泛素化修饰在抗病毒天然免疫反应中的作用

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摘要 蛋白质翻译后修饰几乎调控细胞所有的生命活动,有大量研究报道了其中的泛素化 在病毒感染过程中的作用。受病毒感染时,宿主可利用泛素化修饰起始抗病毒天然免疫反应,从而 抵抗病毒入侵。相应地,病毒也可以利用泛素化修饰逃逸细胞的免疫反应。该文从宿主与病毒两 个角度综述了蛋白质的泛素化修饰在抗病毒天然免疫中的作用及其调控机制,为抗病毒的治疗提 供一些新的策略。

关键词 泛素化修饰; 抗病毒天然免疫反应; 免疫逃逸

The Role of Ubiquitination in Antiviral Innate Immune Response

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Abstract The post translational modification of proteins regulates almost all the life activities of cells. A large number of studies have reported the role of ubiquitination in the process of viral infection. When infected with the virus, host cells can initiate an antiviral innate immune response to restrict viral infection, and correspondingly, viruses can escape the cellular immune response, through ubiquitination pathways. This paper reviews the role and regulatory mechanism of protein ubiquitination from both host and virus perspectives, providing some new strategies for future antiviral treatments.

Keywords ubiquitination; antiviral innate immune response; immune escape

泛素(ubiquitin, UB)是一种普遍存在于真核生物体内的含有76个氨基酸残基的小分子质量蛋白。 UB是一种分子标记蛋白,经由泛素激活酶E1、泛素 偶联酶E2和泛素连接酶E3的级联反应,UB分子C-端 甘氨酸残基的羧基共价连接到底物蛋白质的赖氨酸 残基侧链的ε-氨基上,形成异肽键^[1]。这一过程被称 为蛋白质的泛素化修饰。通过泛素化修饰,蛋白质 的构象、活性、稳定性等多方面都会发生变化,进 而调控多种生物学过程。受病毒感染后,宿主会激 活抗病毒天然免疫反应以抵御病毒对自身的侵害。 而同时,病毒进化出相应的机制以完成免疫逃逸。 本文综述了蛋白质泛素化修饰在抗病毒天然免疫反 应和病毒免疫逃逸中的作用,进而解析在病毒感染 过程中两者的蛋白质网络产生复杂变化的分子机 制。

1 泛素化修饰

根据连接方式不同,可将泛素化修饰分为单泛 素化和多泛素化。单泛素化修饰是指单个泛素分子 与靶蛋白的赖氨酸侧链相连,多泛素化修饰则是靶 蛋白的多个赖氨酸残基与泛素分子结合。泛素分 子的七个赖氨酸残基(K6、K11、K27、K29、K33、 K48和K63)和一个甲硫氨酸残基(M1)均可作为泛素 化位点,已经共价修饰到靶蛋白上的泛素分子还可

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继续发生泛素化修饰,即形成泛素链。目前发现的 细胞中存量最多的2种同型泛素链是K48泛素链和 K63泛素链,其中,K48 泛素链是最主要的泛素修饰 类型,在所有泛素修饰中占到50%以上。其他泛素 链则属于非典型泛素链。泛素链拓扑结构的复杂性 造成了其功能多样性,包括参与囊泡运输途径、调 节组蛋白修饰和病毒出芽等,这对于细胞周期调节、 DNA修复、细胞生长和免疫功能以及激素介导的信 号转导意义重大。

泛素化修饰作为一种可逆的翻译后修饰,能够被去泛素化酶(deubiquitinase, DUB)去除其与底物之间的共价键以及泛素分子之间的共价键。同时DUB能够介导蛋白的降解,调节靶蛋白性质及相关信号通路,进而调控底物的生命进程^[2]。

2 蛋白质泛素化在病毒感染中的作用

当病毒感染细胞后,对于宿主而言,病毒作为 外来物,会刺激宿主启动自身的免疫系统,激活相关 基因及蛋白质的表达,以限制或约束病毒的感染,甚 至清除病毒。相应地,对于病毒而言,则需要尽快地 完成自身基因组的复制、装配及增殖,以逃避甚至 抵御宿主免疫系统的约束。本文将从宿主和病毒两 个不同角度分别阐述蛋白质泛素化修饰在病毒感染 中发挥的作用。

2.1 泛素化对宿主的影响

受病毒感染后,宿主细胞会在第一时间启动自 身的天然免疫系统。通过模式识别受体(pattern recognition receptors, PRRs)识别入侵病毒病原相关分子 模式(pathogen-associated molecular patterns, PAMPs), 起始系列抗病毒级联反应,激活下游相关分子如核 因子 κ 增强子结合蛋白(nuclear factor kappa enhancer binding protein, *NF-* κ *B*)和干扰素调节因子(interferon regulatory factor, *IRF*)的转录,诱导I型干扰素(interferon, IFN-I)和相关细胞因子的表达,从而招募先天 免疫细胞或激活程序性细胞死亡(programmed cell death, PCD),最终建立起细胞抗病毒状态。

宿主细胞的模式识别受体主要有五类: Toll样受体(Toll-like receptors, TLRs)、维甲酸诱导基因I受体(RIG-I like receptors, RLRs)、NOD样受体(NOD-like receptors, NLRs)、黑色素瘤缺乏因子2样受体(AIM2-like receptors, ALRs)以及胞质DNA受体^[3](图1)。

2.1.1 泛素化修饰调控TLRs信号通路 TLRs属

于I型跨膜糖蛋白,具有较高的保守性,由胞外区富 含亮氨酸重复序列、跨膜区含单个α螺旋、胞内区 含Toll-白细胞介素-1受体[Toll/interleukin-1(IL-1) receptor, TIR]信号结构域组成^[4]。到目前为止,已 发现13种TLR家族成员,人类细胞中共发现10种 (TLR1~TLR10) TLRs。其中,TLR3可识别dsRNA, TLR7和TLR8可识别ssRNA,TLR9则可识别病毒未 甲基化的DNA,TLR2和TLR4可识别病毒包膜糖蛋 白^[3]。

根据接头蛋白的不同, TLRs信号通路可分为含 TIR结构域的诱导IFN-β的接头(TIR domain-containing adaptor-inducing IFN-β, TRIF)蛋白依赖的TLRs 信号通路和髓样分化初级反应蛋白88(myeloid differentiation protein antigen 88, MyD88)依赖的TLRs 信号通路。其中, TLR3依赖TRIF接头蛋白作用调 控IFN-I表达, 其他TLRs依赖MyD88街头蛋白调控 NF-κB通路的表达。TLR4则依赖两种途径^[5]。

