

神经肽PACAP对神经内分泌系统的调控

陈尚 范广春 余榕捷*

(暨南大学生命科学技术学院, 生物医药研究院, 广州 510632)

摘要 神经肽类激素垂体腺苷酸环化酶激活多肽(pituitary adenylate cyclase-activating polypeptide, PACAP)最初从牛的脑垂体中被分离; 后续研究发现, PACAP及其受体广泛存在于机体的各个组织器官, 全面参与机体中枢和外周神经内分泌系统的功能调节。在中枢神经内分泌系统层面, PACAP通过促进下丘脑-垂体分泌各种上游激素, 正向调控下游神经内分泌组织和器官的分泌, 发挥神经内分泌网络的“指挥棒”功能; 在外周神经内分泌系统层面, PACAP通过分布在外周神经内分泌组织器官内的受体, 采用自分泌和旁分泌方式, 实现对各种下游激素分泌的精细调控, 发挥维持神经内分泌系统整体和谐平衡的“平衡器”作用。该文就PACAP对中枢和外周神经内分泌系统的调控作用进行整理, 分析得出PACAP对神经内分泌系统的双重调控功能, 为PACAP对神经内分泌系统功能的药用开发奠定理论基础。

关键词 垂体腺苷酸环化酶激活多肽; 神经肽; 激素; 神经内分泌系统; 调控

Regulation of Neuroendocrine System by Neuropeptide PACAP

CHEN Shang, FAN Guangchun, YU Rongjie*

(Institute of Biomedicine, College of Life Science and Technology, Jinan University, Guangzhou 510632, China)

Abstract The neuropeptide hormone PACAP (pituitary adenylate cyclase-activating polypeptide) was originally isolated from the bovine pituitary gland. Subsequent studies have found that PACAP and its receptors are widely present in various tissues and organs of the body, and fully participate in the functional regulation of the central and peripheral neuroendocrine systems. At the level of the central neuroendocrine system, PACAP serve as the “baton” of the neuroendocrine network to regulate the secretion of downstream neuroendocrine tissues and organs by promoting the secretion of various upstream hormones from the hypothalamus-pituitary system. Moreover, at the level of the peripheral neuroendocrine system, PACAP works as the “balancer” to keep the neuroendocrine harmonize by fine adjustment on various downstream hormones secretion through its receptors distributed in the tissues and organs of the peripheral neuroendocrine system and in the way of autocrine and paracrine. This article collates the regulatory effects of PACAP on the central and peripheral neuroendocrine systems, infers the dual regulatory function of PACAP, and lays theoretical foundation for the drug development of PACAP in the neuroendocrine system.

Keywords pituitary adenylate cyclase-activating polypeptide; neuropeptide; hormones; neuroendocrine system; regulation

收稿日期: 2022-04-03 接受日期: 2022-05-05

国家自然科学基金面上项目(批准号: 31670848)和广东省自然科学基金(批准号: 2022A1515011158)资助的课题

*通讯作者。Tel: 020-85220220, E-mail: rongjieyu1123@163.com

Received: April 3, 2022 Accepted: May 5, 2022

This work was supported by the National Natural Science Foundation of China (Grant No.31670848) and the Natural Science Foundation of Guangdong Province (Grant No.2022A1515011158)

*Corresponding author. Tel: +86-20-85220220, E-mail: rongjieyu1123@163.com

神经肽垂体腺苷酸环化酶激活多肽 (pituitary adenylate cyclase-activating polypeptide, PACAP) 属于血管活性肠肽 (vasoactive intestinal polypeptide, VIP)/胰高血糖素/生长激素释放激素/分泌素家族, 在机体各组织器官中广泛分布^[1]。PACAP在体内有 PACAP27 和 PACAP38 两种形式, PACAP27 是 PACAP38 的 N-端截短肽; PACAP27 与 VIP 有高达 68% 的同源性^[2]。PACAP 靶向 3 个 B 类 G 蛋白偶联受体: PAC1-R (pituitary adenylate cyclase 1 receptor)、VPAC1-R (vasoactive intestinal polypeptide receptor 1) 和 VPAC2-R (vasoactive intestinal polypeptide receptor 2), 其中 VPAC1-R 和 VPAC2-R 是 PACAP 和 VIP 的共同受体; PAC1-R 与 PACAP 的亲合力是 VIP 的 1 000 倍, 也是 PACAP 的特异性受体^[3]。作为特异敏感识别 PACAP 的受体, PAC1-R 主要介导了 PACAP 对神经内分泌系统的调控功能。PACAP 的不同基序发挥对 PAC1-R 活性不同调控作用; 其中, PACAP(1~5) 负责正位激活 PAC1-R^[4]; 缺失 PACAP(1~5) 的 PACAP(6~38) 是 PAC1-R 的特异抑制剂^[5]; 此外, PACAP(28~38) 因能有效增强对 PAC1-R 的激活, 发挥了靶向 PAC1-R 的正向别构调节作用^[6]。

