- [80] XIN Y, SHANG X, SUN X, et al. Trefoil factor 3 inhibits thyroid cancer cell progression related to IL-6/JAK/STAT3 signaling pathway [J]. *Evid Based Complement Alternat Med*, 2021, 2021: 2130229.
- [81] HU H, ZHANG Q, CHEN W, et al. MicroRNA-301a promotes pancreatic cancer invasion and metastasis through the JAK/STAT3 signaling pathway by targeting SOCS5 [J]. *Carcinogenesis*, 2020, 41(4): 502-14.
- [82] MASOUMI-DEHGHANI S, BABASHAH S, SADEGHIZADEH M. MicroRNA-141-3p-containing small extracellular vesicles derived from epithelial ovarian cancer cells promote endothelial cell angiogenesis through activating the JAK/STAT3 and NF- κ B signaling pathways [J]. *J Cell Commun Signal*, 2020, 14(2): 233-44.
- [83] CHE S, SUN T, WANG J, et al. MiR-30 overexpression promotes glioma stem cells by regulating Jak/STAT3 signaling pathway [J]. *Tumour Biol*, 2015, 36(9): 6805-11.
- [84] XU G, CAI J, WANG L, et al. MicroRNA-30e-5p suppresses non-small cell lung cancer tumorigenesis by regulating USP22-mediated Sirt1/JAK/STAT3 signaling [J]. *Exp Cell Res*, 2018, 362(2): 268-78.
- [85] LI M, ZHENG R, YUAN F. MiR-410 affects the proliferation and apoptosis of lung cancer A549 cells through regulation of SOCS3/JAK-STAT signaling pathway [J]. *Eur Rev Med Pharmacol Sci*, 2020, 24(22): 11462.
- [86] JIANG C, LONG J, LIU B, et al. MiR-500a-3p promotes cancer stem cells properties via STAT3 pathway in human hepatocellular carcinoma [J]. *J Exp Clin Cancer Res*, 2017, 36(1): 99.
- [87] DELEN E, DOGANLAR O, DOGANLAR Z B, et al. Inhibition of the invasion of human glioblastoma U87 cell line by ruxolitinib: a molecular player of miR-17 and miR-20a regulating JAK/STAT pathway [J]. *Turk Neurosurg*, 2020, 30(2): 182-9.
- [88] ZHANG J, JIA J, ZHAO L, et al. Down-regulation of microRNA-9 leads to activation of IL-6/Jak/STAT3 pathway through directly targeting IL-6 in HeLa cell [J]. *Mol Carcinog*, 2016, 55(5): 732-42.
- [89] NI J S, ZHENG H, OU Y L, et al. MiR-515-5p suppresses HCC migration and invasion via targeting IL6/JAK/STAT3 pathway [J]. *Surg Oncol*, 2020, 34: 113-20.
- [90] CHEN Q, WANG H, LI Z, et al. Circular RNA ACTN4 promotes intrahepatic cholangiocarcinoma progression by recruiting YBX1 to initiate FZD7 transcription [J]. *J Hepatol*, 2022, 76(1): 135-47.
- [91] MARETTO S, CORDENONSI M, DUPONT S, et al. Mapping Wnt/beta-catenin signaling during mouse development and in colorectal tumors [J]. *Proc Natl Acad Sci USA*, 2003, 100(6): 3299-304.
- [92] GIAKOUSTIDIS A, GIAKOUSTIDIS D, MUDAN S, et al. Molecular signalling in hepatocellular carcinoma: role of and crosstalk among WNT/ β -catenin, Sonic Hedgehog, Notch and Dickkopf-1 [J]. *Can J Gastroenterol Hepatol*, 2015, 29(4): 209-17.
- [93] GHAFOURI-FARD S, HAJIESMAEILI M, SHOOREI H, et al. The impact of lncRNAs and miRNAs in regulation of function of cancer stem cells and progression of cancer [J]. *Front Cell Dev Biol*, 2021, 9: 696820.