一方面, MyD88招募下游的白细胞介素-1受体 相关激酶(IL-1R-associated serine/threonine kinases, IRAKs), 后者传递信号到泛素连接酶6(TNF receptor-associated factor 6, TRAF6), 随后募集并活化转 化生长因子-β激活激酶1(TGFβ-activated kinase 1, TAK1)和IKKα/β/γ(也被称作NEMO), 以促进NF-кB 介导的炎症因子的表达。该通路受多种泛素连接 酶调控, 如UBL4A和TRIM26可分别催化TRAF6和 TAB1发生泛素化以增强先天免疫^[6-7]。有趣的是, 宿 主细胞可以通过自身合成的泛素连接酶或DUB负调 控TLRs信号通路, 从而防止发生过度的免疫反应。 如Smurfp1/2、Nrdp1和CYLD可催化MyD88发生泛 素化修饰或去除泛素链进行负调控^[6-8]; A20、IRAK-M、ST2和SIGIRR等同样可以调节TLRs信号的持续 时间和/或强度^[9-12]。

另一方面, TRIF通过招募 TRAF3, 激活 TANK 结合激酶 1(TANK-binding kinase 1, TBK1)和核因 子 κ B激酶抑制剂 ϵ (inhibitor- κ b kinase ϵ , IKK ϵ), 促进 IRF3诱导的IFN-I产生。该通路受多种酶调控,包括 cIAP1/2、Peli1、Triad3A、Mint3、Nedd4l、USP1 都能够正向调节TRIF介导的抗病毒免疫应答^[13-21]。 类似地,Triad3A、USP19、WWP2和TRIM32可降 解TRIF防止过度免疫的发生^[22-25]; RNF99则可促进 TAB2发生K48泛素化修饰进入蛋白酶体被降解,抑 制TAK-TABs复合体形成^[26]。不仅如此,有的泛素连



Fig.1 Ubiquitination regulates the host's natural immune response signaling pathways

接酶还可针对IFN-I产生通路外的蛋白进行调控,例如RNF182通过K48泛素化促进胞质p65的降解,进 而抑制炎症反应^[27]。由此可见,宿主细胞自身对天 然抗病毒免疫翻译的正调控和负调控对于维持机体 稳态至关重要。

2.1.2 泛素化修饰调控RLRs信号通路 RLRs家族 成员包括视黄酸(维甲酸)诱导基因蛋白-I(retinoic acid-inducible gene-I, RIG-I,也被称为DDX58)、黑色素 瘤分化相关基因5(melanoma differentiation associated factor 5, MDA5)和遗传与生理学实验室蛋白2(laboratory of genetics and physiology 2, LGP2),主要通过识 别胞质中的病毒RNA启动抗病毒天然免疫反应^[28]。 RIG-I和MDA5具有相似的序列,均由两个N-端的半 胱天冬酶招募结构域(caspase-recruitment domains, CARDs)、一个具有RNA结合活性的DExD/H-box解 旋酶结构域和一个具有RNA识别功能的C-端结构域 (C-terminal domain, CTD,也被称为调节或抑制结构 域)组成。相比之下, LGP2缺乏N-端CARDs。

在胞质中, RIG-I和 MDA5可通过 CTD 与病毒 RNA结合,改变自身构象以结合线粒体抗病毒信号 蛋白(mitochondrial antiviral sig naling protein, MAVS, 也称 VISA/Cardif/IPS-1),后者活化 TBK1/IKKi,最 终促进I型干扰素的产生,调控天然免疫反应信号 通路。当感染水泡性口炎病毒(vesicular stomatitis virus, VSV)时, Riplet(也称RNF135)催化RIG-I的C末 端发生K63泛素化,介导IFN-β启动子激活,最终参 与人类抗RNA病毒感染的先天免疫^[29]。除C末端外, RIG-I氨基端CARD结构域也可发生泛素化修饰,如 TRIM25、TRIM4与MEX3C可介导CARD结构域多 个位点发生K63多聚泛素化修饰,促进其对MAVS的 招募与结合,正向调控干扰素信号通路^[30-32]。而负 调控方面通过去泛素化酶介导,如OTUD3可去除由 Riplet催化的RIG-I的K63泛素链^[33]。另外,有研究发 现,由血红素氧化IRP2泛素连接酶1L(heme-oxidized IRP2 ubiquitin ligase 1L, HOIL-1L)和HOIL1接头蛋 白(HOIL-1-interacting protein, HOIP)构成的复合体 LUBAC(linear ubiquitin chain assembly complex), 可 以对RIG-I进行线性泛素化修饰并导致其降解;同时, LUBAC可以与TRIM25竞争性结合RIG-I,进而抑制 RLRs所介导的信号通路^[34-35]。

当机体感染脑心肌炎病毒(encephalomyocarditis virus, EMCV)时, 识别ssRNA的MDA5被TRIM65 催化发生K63泛素化, TRIM65缺失会减弱由EMCV 诱导的INF-I表达^[36]。同样, 为防止过度免疫, 泛素 连接酶可催化底物发生K48泛素链修饰使其进入蛋白酶体途径被降解,如RNF125、Parkin、PSMA7、TRIM44 L、TRIM13和TRIM40可通过泛素化修饰MDA5负调控RLRs通路,抑制病毒介导的天然免疫反应^[37-42]。

另外, MAVS复合物的泛素化调控也很有趣, 一 方面, TRIM31可以对MAVS多位点进行K63泛素化 修饰, 促进MAVS朊蛋白样多聚体的形成, 激活I型干 扰素的分泌, 抑制病毒的复制^[43]。另一方面, YOD1 可通过催化MAVS的K63链去泛素化进而抑制其聚 集和激活, 反向调控RLRs通路^[44]。也有文章报道 支架蛋白FAF1与TRIM31竞争MAVS来拮抗MAVS 的多泛素化和聚集。病毒感染后, FAF1的Ser556位 点发生磷酸化促进其被溶酶体降解, 从而缓解FAF1 对MAVS的抑制^[45]。此外, RNF5可通过催化激活的 IRF3发生K48泛素化, 减少I型干扰素的产生, 抑制天 然免疫反应^[46]。

2.1.3 泛素化修饰调控NLRs信号通路 NLRs具有 一个N-端效应结构域、一个中心核苷酸结合和寡聚 化结构域(oligomerization domain, NOD)、一个C-端 富亮氨酸重复序列(leucine-rich repeat, LRR)。根据 N-端效应结构域不同,动物NLRs可以分为NLRA、 NLRB/NAIP、NLRC和NLRP等亚科,四者分别具 有一个酸性反转录激活结构域(acidic transactivation, AD)、三个串联杆状病毒调亡抑制剂(inhibitor of apoptosis, IAP)重复序列(baculovirus inhibitor of apoptosis repeats, BIRs)、一个半胱天冬酶激活和 招募结构域(caspase activation and recruitment domain, CARD)和一个PYRIN结构域(PYD)^[47]。NLRs 参与炎症小体多蛋白复合物的形成,该复合物由 含有CARD的适配器凋亡相关斑点样蛋白(adaptor apoptosis-associated speck-like protein, ASC)和含有 CARD结构域的pro-caspase-1组成, 激活 caspase-1, 催化亲白介素(IL)-1β和亲IL-18的蛋白水解裂解,然 后释放IL-1β和IL-18,导致促炎反应^[48]。