PACAP 进化过程极其保守, 人和青蛙的 PACAP 只有 1 个氨基酸的差异。PACAP 及其受体广泛存在于机体神经内分泌系统的各个组织和器官; 已有的研究提示, PACAP 不仅可以作为“指挥棒”调控下丘脑-垂体分泌上游激素, 调节下游神经内分泌系统相关的组织和器官的分泌; 还可以作为一种“平衡器”, 通过广泛存在于神经内分泌组织器官中的受体, 采用自分泌和旁分泌方式对下游神经内分泌进行精细调控; 本文就 PACAP 分别对中枢和外周神经内分泌系统的调节作用进行归纳和综述, 整理 PACAP 对神经内分泌系统的调控规律, 为 PACAP 对神经内分泌系统调控作用相关的药物开发奠定药理学研究的理论基础。

1 PACAP 调控下丘脑-垂体激素分泌

1.1 生长激素 (growth hormone, GH)

研究人员通过对动态灌流的大鼠垂体瘤细胞分泌 GH 的观测发现: PACAP38、PACAP27 和 VIP 均可促进 GH 的释放; 在 1 nmol/L 浓度下, 三者作用效果相当, 但 PACAP38 有效作用时间显著较 PACAP27 和 VIP 延长; PAC1-R 的特异抑制剂 PACAP6-38 和 VIP

拮抗剂可分别有效抑制 PACAP38 和 VIP 的促 GH 释放的活性^[7-8]。在产生 GH 的垂体细胞, 如大鼠垂体瘤 GH3 细胞中, 检测到 PACAP 的多种受体的表达, 包括: PAC1-R 的剪接变体 PAC1-Hop^[9] 和 VPAC2-R^[10]。在绵羊和牛的垂体细胞中, 均发现 PACAP 可有效促进 GH 的分泌^[11-12]。以上发现提示, PACAP 通过受体 PAC1-R 和 VPACR 刺激生长激素分泌细胞释放生长激素, 且 PAC1-R 介导促生长激素分泌的效果更为显著。

1.2 催乳素 (prolactin, PRL)

在大鼠垂体产 PRL 的 GH3 细胞中, PACAP 可通 ERK 和 cAMP/PKA 信号通路增强 PRL 的合成和分泌^[13]。用增强 PAC1-R 表达的载体转染 GH3 细胞后, PACAP 诱导的催乳素基因表达的作用可随 PAC1-R 表达水平的增加而加强, 这提示是 PAC1-R 介导了 PACAP 促 PRL 分泌的作用。此外, PACAP 还协助增强了促甲状腺激素释放激素 (thyrotropin-releasing hormone, TRH) 促进 PRL 分泌的作用, 而且 PACAP 协同 TRH 促 PRL 分泌的作用与 PAC1-R 密切相关^[14]。

1.3 促性腺激素释放激素 (gonadotropin-releasing hormone, GnRH)

1.3.1 GnRH 研究发现, PACAP 可以通过 PAC1-R 上调 GnRH 受体的表达^[15]; 同样 GnRH 可以诱导 PACAP 和 PAC1-R 的表达^[16], 这表明了 PACAP 和 GnRH 之间存在交流 (crosstalk), 可通过相互协同的作用, 以自分泌和旁分泌的方式上调促进性腺激素的表达。

1.3.2 促卵泡激素 (follicle-stimulating hormone, FSH) 在大鼠垂体细胞体外培养系中发现, PACAP 不仅增加了表达卵泡抑素 (follistatin) 的细胞数量, 而且增加了分泌 FSH 的垂体细胞内的颗粒数量; 而分泌卵泡抑素和 FSH 的垂体细胞均高表达 PAC1-R^[17]。体内研究则发现: 成熟大鼠间脑室旁核中 PACAP 与 FSH 的 β 亚基的表达之间的关联密切^[18]; 另有机制研究则提示, PACAP 通过 MAPK/cFOS 途径诱导 FSH 的 β 亚基的表达^[19]。除了可以直接促进垂体细胞分泌 FSH 外^[20], PACAP 还被发现通过其特异 PAC1-R 有效增强 GnRH 对 FSH 表达的调控, 即 PACAP 对 GnRH 的调控具有协同作用^[21]。