- [94] CHEN Z Q, YUAN T, JIANG H, et al. MicroRNA-8063 targets heterogeneous nuclear ribonucleoprotein AB to inhibit the self-renewal of colorectal cancer stem cells via the Wnt/ β -catenin pathway [J]. *Oncol Rep*, 2021, 46(4): 219.

- [95] LI X, ZHANG C, YUAN Y, et al. Downregulation of ARMC8 promotes tumorigenesis through activating Wnt/beta-catenin pathway and EMT in cutaneous squamous cell carcinomas [J]. *J Dermatol Sci*, 2021, 102(3): 184-92.
- [96] MARINI F, GIUSTI F, PALMINI G, et al. Genetics and epigenetics of parathyroid carcinoma [J]. *Front Endocrinol*, 2022, 13: 834362.
- [97] HASHEMI M, TAHERIAZAM A, DANEII P, et al. Targeting PI3K/Akt signaling in prostate cancer therapy [J]. *J Cell Commun Signal*, 2022, doi: 10.1007/s12079-022-00702-1.
- [98] CHI B, ZHENG Y, XIE F, et al. Increased expression of miR-194-5p through the circPVRL3/miR-194-5p/SOCS2 axis promotes proliferation and metastasis in pancreatic ductal adenocarcinoma by activating the PI3K/AKT signaling pathway [J]. *Cancer Cell Int*, 2022, 22(1): 415.
- [99] GUBENKO M S, LOGINOV V I, BURDENNYI A M, et al. Changes in the level of methylation of a group of microRNA genes as a factor in the development and progression of non-small cell lung cancer [J]. *Bull Exp Biol Med*, 2022, 174(2): 254-8.
- [100] HUANG P, XIA L, GUO Q, et al. Genome-wide association studies identify miRNA-194 as a prognostic biomarker for gastrointestinal cancer by targeting ATP6V1F, PPP1R14B, BTF3L4 and SLC7A5 [J]. *Front Oncol*, 2022, 12: 1025594.
- [101] RUI T, ZHANG X, GUO J, et al. Serum-exosome-derived miRNAs serve as promising biomarkers for HCC diagnosis [J]. *Cancers*, 2022, 15(1): 205.
- [102] ULUSAN M, SEN S, YILMAZER R, et al. The let-7 microRNA binding site variant in KRAS as a predictive biomarker for head and neck cancer patients with lymph node metastasis [J]. *Pathol Res Pract*, 2022, 239: 154147.
- [103] ALY D M, GOHAR N A, ABD EL-HADY A A, et al. Serum microRNA let-7a-1/let-7d/let-7f and miRNA 143/145 gene expression profiles as potential biomarkers in HCV induced hepatocellular carcinoma [J]. *Asian Pac J Cancer Prev*, 2020, 21(2): 555-62.
- [104] BAI J, SHI Z, WANG S, et al. MiR-21 and let-7 cooperation in the regulation of lung cancer [J]. *Front Oncol*, 2022, 12: 950043.
- [105] WANG S, SONG X, WANG K, et al. Plasma exosomal miR-320d, miR-4479, and miR-6763-5p as diagnostic biomarkers in epithelial ovarian cancer [J]. *Front Oncol*, 2022, 12: 986343.
- [106] WROBLEWSKA J P, LACH M S, RUCINSKI M, et al. MiRNAs from serum-derived extracellular vesicles as biomarkers for uveal melanoma progression [J]. *Front Cell Dev Biol*, 2022, 10: 1008901.
- [107] ABO-ELELA D A, SALEM A M, SWELLAM M, et al. Potential diagnostic role of circulating MiRNAs in colorectal cancer [J]. *Int J Immunopathol Pharmacol*, 2023, 37: 3946320221144565.
- [108] SATI I, PARHAR I. MicroRNAs regulate cell cycle and cell death pathways in glioblastoma [J]. *Int J Mol Sci*, 2021, 22(24): 13550.