目前,对于NLRs通路的泛素化调控报道较少。 NLRs炎症小体包括NLRP1、NLRP3、NLRP6和 NAIP/NLRC4,其中,对NLRP3研究相对最多。目 前仅知ALNEMRI等^[49]证明NLRP3的去泛素化是 NLRs形成炎症小体的关键步骤,而去泛素酶BRCC3 介导的NLRP3去泛素化对其激活至关重要^[50]。此 外,NLRs也可协同参与到TLRs通路中,NLRP3可 被TLR4识别信号诱导线粒体活性氧的产生而去泛 素化激活以介导炎性小体的组装和激活,去泛素酶 USP50可以通过去除ASC的K63泛素化,介导ASC寡 聚和NLRP3炎性小体激活^[49,51]。

2.1.4 泛素化修饰调控cGAS-STING信号通路 环 状鸟苷单磷酸(cyclic guanosine monophosphate-adenosine monophosphate, cGAMP)合酶(cyclic GMP-AMP synthase, cGAS)是一种可以识别多类dsDNA的 胞质DNA受体,可催化合成内源性 cGAMP,后者可 被其下游接头蛋白STING识别,进而使TBK1、IRF3 发生磷酸化,最终促进I型干扰素的产生^[52]。也有文 献报道,在SARS-CoV-2感染时,STING与cGAMP结 合后会回到高尔基体,通过招募TBK1和IRF3激活 NF-κB通路^[53]。

STING通路受多种泛素化修饰调控。STING 的多个位点可被TRIM32、TRIM56、TRIM29、 DAPK3等催化发生泛素化修饰,促进其与下游信 号分子TBK1结合,诱导I型干扰素的产生^[24,54-56]。 MUL1催化STING的Lys224位点泛素化,阻断Lys224 泛素化可以特异性地阻止IRF3表达减少I型干扰素 产生^[57]。另外,去泛素酶OTUD5与STING相互作用, 去除其K48泛素链并增强其稳定性,敲除*OTUD5*后, 小鼠更容易感染I型单纯疱疹病毒(HSV-1)^[58]。cGAS 上的泛素化修饰也可由TRIM56催化^[59]。

此外, cGAS-STING通路也参与抗RNA病毒的 天然免疫反应, 麻疹病毒 (measles virus, MeV)和尼 帕病毒 (nipah virus, NiV)感染细胞后, STING发生 K63泛素化以调控cGAS-STING信号通路, 最终产生 抗病毒效应^[52]。而TRIM13则通过催化STING发生 K6连接的泛素化导致STING降解, 负向调控cGAS-STING信号通路^[60]。另外, cGAS被泛素连接酶 MARCH8修饰后无法与DNA结合, 在小鼠DNA病毒 感染模型中, *MARCH8*敲除的小鼠对HSV-1敏感性 更低^[61]。

2.2 病毒利用泛素化进行免疫逃逸

泛素化修饰作为促进蛋白质相互作用的关键 一环,可以促进和协调宿主的抗病毒天然免疫反应。 同样,病毒也会利用宿主泛素化系统负向调控先天 免疫途径并促进其增殖(表1)。

一些病毒可以通过自身合成的蛋白对宿主先天 免疫反应通路中的重要蛋白进行泛素化修饰来实施 免疫逃逸,目前研究较多的是针对RIG-I通路的泛素

Table 1The effects of protein ubiquitination on virus			
病毒	病毒蛋白	底物	影响
Viruses	Virus protein	Substrates	Effects
Arterivirus	Arterivirus OTU	RIG-I ^[75]	Reduce the production of IFN-I
DENV	sfRNA	USP15 ^[71]	Reduce the production of IFN-I
EBOV	VP35	TRIM6 ^[89]	Reduce the production of IFN-I
FMDV	L	RIG-I ^[87]	Reduce the production of IFN-I
EBV	BPLF1	P62 ^[92]	Promote viral infection
HBV	HBx	RIG-I, TRAF3 ^[77]	Reduce the production of IFN-I
HIV-1	RNF39	DDX3X ^[80]	Decrease the signal of RLRs
HCV	/	Riplet ^[29,62]	Reduce the production of IFN-I
HEV	PCP	RIG-I, TBK1 ^[76]	Reduce the production of IFN-I
HSV	VP1-2	STING ^[83]	Reduce the production of IFN-I
HSV	UL36USP	IkBa ^[84]	Decrease the signal of NF-kB
IAV	NS1	Riplet, TRIM25 ^[30,65]	Reduce the production of IFN-I
KSHV	ORF64	RIG-I ^[93-94]	Reduce the production of IFN-I
Nairo	Nairovirus OTU	RIG-I ^[75]	Decrease the signal of RLRs
NiV	NiV M	TRIM6 ^[88]	Decrease the phosphorylation of IKKE
PDCoV	PDCoV N	pRiple ^[63]	Decrease the signal of RLRs
		IRF7 ^[64]	Reduce the production of IFN-I
PEDV	PLP2	RIG-I ^[74]	Reduce the production of IFN-I
RGNNV	/	LjRNF114 ^[95]	Decrease the signal of RLRs
SARS-CoV	Ν	TRIM25 ^[69]	Reduce the production of IFN-I
	PLP	TBK1, TRAF3, TRAF6 ^[86]	Reduce the production of IFN-I
SVCV	SVCV N	p53 ^[90]	Decrease p53-mediated innate immune response
	SVCV P	p53 ^[90]	Decrease p53-mediated innate immune response
SVV	3Cpro	RIG-I, TBK1, TRAF3 ^[78]	Decrease the expression of IFN- β and ISG56
TOSV	NSs	RNF5 ^[72-73]	Reduce the production of IFN-I
EV71	3Cpro	miR-526 ^[81-82]	Reduce the production of IFN-I
HRSV	NS1	TRIM25 ^[66]	Decrease the signal of RLRs
HPV	HPV E6	USP15 ^[70]	Decrease the signal of RLRs
WNV	NS1	RIG-I ^[79]	Reduce the production of IFN-I
SFTSV	NSs	TRIM25 ^[68]	Promote viral infection

表1 蛋白质泛素化修饰对病毒的影响

/: 未确定。

/ / /

/: not determined.