1.3.3 黄体生成素 (luteinizing hormone, LH) 在雄性大鼠中发现, 脑室内注射 PACAP 会以剂量依赖的方式增加血浆中 LH 的水平^[22]。同样地, 在垂体细胞

L β T2中发现,一方面,单独的PACAP可促进LH α 和 β 亚基的表达;另一方面,细胞中PAC1-R的存在能够显著增强GnRH对LH表达的影响^[23]。这些研究表明了PACAP可通过多个途径调控LH的分泌。

研究发现,PACAP对LH的调控与发情期和昼夜节律密切相关:处于发情期的雌性或雄性大鼠的垂体中PACAP释放量显著高于发情间期,并且在发情期的雌性大鼠中,PACAP促进LH分泌的作用在清晨与在中午或晚上相比显著增强;而在发情间期开始的前期10~12 h,PACAP会使LH的分泌降低,在间情期后期(16 h)PACAP则不显著影响LH的释放^[24]。

1.3.4 促性腺激素(gonadotropin, GTH) 整理发现,PACAP对下丘脑-垂体分泌GTH的调控作用存在明显的性别差异:雄性大鼠下丘脑的PACAP对促性腺激素的调控作用主要是激动性的(正向);而在雌性大鼠中,它对促性腺激素分泌的调控作用具有激动(正向)和抑制(反向)的双向性^[25],而且作用的方向与雌性的生理周期密切相关;这些发现与PACAP及其受体的表达有显著性别差异相吻合。

1.4 促肾上腺皮质释放激素(corticotropin-releasing hormone, CRH)和促肾上腺皮质激素(adrenocorticotrophic hormone, ACTH)

PACAP在调控下丘脑-垂体-肾上腺轴(hypothalamic-pituitary-adrenal axis, HPA)中发挥显著正向调节作用,即PACAP能有效促进CRH和ACTH在下丘脑-垂体中的分泌。其中,CRH在调节HPA轴方面起着核心作用,已发现PACAP可以通过cAMP/PKA信号通路^[26]刺激CRH基因在下丘脑细胞中的表达^[27]。而对于ACTH,体内研究发现小鼠脑室单独给药PACAP,就可有效促进ACTH分泌的增加^[28];而利用大鼠垂体细胞克隆系ATT-20进行的研究,则发现PACAP不仅单独用药就可以刺激分泌ACTH,并且当CRH与PACAP共同作用ATT-20细胞时,ACTH的分泌更加显著^[29];此结果提示PACAP对ACTH的正向调控可通过多个途径实现。

1.5 促甲状腺激素(thyroid-stimulating hormone, TSH)

在非哺乳动物中,研究发现PACAP和VIP通过PAC1-R和VPAC2-R剂量依赖性刺激牛蛙垂体细胞释放TSH^[30]。而在哺乳动物中,数据较少,只有早期的研究表明体内注射PACAP不影响大鼠^[31]和人类^[32]的血浆TSH浓度,且共定位研究发现,在垂体前叶细

胞中,只有少数细胞既表达TSH又结合PACAP^[33],即PACAP并不改变大鼠垂体前叶细胞TSH的分泌^[34]。由于最新研究相关数据缺乏,PACAP对哺乳动物促甲状腺激素分泌的具体影响还需要更多的研究来证明。

1.6 促黑素细胞激素(melanocyte-stimulating hormone, MSH)

黑皮素原(proopiomelanocortin, POMC)是具有调控应激和抗炎作用的激素MSH的前体。在非洲爪蟾黑素细胞中,发现PACAP通过显著刺激POMC表达进而促进MSH的分泌^[35]。此外,PACAP被发现能够通过cAMP/PKA和PKC信号通路来增加小鼠垂体细胞^[36]和神经中叶^[37]中POMC的mRNA水平,从而促进MSH的分泌;在大鼠中也有相同的发现^[38]。