化调控。本文将病毒针对RIG-I通路的泛素化调控免疫逃逸分为以下两种机制。一种是病毒蛋白通过宿主的E3泛素连接酶TRIM25或Riplet发挥作用:RIG-I在病毒感染细胞中的激活依赖于TRIM25或Riplet,TRIM25的SPRY结构域可与RIG-IN末端的第一个CARD结合,催化第二个CARD的Lys172发生K63多聚泛素化,随后RIG-I与MAVS结合激活RLRs信号通路。而Riplet则通过促进RIG-I的CARD结构域和CTD结构域发生K63多聚泛素化激活RIG-I,且Riplet诱导RIG-I的Lys788泛素化可能是TRIM25发挥修饰功能的前提。因此,TRIM25和Riplet自然而然地成为了病

毒攻击的靶点蛋白。丙型肝炎病毒(hepatitis C virus, HCV)感染肝脏细胞后会下调Riplet的表达水平,影响其所介导的RIG-I的泛素化^[29,62]。这种情况也同样发生在猪三角洲冠状病毒(porcine delta coronavirus, PDCoV)感染中,不仅如此,PDCoV N还与IRF7相互作用,促进后者通过蛋白酶体被降解,最终实现免疫逃逸^[63-64]。而甲型流感病毒(influenza A virus, IAV)菌株除与Riplet互作外,还能够利用TRIM25发生免疫逃逸。其机制类似于人呼吸道合胞体病毒(human respiratory syncytial virus, HRSV), IAV的NS1可以通过与TRIM25结合从而抑制RIG-I泛素化^[30,65-66]。缺

失NS1效应区的EALQR基序(AA 191~195)会减弱其 对宿主IFN相关细胞因子表达的抑制作用,从而表 现出更低的病毒毒性,因而EALQR基序或成为未来 小分子药物和疫苗的潜在靶点。严重急性呼吸综合 征冠状病毒(severe acute respiratory syndrome coronavirus, SARS-CoV)的核衣壳蛋白可以与TRIM25的 SPRY结构域结合而破坏TRIM25介导的泛素化,而 重度发热伴血小板减少综合征病毒(severe fever with thrombocytopenia syndrome virus, SFTSV)则可利 用自编码的NSs蛋白将TRIM25劫持到病毒包涵体 中[67-69]。病毒免疫逃逸的智慧之处不仅如此,利用 TRIM25需要通过USP15去泛素化激活的特性,某些 病毒,如人乳头瘤病毒(human papilloma virus, HPV) 编码的癌蛋白HPV E6、登革病毒(DENV)血清型二 株(PR-2B)的黄病毒亚基因组RNA(sfRNA)可通过自 身编码的蛋白抑制TRIM25去泛素进而抑制RIG-I的 激活[70-71]。

另一种机制是病毒蛋白直接阻碍或去除 RIG-I 的泛素化修饰。如托斯卡纳病毒 (toscana virus, TOSV)、猪流行性腹泻病毒 (porcine epidemic diarrhea virus, PEDV)、卡波西肉瘤相关疱疹病毒(Kaposi's sarcoma-associated herpes virus, KSHV),以及动脉 炎病毒 (arterivirus)和 Nairo等病毒编码的蛋白可直接 去除 RIG-I的泛素链^[72-75]。有些病毒编码的蛋白可直接 去除 RIG-I的泛素链^[72-75]。有些病毒编码的去泛素化 蛋白还可以作用于多个受体,如戊型肝炎病毒 (hepatitis E virus, HEV)、乙型肝炎病毒 (hepatitis E virus, HEV)、乙型肝炎病毒 (hepatitis B virus, HBV)、塞内卡谷病毒 (seneca valley virus, SVV)及西 尼罗河病毒(west nile virus, WNV)等^[76-79]。

此外, RLRs信号通路也有一些特殊的免疫逃逸 机制,如RNA解旋酶家族成员DDX3X可识别1型人 类免疫缺陷病毒(human immunodeficiency virus 1, HIV-1)的 ssRNA,通过促进MAVS-DDX3X-TRAF3 复合体的形成,增强抗RNA病毒免疫。而HIV-1感 染诱导表达的RNF39可以促进DDX3X的55、138和 162位点的赖氨酸发生K48偶联的泛素化修饰,抑制 DDX3X表达,阻断MAVS-DDX3-TRAF3复合体的形 成,从而抑制抗病毒天然免疫反应,实现病毒的免疫 逃逸^[80]。肠道病毒71(enterovirus type 71, EV71)编 码的3Cpro抑制宿主miR-526的表达,后者过表达导 致CYLD的上调,由此去除RIG-I的K63泛素链,阻断 RIG-I介导的免疫信号,促进病毒复制^[81-82]。这不难 看出,病毒通过编码蛋白将RLRs通路信号因子去泛 素化,是病毒进行免疫逃逸的重要内容之一。

在长期与宿主免疫系统博弈的过程中,病毒 也进化出针对其他通路信号因子的免疫逃逸机制, 如单纯疱疹病毒(herpes simplex virus, HSV)可调控 cGAS-STING和NF-κB信号通路的泛素化以促进其 在大脑中的免疫逃逸^[83-84];新城疫病毒(newcastle disease virus, NDV)、SARS-CoV、FMDV、NiV及 埃博拉病毒(Ebola virus, EBOV)等病毒可通过自身 编码的蛋白对TLRs信号通路的多种蛋白进行泛素 化调控^[85-89]。有的病毒如鲤春病毒血症病毒(spring virernia of carp virus, SVCV)可通过p53介导的先天 免疫进行逃逸^[90-91];有的病毒如爱泼斯坦巴尔病毒 (epstein barr, EBV)可以编码去泛素化酶BPLF1作用 于p62进行免疫逃逸^[92]。总体来看,目前研究中,针 对哺乳动物病毒的免疫逃逸机制研究较为透彻,但 对于昆虫病毒尤其是杆状病毒逃逸机制知之甚少。

病毒已进化出针对天然免疫反应通路的多种 受体蛋白的免疫逃逸能力,因此研究其蛋白质网络 的复杂变化,进而开发相应的去泛素化酶小分子抑 制剂药品则成为重中之重。

3 DUBs抑制剂在抗病毒治疗中的应用

病毒自编码蛋白可以去除TLRs、RLRs和STING 通路重要分子的泛素链,以逃避宿主的抗病毒天然免 疫反应,因此,针对病毒免疫逃逸开发特定去泛素化 酶抑制剂十分重要。而目前对于DUBs抑制剂的开 发主要应用于恶性肿瘤、神经退行性疾病的治疗, 如针对USP7的小分子抑制剂P5091、GW7647等都 已进入临床应用阶段。

在病毒感染性疾病的治疗中,研究较多的是针 对USP14抑制剂的开发,本文主要介绍两种USP14小 分子抑制剂。第一种USP14的特异性抑制剂IU1,对 于黄病毒尤其是DENV具有良好的抗病毒活性^[96]。 DENV感染尚无特异治疗方法,而疫苗接种仅针对 具有DENV感染史的人群有效。作为一个强活性的、 特异的巯基蛋白酶抑制剂,IU1可以快速并且可逆地 结合激活的USP14,改变USP14的空间结构,从而降 低USP14的活性,防止其结合蛋白酶体,感染DENV 的宿主细胞内IU1浓度达到100 μmol/L时可抑制病 毒的复制,并且IU1对西尼罗河病毒也具有抗病毒活 性^[97]。这表明IU1是黄病毒感染患者治疗的一种潜 在药物。 另一种USP14抑制剂WP1130是最初被认为是 JAK-STAT信号通路的抑制剂,随后发现其可直接抑 制USP9x、USP5、USP14和UCH37的去泛素化活性, 在治疗慢性粒细胞白血病和黑色素瘤中应用较多。 WP1130可以显著抑制小鼠巨噬细胞中的诺如病毒 (murine norovirus 1, MNV-1)的感染,进一步的研究 表明,WP1130可以通过显著抑制USP14介导的肌醇 酶1(inositol-requiring enzyme 1)的失活,激活未折叠 的蛋白反应,抑制病毒感染,并且在其他RNA病毒 如EMCV、Sindbis病毒和拉克罗斯病毒(La Crosse virus)中也表现出类似的抑制作用^[98]。

这些证据表明, DUBs是治疗病毒性传染病的有希望的药物靶点, 针对特定DUBs的抑制剂的开发是一种有效的抗病毒药物筛选策略, 具有良好的前景。

4 总结与展望

天然免疫反应受泛素化调控的机理及病毒的 免疫逃逸机制还有很多未被阐明的地方,如泛素化 介导的病毒免疫逃逸受哪些E3泛素连接酶和相应去 泛素化酶调控,除RIG-I外,还有哪些天然免疫信号 通路因子可被病毒去泛素化,病毒是否会使泛素偶 联酶E2丧失活性,以及面对病毒的逃逸,宿主可以采 取何种方式对抗,诸如此类问题还需要进一步探索。

本文主要从宿主和病毒方面,总结了近年来泛 素化和去泛素化参与调控宿主的抗病毒天然免疫反 应的新发现以及其对病毒免疫逃逸的影响,为后续 研究提供了理论基础。同时,揭示泛素化和去泛素 化在宿主与病毒博弈过程中的作用及其分子调控机 制,有助于寻找病毒性疾病的药物靶点,发现抗病毒 治疗新靶标,为抗病毒药物和疫苗的研发提供新的 思路。

参考文献 (References)

- WITTING K F, MULDER M P C, OVAA H. Advancing our understanding of ubiquitination using the Ub-Toolkit [J]. J Mol Model, 2017, 429(22): 3388-94.
- [2] KOMANDER D, CLAGUE M J, URBE S. Breaking the chains: structure and function of the deubiquitinases [J]. Nat Rev Mol Cell Bio, 2009, 10(8): 550-63.
- [3] HAYWARD J A, MATHUR A, NGO C, et al. Cytosolic recognition of microbes and pathogens: inflammasomes in action [J]. Microbiol Mol Biol R, 2018, 82(4): e00015-18.
- [4] DOHERTY T M, ARDITI M. TB, or not TB: that is the question does TLR signaling hold the answer [J]? J Clin Investig, 2004, 114(12): 1699-703.
- [5] YAMAMOTO M, SATO S, HEMMI H, et al. TRAM is specifi-

cally involved in the Toll-like receptor 4-mediated MyD88independent signaling pathway [J]. Nat Immunol, 2003, 4(11): 1144-50.

- [6] LEE Y S, PARK J S, KIM J H, et al. Smad6-specific recruitment of smurf E3 ligases mediates TGF-beta 1-induced degradation of MyD88 in TLR4 signalling [J]. Nat Commun, 2011, doi: 10.1038/ncomms1469.
- [7] WANG C, CHEN T Y, ZHANG J, et al. The E3 ubiquitin ligase Nrdp1 'preferentially' promotes TLR-mediated production of type I interferon [J]. Nat Immunol, 2009, 10(7): 744-52.
- [8] LEE B C, MIYATA M, LIM J H, et al. Deubiquitinase CYLD acts as a negative regulator for bacterium NTHi-induced inflammation by suppressing k63-linked ubiquitination of MyD88 [J]. Proc Natl Acad Sci USA, 2016, 113(2): E165-E71.
- [9] WALD D, QIN J Z, ZHAO Z D, et al. SIGIRR, a negative regulator of Toll-like receptor-interleukin 1 receptor signaling [J]. Nat Immunol, 2003, 4(9): 920-7.
- [10] ZHOU H, YU M, FUKUDA K, et al. IRAK-M mediates Toll-like receptor/IL-1R-induced NFkB activation and cytokine production [J]. EMBO J, 2013, 32(4): 583-96.
- [11] BRINT E K, XU D M, LIU H Y, et al. ST2 is an inhibitor of interleukin 1 receptor and Toll-like receptor 4 signaling and maintains endotoxin tolerance [J]. Nat Immunol, 2004, 5(4): 373-9.
- PARVATIYAR K, BARBER G N, HARHAJ E W. TAX1BP1 and A20 inhibit antiviral signaling by targeting TBK1-IKKi kinases
 J. J Biol Chem, 2010, 285(20): 14999-5009.
- [13] MAO A P, LI S, ZHONG B, et al. Virus-triggered ubiquitination of TRAF3/6 by cIAP1/2 is essential for induction of interferonbeta (IFN-beta) and cellular antiviral response [J]. J Biol Chem, 2010, 285(13): 9470-6.
- [14] GUICCIARDI M E, MOTT J L, BRONK S F, et al. Cellular inhibitor of apoptosis 1 (cIAP-1) degradation by caspase 8 during TNF-related apoptosis-inducing ligand (TRAIL)-induced apoptosis [J]. Exp Cell Res, 2011, 317(1): 107-16.
- [15] XIAO Y C, JIN J, CHANG M Y, et al. Peli1 promotes microgliamediated CNS inflammation by regulating Traf3 degradation [J]. Nat Med, 2013, 19(5): 595-602.
- [16] WANG H, MENG H, LI X, et al. PELI1 functions as a dual modulator of necroptosis and apoptosis by regulating ubiquitination of RIPK1 and mRNA levels of c-FLIP [J]. Proc Natl Acad Sci USA, 2017, 114(45): 11944-9.
- [17] NAKHAEI P, MESPLEDE T, SOLIS M, et al. The E3 ubiquitin ligase Triad3A negatively regulates the RIG-I/MAVS signaling pathway by targeting TRAF3 for degradation [J]. PLoS Pathog, 2009, 5(11): e1000650.
- [18] HUAI W, SONG H, YU Z, et al. Mint3 potentiates TLR3/4-and RIG-I-induced IFN-beta expression and antiviral immune responses [J]. Proc Natl Acad Sci USA, 2016, 113(42): 11925-30.
- [19] CHUNG H Y, MORITA E, VON SCHWEDLER U, et al. NED-D4L overexpression rescues the release and infectivity of human immunodeficiency virus type 1 constructs lacking PTAP and YPXL late domains [J]. J Virol, 2008, 82(10): 4884-97.
- [20] GAO P, MA X, YUAN M, et al. E3 ligase Nedd4l promotes antiviral innate immunity by catalyzing k29-linked cysteine ubiquitination of TRAF3 [J]. Nat Commun, 2021, 12(1): 1194.
- [21] YU Z, SONG H, JIA M, et al. USP1-UAF1 deubiquitinase complex stabilizes TBK1 and enhances antiviral responses [J]. J Exp Med, 2017, 214(12): 3553-63.