2 PACAP调控松果体的分泌

PACAP很早已被发现在大鼠松果体中大量存在,并且其分泌具有昼夜节律^[39],而且在松果腺中亦存在高密度的PACAP结合位点^[40]。体外研究表明,PACAP可以有效刺激灌流的大鼠松果腺和培养的松果体细胞分泌褪黑素(melatonin, MT),并参与调节视交叉神经活性的昼夜周期^[41]。而体内的研究证明,PACAP有效参与大鼠和鸡的松果体中褪黑素的释放^[42],即PACAP对松果体MT的释放具有显著正向调控作用。另外,研究发现PACAP刺激松果体MT的释放是通过松果体自身细胞中的PAC1-R受体介导的;并且具有与光照密切相关的作用特点,表现于:如果在黑暗条件下,MT的释放量不会随PACAP刺激的时间变化而发生规律性改变^[43]。同样地,有数据显示PACAP存在于鸡松果体的神经纤维中,松果体细胞中含有PACAP的敏感受体,PACAP可规律性调节MT的释放但不改变松果体的固有生物钟^[44]。这提示了松果体是可以通过自分泌PACAP在PAC1-R的介导下上调自身MT的释放,并调节视交叉神经的昼夜节律的。

3 PACAP调控甲状腺的分泌

PACAP及其3个受体被发现在大鼠甲状腺和甲状旁腺中有分布^[45];在小鼠甲状腺滤泡细胞中则发现存在有VPAC2-R^[46]。在人的甲状腺细胞中发现存在PACAP和PAC1-R的表达^[47];而在哺乳动物中,PA-

CAP27和PACAP38均可快速高效地增加猪甲状腺细胞中cAMP的产生量,并高效增加小鼠体内甲状腺T4的产生量,这两种PACAP均可抑制TSH与甲状腺膜的结合,其中PACAP38的抑制效果更显著^[48]。而最新的研究发现,参与甲状腺激素(thyroid hormone, TH)代谢转化的2型脱碘酶(type 2 deiodinase, D2)可被PACAP上调,提示PACAP直接参与介导了TH的代谢反馈^[49]。

4 PACAP调控肾上腺的分泌

研究发现,PACAP38在大鼠的肾上腺髓质中高表达^[50],且PACAP38的浓度显著高于PACAP27^[51]。此外,PACAP显著上调肾上腺儿茶酚胺(catecholamines, CA)的分泌在犬和大鼠中均有发现^[52-53];并且其上调作用均可被PAC1-R特异拮抗剂以剂量依赖方式所抑制;这提示PACAP是通过PAC1-R介导刺激肾上腺分泌CA的。已有的研究还表明,PACAP既可以作为肾上腺交感神经递质参与儿茶酚胺的调节^[54],又可以作为神经调节剂参与肾上腺的局部调节^[55]。

关于PACAP上调CA的机制研究发现,除PAC1-R介导PACAP刺激CA分泌外,在大鼠肾上腺髓质细胞存在的VIP/PACAP共同受体VPAC1-R和VPAC2-R也能介导PACAP刺激CA的分泌;其中,PAC1-R主要与腺苷酸环化酶级联耦合,VPAC1-R介导与腺苷酸环化酶和磷脂酶C依赖的级联耦合,VPAC2受体仅与磷脂酶C依赖级联耦合,即PACAP刺激儿茶酚胺分泌的途径包括腺苷酸环化酶途径和磷脂酶C途径^[56]。

另外,值得注意的是,PACAP不仅通过促进上游激素CRH和ACTH的分泌,调控肾上腺的分泌,而且可以通过分布在肾上腺中的受体,直接诱导去甲肾上腺素的释放,这在利用PACAP27通过上胰十二指肠动脉局部输注到胰腺和肾上腺中观察去甲肾上腺素浓度的实验中已经得到了证明^[57]。

5 PACAP调控多巴胺的分泌

已有的研究均显示,PACPA与多巴胺(dopamine, DA)的表达之间存在紧密联系,PACAP能够有效减缓DA随龄分泌的衰退;例如:在老年帕金森病的大鼠模型中,脑室注射PACAP可减少老化大鼠脑部DA的随龄损失,并提高具有神经保护作用的帕金森病蛋白7(parkinson disease protein 7, PARK7)的表达水平^[58]。此外,PACAP在保护多巴胺能神经元方面也

发挥显著积极作用,例如:在胚胎大鼠的原代培养物中,PACAP能够阻止6-羟基多巴胺诱导的多巴胺能神经元的死亡^[59];在鱼藤酮(rotenone)诱导的蜗牛帕金森病模型和6-羟基多巴胺诱导的大鼠帕金森病模型中,也同样发现了PACAP能够挽救多巴胺神经元,改善6-羟基多巴胺诱导的帕金森症状大鼠运动能力损伤^[60]。此外,PACAP还可以剂量依赖的方式,减少内源性多巴胺代谢物Salsolinol所导致的多巴胺能神经元凋亡,这种抗凋亡作用可以被PACAP特异性受体PAC1-R拮抗剂PACAP(6-38)所阻断^[61];这提示了PACAP是在PAC1-R的介导下调控多巴胺神经元的保护作用的。