- [22] WU X, LEI C, XIA T, et al. Regulation of TRIF-mediated innate immune response by k27-linked polyubiquitination and deubiquitination [J]. Nat Commun, 2019, 10(1): 4115.
- [23] YANG Y, LIAO B, WANG S Y, et al. E3 ligase WWP2 negatively regulates TLR3-mediated innate immune response by targeting TRIF for ubiquitination and degradation [J]. Proc Natl Acad Sci USA, 2013, 110(13): 5115-20.
- [24] ZHANG J, HU M M, WANG Y Y, et al. TRIM32 protein modulates type I interferon induction and cellular antiviral response by targeting MITA/STING protein for k63-linked ubiquitination [J]. J Biol Chem, 2012, 287(34): 28646-55.
- [25] CHUANG T H, ULEVITCH R J. Triad3A, an E3 ubiquitin-protein ligase regulating Toll-like receptors [J]. Nat Immunol, 2004, 5(5): 495-502.
- [26] ZHANG J, CAO L, GAO A M Y, et al. E3 ligase RNF99 negatively regulates TLR-mediated inflammatory immune response via k48-linked ubiquitination of TAB2 [J]. Cell Death Differ, 2023, 30(4): 966-78.
- [27] CAO Y, SUN Y, CHANG H Y, et al. The E3 ubiquitin ligase RNF182 inhibits TLR-triggered cytokine production through promoting p65 ubiquitination and degradation [J]. FEBS Lett, 2019, 593(22): 3210-9.
- [28] WICHERSKA-PAWLOWSKA K, WROBEL T, RYBKA J. Tolllike receptors (TLRs), NOD-like receptors (NLRs), and RIG-I-like receptors (RLRs) in innate immunity. TLRs, NLRs, and RLRs ligands as immunotherapeutic agents for hematopoietic diseases [J]. Int J Mol Sci, 2021, 22(24): 13397.
- [29] OSHIUMI H, MATSUMOTO M, HATAKEYAMA S, et al. Riplet/RNF135, a RING finger protein, ubiquitinates RIG-I to promote interferon-beta induction during the early phase of viral infection [J]. J Biol Chem, 2009, 284(2): 807-17.
- [30] GACK M U, ALBRECHT R A, URANO T, et al. Influenza A virus NS1 targets the ubiquitin ligase TRIM25 to evade recognition by the host viral RNA sensor RIG-I [J]. Cell Host Microbe, 2009, 5(5): 439-49.
- [31] YAN J, LI Q, MAO A P, et al. TRIM4 modulates type I interferon induction and cellular antiviral response by targeting RIG-I for k63-linked ubiquitination [J]. Mol Cell Biol, 2014, 6(2): 154-63.
- [32] KUNIYOSHI K, TAKEUCHI O, PANDEY S, et al. Pivotal role of RNA-binding E3 ubiquitin ligase MEX3C in RIG-I-mediated antiviral innate immunity [J]. Proc Natl Acad Sci USA, 2014, 111(15): 5646-51.
- [33] CAI X, ZHOU Z, ZHU J, et al. Opposing effects of deubiquitinase OTUD3 in innate immunity against RNA and DNA viruses [J]. Cell Rep, 2022, 39(10): 110920.
- [34] INN K S, GACK M U, TOKUNAGA F, et al. Linear ubiquitin assembly complex negatively regulates RIG-I- and TRIM25mediated type I interferon induction [J]. Mol Cell, 2011, 41(3): 354-65.
- [35] OIKAWA D, SATO Y, ITO H, et al. Linear ubiquitin code: its writer, erasers, decoders, inhibitors, and implications in disorders [J]. Int J Mol Sci, 2020, 21(9): 3381.
- [36] LANG X T, TANG T T, JIN T C, et al. TRIM65-catalized ubiquitination is essential for MDA5-mediated antiviral innate immunity [J]. J Exp Med, 2017, 214(2): 459-73.
- [37] ARIMOTO K I, TAKAHASHI H, HISHIKI T, et al. Negative regulation of the RIG-I signaling by the ubiquitin ligase RNF125

[J]. Proc Natl Acad Sci USA, 2007, 104(18): 7500-5.