6 PACAP调控胰岛腺的分泌

PACAP27和PACAP38均可以显著刺激胰岛素分泌^[52]。现已有研究发现,PAC1-R、VPAC1-R和VPAC2-R均存在于胰岛细胞中^[62-63],且PACAP38主要在PAC1-R的介导下通过cAMP及其下游效应的PKA途径增加胞内钙离子浓度刺激的胰岛素分泌^[64]。PACAP在体内对葡萄糖诱发的胰岛素释放具有协同促进的作用;表现在单独使用葡萄糖或PACAP进行胰腺局部注射,均能引起胰岛素分泌水平的升高;而当使用葡萄糖和PACAP共同进行注射时,相比单独注射其中一者,共同使用引起胰岛素分泌增加的效果更为显著^[65]。总之,PACAP可以显著正向调控胰岛分泌胰岛素。

7 PACAP调控性腺的分泌

7.1 卵巢

研究表明,PACAP在下丘脑-垂体-性腺轴中发挥重要的调节作用,既可以通过调节神经内分泌活动参与调节卵巢卵泡的发育过程,也可以通过分布在卵巢中的受体,采用自分泌和旁分泌的方式对卵巢的分泌功能进行精细调控。目前,除在大鼠排卵前期的卵泡中检测到了PACAP的表达外,在排卵期的卵巢细胞中也发现存在有PACAP,包括有颗粒细胞、基质细胞,甚至是支配卵巢的神经纤维也存在PACAP^[66]。而在卵泡发育过程中,PACAP可以抑制原代卵泡细胞的转变并抑制其增殖^[67],PACAP和VIP添加到FSH刺激的卵泡中,可抑制卵泡生长^[68];PACAP可使未成熟卵泡细胞的雌二醇分泌增加^[69]。另外,在利用孕马血清促性腺激素和人绒毛膜促性

腺激素治疗后的大鼠卵巢颗粒细胞中,均发现了PACAP以及其特异性受体PAC1-R的存在^[70-71],这表明了PACAP可能是在特异性PAC1-R的介导下发挥调控卵泡发育过程的,但是其他受体是否参与介导PACAP作用,这需要更多的研究证明。

最新的发现,PACAP与松弛素(relaxin)协同作用,不仅可促进大鼠卵巢膜间质细胞和颗粒细胞分泌参与基质重塑的明胶酶(gelatinase),并且PACAP通过cAMP通路,可改变颗粒细胞和膜间质细胞的形状,进而调节卵巢分泌功能^[72],而松弛素单独作用则不能改变颗粒细胞和膜间质细胞的形状。这也表明了PACAP调节卵巢的过程是通过cAMP相关的信号通路介导的,也支持了PACAP本身可以作为卵巢响应促性腺激素的生理介质,促进排卵过程的观点。

相关PACAP促进卵巢激素分泌的研究还表明,PACAP可以刺激大鼠卵巢中的cAMP积累和类固醇生成^[73];在大鼠卵巢颗粒细胞中,PACAP不仅单独增加了雌激素、孕激素和20 α -二氢黄体酮的生成,而且还能够协同增强FSH诱导的黄体酮和20 α -二氢黄体酮积累^[74-75],这表明PACAP在正向调节卵巢激素分泌方面也是可以通过多个途径实现的。

值得注意的是,虽然单独PACAP会促进黄体酮和cAMP积累^[76],但是在LH存在时,高浓度的PACAP则会抑制LH诱导的cAMP的积累和黄体酮的产生^[70],并且PACAP会抑制黄体细胞中LH受体的增加;此研究提示PACAP对卵巢的分泌具有双向性,既可通过PKA途径的参与刺激黄体酮的产生,也参与对黄体酮分泌的负反馈调控,从而发挥全面而精细调节卵巢激素分泌的功能。