- [38] BU L, WANG H, HOU P P, et al. The ubiquitin E3 ligase parkin inhibits innate antiviral immunity through k48-linked polyubiquitination of RIG-I and MDA5 [J]. Front Immunol, 2020, 11: 1926.
- [39] JIA Y X, SONG T, WEI C W, et al. Negative regulation of MAVS-mediated innate immune response by PSMA7 [J]. J Immunol, 2009, 183(7): 4241-8.
- [40] ZHENG J Y, ZHANG Y, ZHI L Y, et al. The novel gene TRIM44L from orange-spotted grouper negatively regulates the interferon response [J]. Fish Shellfish Immun, 2019, 92: 746-55.
- [41] NARAYAN K, WAGGONER L, PHAM S T, et al. TRIM13 is a negative regulator of MDA5-mediated type I interferon production [J]. J Virol, 2014, 88(18): 10748-57.
- [42] ZHAO C Y, JIA M T, SONG H, et al. The E3 ubiquitin ligase TRIM40 attenuates antiviral immune responses by targeting MDA5 and RIG-I [J]. Cell Rep, 2017, 21(6): 1613-23.
- [43] LIU B, CHU H, WU H, et al. The ubiquitin E3 ligase TRIM31 promotes aggregation and activation of the signaling adaptor MAVS through lys63-linked polyubiquitination [J]. Eur J Immunol, 2019, 18(2): 214-24.
- [44] LIU C, HUANG S, WANG X L, et al. The otubain YOD1 suppresses aggregation and activation of the signaling adaptor MAVS through lys63-linked deubiquitination [J]. J Immunol, 2019, 202(10): 2957-70.
- [45] DAI T, WU L, WANG S, et al. FAF1 regulates antiviral immunity by inhibiting MAVS but is antagonized by phosphorylation upon viral infection [J]. Cell Host Microbe, 2018, 24(6): 776-90.
- [46] ZHONG B, ZHANG L, LEI C Q, et al. The ubiquitin ligase RNF5 regulates antiviral responses by mediating degradation of the adaptor protein MITA [J]. Immunity, 2009, 30(3): 397-407.
- [47] D'AMBROSIO E A, DRAKE W R, MASHAYEKHD S, et al. Modulation of the NOD-like receptors NOD1 and NOD2: a chemist's perspective [J]. Bioorg Med Chem Lett, 2019, 29(10): 1153-61.
- [48] BROZ P, DIXIT V M. Inflammasomes: mechanism of assembly, regulation and signalling [J]. Nat Rev Immunol, 2016, 16(7): 407-20.
- [49] JULIANA C, FERNANDES-ALNEMRI T, KANG S, et al. Nontranscriptional priming and deubiquitination regulate NLRP3 inflammasome activation [J]. J Biol Chem, 2012, 287(43): 36617-22.
- [50] PY B F, KIM M S, VAKIFAHMETOGLU-NORBERG H, et al. Deubiquitination of NLRP3 by BRCC3 critically regulates inflammasome activity [J]. Mol Cell, 2013, 49(2): 331-8.
- [51] LEE J Y, SEO D, YOU J, et al. The deubiquitinating enzyme, ubiquitin-specific peptidase 50, regulates inflammasome activation by targeting the ASC adaptor protein [J]. FEBS Lett, 2017, 591(3): 479-90.
- [52] IAMPIETRO M, DUMONT C, MATHIEU C, et al. Activation of cGAS/STING pathway upon paramyxovirus infection [J]. Iscience, 2021, 24(6): 102519.
- [53] NEUFELDT C J, CERIKAN B, CORTESE M, et al. SARS-CoV-2 infection induces a pro-inflammatory cytokine response through cGAS-STING and NFκB [J]. Commun Biol, 2022, 5(1): 45.
- [54] TSUCHIDA T, ZOU J A, SAITOH T, et al. The ubiquitin ligase TRIM56 regulates innate immune responses to intracellular

double-stranded DNA [J]. Immunity, 2010, 33(5): 765-76.

- [55] LI Q, LIN L, TONG Y, et al. TRIM29 negatively controls antiviral immune response through targeting STING for degradation [J]. Cell Discov, 2018, 4: 13.
- [56] TAKAHASHI M, LIO C J, CAMPEAU A, et al. The tumor suppressor kinase DAPK3 drives tumor-intrinsic immunity through the STING-IFN-beta pathway [J]. Nat Immunol, 2021, 22(4): 485-96.
- [57] NI G, KONNO H, BARBER G N. Ubiquitination of STING at lysine 224 controls IRF3 activation [J]. Sci Immunol, 2017, 2(11): e7119.
- [58] GUO Y Y, JIANG F, KONG L L, et al. OTUD5 promotes innate antiviral and antitumor immunity through deubiquitinating and stabilizing STING [J]. Cell Mol Immunol, 2021, 18(8): 1945-55.
- [59] SEO G J, KIM C, SHIN W J, et al. TRIM56-mediated monoubiquitination of cGAS for cytosolic DNA sensing [J]. Nat Commun, 2018, 9(1): 613.
- [60] LI X, YU Z, FANG Q, et al. The transmembrane endoplasmic reticulum-associated E3 ubiquitin ligase TRIM13 restrains the pathogenic-DNA-triggered inflammatory response [J]. Sci Adv, 2022, 8(4): eabh0496.
- [61] YANG X, SHI C, LI H, et al. MARCH8 attenuates cGAS-mediated innate immune responses through ubiquitylation [J]. Science Signaling, 2022, 15(732): eabk3067.
- [62] OSHIUMI H, MIYASHITA M, MATSUMOTO M, et al. A distinct role of Riplet-mediated k63-linked polyubiquitination of the RIG-I repressor domain in human antiviral innate immune responses [J]. PLoS Pathog, 2013, 9(8): e1003533.
- [63] JI L K, LI S S, ZHU W X, et al. Porcine deltacoronavirus nucleocapsid protein suppressed IFN-beta production by interfering porcine RIG-I dsRNA-binding and k63-linked polyubiquitination [J]. Front Immunol, 2019, 10: 1024.
- [64] JI L K, WANG N, MA J J, et al. Porcine deltacoronavirus nucleocapsid protein species-specifically suppressed IRF7-induced type I interferon production via ubiquitin-proteasomal degradation pathway [J]. Vet Microbiol, 2020, 250: 108853.
- [65] RAJSBAUM R, ALBRECHT R A, WANG M K, et al. Speciesspecific inhibition of RIG-I ubiquitination and IFN induction by the influenza A virus NS1 protein [J]. PLoS Pathog, 2012, 8(11): e1003059.
- [66] BAN J, LEE N R, LEE N J, et al. Human respiratory syncytial virus NS 1 targets TRIM25 to suppress RIG-I ubiquitination and subsequent RIG-I-mediated antiviral signaling [J]. Viruses, 2018, 10(12): 716.
- [67] MIN Y Q, NING Y J, WANG H L, et al. A RIG-I-like receptor directs antiviral responses to a bunyavirus and is antagonized by virus-induced blockade of TRIM25-mediated ubiquitination [J]. J Biol Chem, 2020, 295(28): 9691-711.
- [68] MORIYAMA M, IGARASHI M, KOSHIBA T, et al. Two conserved amino acids within the NSs of severe fever with thrombocytopenia syndrome phlebovirus are essential for anti-interferon activity [J]. J Virol, 2018, 92(19): e00706-18.
- [69] GORI SAVELLINI G, ANICHINI G, GANDOLFO C, et al. SARS-CoV-2 N protein targets TRIM25-mediated RIG-I activation to suppress innate immunity [J]. Viruses, 2021, 13(8): 1439.
- [70] CHIANG C, PAULI E K, BIRYUKOV J, et al. The human papillomavirus E6 oncoprotein targets USP15 and TRIM25 to sup-

press RIG-I-mediated innate immune signaling [J]. J Virol, 2018, 92(6): e01737-17.