7.2 睾丸

PACAP在睾丸中存在并被大量合成,研究已经发现在大鼠的睾丸和附睾中存在有PACAP mRNA,并且PACAP mRNA的表达也会受到睾丸内部温度的调控^[77]。PACAP特异受体PAC1-R的mRNA也被发现存在大鼠睾丸中^[78]。PACAP27和PACAP38均可刺激大鼠睾丸间质细胞睾酮及cAMP的产生,而且PACAP38的作用比PACAP27更为显著^[79],这表明了PACAP单独作用即可刺激睾丸分泌睾酮。另外,存在于睾丸中的PACAP及其受体PAC1-R在繁殖季的表达明显高于非繁殖季^[80]。因此,可以得出结论,睾丸可以通过自分泌PACAP,在PAC1-R的介导下刺激睾丸分泌睾酮的产生,且在繁殖季节PACAP刺激睾丸

分泌睾酮可能比非繁殖季更显著。除此之外,缺乏PACAP会导致精子发生中的信号受到干扰,造成小鼠生育能力降低^[81],这也表明了PACAP具有一定改善雄性小鼠生育的能力。

然而关于睾丸中介导PACAP发挥作用的受体种类存在不同观点。有研究发现生殖细胞精子中不存在PAC1-R mRNA和VPAC1-R mRNA,而VPAC2-R是可以很容易被检测到的^[82],表明PACAP是在VPAC2-R受体的介导下在睾丸中发挥作用的。还有研究表明了PACAP、PAC1-R、VPAC1-R、VPAC2-R在麝鼠的胚芽和体细胞睾丸中均有广泛的表达^[80]。那么睾丸中介导PACAP发挥作用的是哪一种受体,这还需要更多研究来证明。

总之,上述的整理清楚地表明了PACAP对性腺激素具有多方面的调控,PACAP既可以在下丘脑-垂体-性腺轴中通过促进GnRH、FSH和LH的分泌来对性腺分泌性腺激素进行调控,也可以通过分布在性腺中的受体,通过自分泌和旁分泌,或直接促进性激素释放,或对FSH和LH在性腺中作用进行精细调节。

8 PACAP调控脂肪组织的分泌

PACAP及其3种受体均在脂肪细胞中被发现;并且PACAP与VIP均可以诱导大鼠脂肪细胞的脂肪分解^[83]。对于介导脂肪分解的受体类型,有研究通过利用3种受体的拮抗剂和激动剂,发现只有VPAC2-R的激动剂在脂肪分解方面介导和PACAP、VIP相似的活性^[83];研究发现,主要由VPAC2-R介导了PACAP和VIP的促脂肪分解作用^[83-84]。除此之外,PACAP也被报道参与了脂肪细胞分泌的、具有抗炎、抗糖尿病和抗动脉粥样硬化作用的脂联素(adiponectin)的分泌调控^[85]。另有研究发现,VPAC1-R激动剂在降低脂肪增长方面也起着一定作用,使用VPAC1-R激动剂治疗的小鼠,其因高脂肪饮食诱导的体重和白色脂肪组织(附睾和背侧)重量的增加被抑制,高脂肪饮食诱导的血浆葡萄糖、胆固醇和甘油三酯水平的增加也因使用VPAC1-R激动剂被下调^[86]。这说明了VPAC1-R激动剂可能有助于抑制高脂肪饮食诱导的肥胖相关代谢疾病的发病率。

9 PACAP调控胃肠道组织的分泌

PACAP27和PACAP38及其3类受体在胃肠道的表达均已得到证实^[87]。通过对PACAP^{-/-}突变小鼠和

PACAP野生型小鼠分别给药葡聚糖硫酸钠诱导其发生结肠炎, 研究人员发现, PACAP可有效抑制葡聚糖硫酸钠所诱发的促炎细胞因子的产生^[88]。另外, PACAP和VIP均可以放松大鼠平滑肌, 减少胃肠道的基础平滑肌收缩, 且PACAP的作用是VIP的100倍^[89]。

PACAP可通过抑制胃泌素分泌, 从而抑制胃酸的分泌^[90]。促胰液素、生长抑素和前列腺素E₂也被报道参与了PACAP对大鼠胃酸分泌的抑制作用^[91]。但是, 与此存在争议的是, 有研究认为, PACAP通过刺激能够调节胃酸分泌的肠嗜铬样细胞释放组胺, 进而诱导大鼠分泌胃酸^[92], 并且在PAC1-R介导下的PACAP比胃泌素能够更加有效地调节肠嗜铬样细胞的增殖, 这也说明了PACAP对胃酸分泌有正向调节作用。那么, PACAP对胃酸分泌的调控作用到底是刺激还是抑制, 或者是先抑制再刺激, 或先刺激再抑制, 这需要更多的深入研究来探索证明。