- [71] MANOKARAN G, FINOL E, WANG C L, et al. Dengue subgenomic RNA binds TRIM25 to inhibit interferon expression for epidemiological fitness [J]. Science, 2015, 350(6257): 217-21.
- [72] SAVELLINI G G, ANICHINI G, GANDOLFO C, et al. Toscana virus non-structural protein NSs acts as E3 ubiquitin ligase promoting RIG-I degradation [J]. PLoS Pathog, 2019, 15(12): e1008186.
- [73] GORI-SAVELLINI G, VALENTINI M, CUSI M G. Toscana virus NSs protein inhibits the induction of type I interferon by interacting with RIG-I [J]. J Virol, 2013, 87(12): 6660-7.
- [74] XING Y, CHEN J, TU J, et al. The papain-like protease of porcine epidemic diarrhea virus negatively regulates type I interferon pathway by acting as a viral deubiquitinase [J]. J Gen Virol 2013, 94: 1554-67.
- [75] VAN KASTEREN P B, BEUGELING C, NINABER D K, et al. Arterivirus and nairovirus ovarian tumor domain-containing deubiquitinases target activated RIG-I to control innate immune signaling [J]. J Virol, 2012, 86(2): 773-85.
- [76] NAN Y, YU Y, MA Z, et al. Hepatitis E virus inhibits type I interferon induction by ORF1 products [J]. J Virol, 2014, 88(20): 11924-32.
- [77] JIANG J, TANG H. Mechanism of inhibiting type I interferon induction by hepatitis B virus X protein [J]. Protein Cell, 2010, 1(12): 1106-17.
- [78] XUE Q, LIU H S, ZHU Z X, et al. Seneca valley virus 3C protease negatively regulates the type I interferon pathway by acting as a viral deubiquitinase [J]. Antiviral Res, 2018, 160: 183-9.
- [79] ZHANG H L, YE H Q, LIU S Q, et al. West Nile virus NS1 antagonizes interferon beta production by targeting RIG-I and MDA5 [J]. J Virol, 2017, 91(18): e02396-16.
- [80] WANG W, JIA M, ZHAO C, et al. RNF39 mediates k48-linked ubiquitination of DDX3X and inhibits RLR-dependent antiviral immunity [J]. Sci Adv, 2021, 7(10): eabe5877.
- [81] FRIEDMAN C S, O'DONNELL M A, LEGARDA-ADDISON D, et al. The tumour suppressor CYLD is a negative regulator of RIG-I-mediated antiviral response [J]. EMBO Rep, 2008, 9(9): 930-6.
- [82] XU C Z, HE X, ZHENG Z R, et al. Downregulation of microRNA miR-526a by enterovirus inhibits RIG-I-dependent innate immune response [J]. J Virol, 2014, 88(19): 11356-68.
- [83] BODDA C, REINERT L S, FRUHWURTH S, et al. HSV1 VP1-2 deubiquitinates STING to block type I interferon expression and promote brain infection [J]. J Exp Med, 2020, 217(7): e20191422.
- [84] WANG S, WANG K Z, LI J, et al. Herpes simplex virus 1 ubiquitin-specific protease UL36 inhibits beta interferon production by deubiquitinating TRAF3 [J]. J Virol, 2013, 87(21): 11851-60.
- [85] SUN Y J, ZHENG H, YU S Q, et al. Newcastle disease virus V protein degrades mitochondrial antiviral signaling protein to inhibit host type I interferon production via E3 ubiquitin ligase RNF5 [J]. J Virol, 2019, 93(18): e00322-19.
- [86] SUN L, XING Y, CHEN X, et al. Coronavirus papain-like proteases negatively regulate antiviral innate immune response through disruption of STING-mediated signaling [J]. PLoS One, 2012, 7(2): e30802.

- [87] WANG D, FANG L, LI P, et al. The leader proteinase of footand-mouth disease virus negatively regulates the type I interferon pathway by acting as a viral deubiquitinase [J]. J Virol, 2011, 85(8): 3758-66.
- [88] BHARAJ P, WANG Y E, DAWES B E, et al. The matrix protein of nipah virus targets the E3-ubiquitin ligase TRIM6 to inhibit the IKK epsilon kinase-mediated type-I IFN antiviral response [J]. PLoS Pathog, 2016, 12(9): e1005880.
- [89] BHARAJ P, ATKINS C, LUTHRA P, et al. The host E3-ubiquitin ligase TRIM6 ubiquitinates the ebola virus VP35 protein and promotes virus replication [J]. J Virol, 2017, 91(18): e00833-17.
- [90] LI S, LU L F, LIU S B, et al. Spring viraemia of carp virus modulates p53 expression using two distinct mechanisms [J]. PLoS Pathog, 2019, 15(3): e1007695.
- [91] SONG Y A, FAN S J, ZHANG D W, et al. Zebrafish maoc1 attenuates spring viremia of Carp Virus propagation by promoting autophagy-lysosome-dependent degradation of viral phosphoprotein [J]. J Virol, 2023, 97(2): e0133822.
- [92] YLA-ANTTILA P, GUPTA S, MASUCCI M G. The Epstein-Barr virus deubiquitinase BPLF1 targets SQSTM1/p62 to inhibit selective autophagy [J]. Autophagy, 2021, 17(11): 3461-74.
- [93] INN K S, LEE S H, RATHBUN J Y, et al. Inhibition of RIG-I-

mediated signaling by Kaposi's sarcoma-associated herpesvirusencoded deubiquitinase ORF64 [J]. J Virol, 2011, 85(20): 10899-904.

- [94] GONZALEZ C M, WANG L, DAMANIA B. Kaposi's sarcomaassociated herpesvirus encodes a viral deubiquitinase [J]. J Virol, 2009, 83(19): 10224-33.
- [95] XIANG Y X, ZHANG W W, JIA P, et al. E3 ubiquitin ligase RNF114 inhibits innate immune response to red-spotted grouper nervous necrosis virus infection in sea perch by targeting MAVS and TRAF3 to mediate their degradation [J]. J Immunol, 2021, 206(1): 77-88.
- [96] NAG D K, FINLEY D. A small-molecule inhibitor of deubiquitinating enzyme USP14 inhibits dengue virus replication [J]. Virus Res, 2012, 165(1): 103-6.
- [97] WANG Y, JIANG Y, DING S, et al. Small molecule inhibitors reveal allosteric regulation of USP14 via steric blockade [J]. Cell Res, 2018, 28(12): 1186-94.
- [98] PERRY J W, AHMED M, CHANG K O, et al. Antiviral activity of a small molecule deubiquitinase inhibitor occurs via induction of the unfolded protein response [J]. PLoS Pathog, 2012, 8(7): e1002783.