除此之外, PACAP还可以对胃十二指肠碳酸氢盐的分泌产生影响, PACAP可由cAMP介导十二指肠碳酸氢盐的分泌, 但不介导胃中碳酸氢盐分泌^[93]; 这些研究表明了PACAP能够对十二指肠起到一定的保护作用。

10 PACAP调控脑肠肽的分泌

已知大脑和肠道之间存在的脑肠轴是由神经内分泌介导的双向应答系统, 脑肠轴通过“脑-肠互动”进行调控。脑肠肽及其受体在大脑和胃肠道中双重分布, 是具有神经递质和激素双重功能的小分子多肽物质, 是脑肠轴发挥重要作用的物质基础。

已有研究发现, PACAP对多种脑肠肽的分泌具有显著的调控作用^[94]。

10.1 胃泌素释放肽(gastrin releasing peptide, GRP)

GRP及其受体广泛分布于哺乳动物的大脑和胃肠道中, 在下丘脑、海马、杏仁核等区域表达较高。目前, 已有研究发现, PACAP对GRP存在正向的调控作用: 在离体的、孤立的猪胃窦组织中, PACAP的免疫反应定位于黏膜层和黏膜下层肌肉; PACAP38(10^{-9} mol/L)即可有效促进孤立的胃窦组织分泌GRP、物质P和VIP^[94]。

10.2 胆囊收缩素(cholecystokinin, CCK)

CCK作为脑肠肽, 既调节胃肠道, 也作用于中枢神经系统^[95]。在小鼠肠肿瘤细胞系STC-1中发现, PACAP能够通过PKA途径调控CCK基因的转录, 从

而促进CCK的分泌^[96-97]。

10.3 降钙素基因相关肽(calcitonin gene-related peptide, CGRP)

脑肠肽CGRP广泛分布于人体的中枢和外周神经内分泌系统, 参与调节胃黏膜血流、胃肠道平滑肌收缩、胃肠道蠕动和迷走神经调控的胃肠黏膜分泌作用^[98]。有研究发现, PACAP38可有效诱导CGRP从三叉神经核的释放^[99]; 以及PACAP作为神经元的营养因子, 可有效调节背根神经节中CGRP的表达^[100]。

10.4 瘦素(leptin)

已有研究发现: 在PAC1-R的介导下, PACAP27和PACAP38均可导致小鼠摄食量明显减少^[101]。在下丘脑-垂体中, PACAP介导的信号转导在瘦素控制进食行为和脂质代谢交感神经流出方面起着关键作用; PAC1-R特异抑制剂PACAP(6-38)可抑制瘦素对脂质代谢交感神经的调控^[102]; 因此, 可以得出结论, PACAP主要通过PAC1-R来调控瘦素分泌进而影响机体食物摄入与肥胖的发生发展, 这也为利用PACAP或PAC1-R激动剂治疗肥胖病症提供了可能。

10.5 神经肽Y(neuropeptide Y, NPY)

NPY是一种广泛存在于中枢和外周神经内分泌系统, 维持内环境稳态的脑肠肽。PACAP对NPY存在正向调控作用: 有研究发现VIP、PACAP27和PACAP38都对交感神经分泌NPY表现出持续有效的刺激作用; 且PACAP27和PACAP38的作用比VIP更强^[103], 其中PACAP38可以使NPY分泌速率增加7至9倍^[104]。关于介导PACAP的刺激作用的受体类型, 有研究发现大部分的NPY神经元表达PAC1-R或VPAC2-R mRNA^[105], 而且NPY相关颈上交感神经节神经元高表达PACAP特异性受体PAC1-R; 研究提示, 在PAC1-R的介导下, PACAP刺激NPY的分泌过程主要与PKA/PAC途径有关^[103]。

此外, CGRP、VIP、PACAP和NPY都在偏头痛的发病机制中起着关键作用。利用电刺激三叉神经节诱导建立偏头痛大鼠模型中, 经过单次电刺激后, 三叉神经节中的CGRP、VIP、PACAP和NPY在不同时间点的表达显著升高并达到峰值, 在反复连续电刺激后, CGRP、PACAP和NPY水平升高更加显著^[106]。这表明了这些神经肽可能在偏头痛的发病机制中起重要作用, 也将为今后的偏头痛的药物研发提供帮助。

以上研究整理发现, PACAP及其受体不仅在神

经内分泌系统表达, 而且在胃肠道内也高密度表达; PACAP不仅作为神经递质, 通过自主神经系统促进脑肠肽的分泌, 而且作为神经调质, 通过分布在肠道内的受体, 采用自分泌或旁分泌, 对脑肠肽的分泌进行精细调控。

11 其他

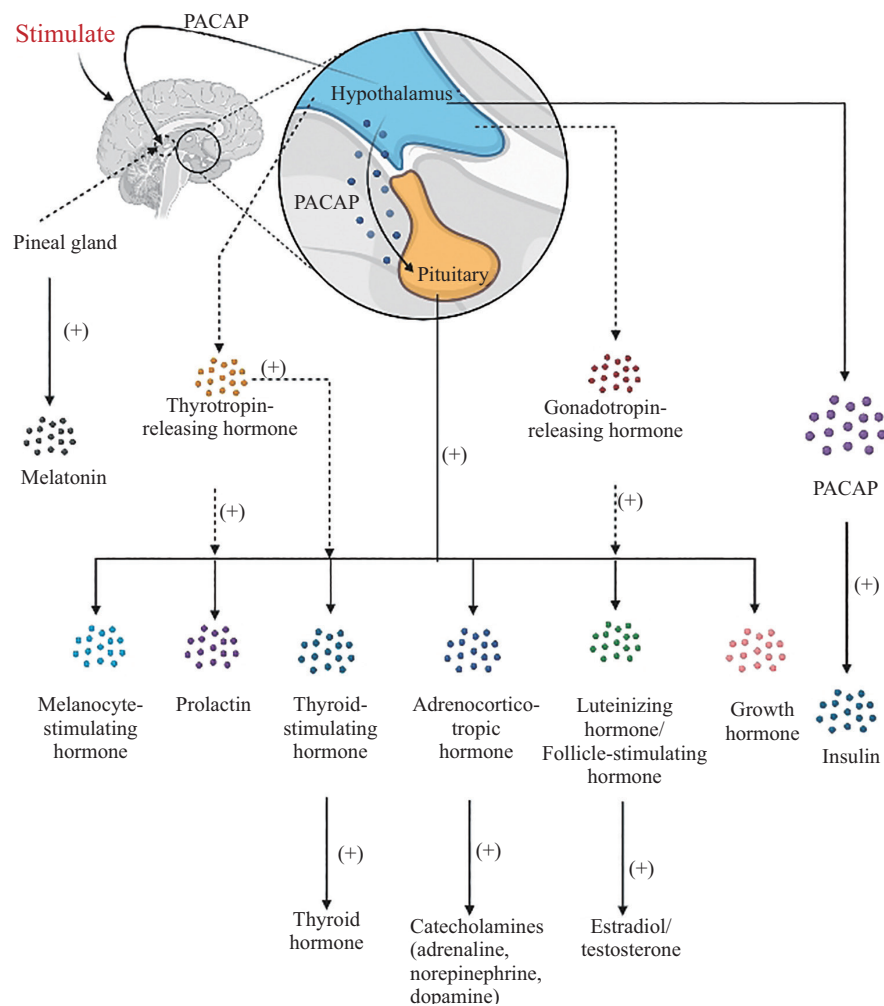
脑啡肽(enkephalin), 是中枢神经系统的类吗啡性神经递质, 可有效抑制疼痛。在大鼠的脑室下区和相邻新皮层区域, 发现PACAP(主要是PACAP38)可显著提高脑啡肽的mRNA水平^[107]。另外, 在牛脑的嗜铬细胞中, 研究人员也发现PACAP提高了甲基脑啡肽和原脑啡肽mRNA水平^[108]。此外, PACAP调控的脑啡肽分泌过程主要是与PAC1-R和钙离子有关, 在PAC1-R的介导下, 其通过激活钙离子通路, 限

制钙流入以促进脑啡肽分泌水平^[107]。

12 总结与展望

总的来说, PACAP对神经内分泌系统的调控具有双重性: 一方面, 在中枢神经内分泌系统层面, 下丘脑-垂体分泌的PACAP既可以通过刺激下丘脑-垂体分泌各种上游和上上游激素来调控下游的外周神经内分泌组织器官的分泌(图1); 另一方面, 在各个外周神经内分泌组织和器官层面, 通过分布在广泛机体各个组织和器官的受体, PACAP以自分泌和旁分泌的方式, 参与各类下游激素分泌的精细调控(图2)。由此可见, PACAP对内分泌的调控具有全面性和精准性的特点。

由以上整理发现, 从整体上看, PACAP作为“指挥棒”, 可通过促进中枢内分泌的垂体分泌各类促激

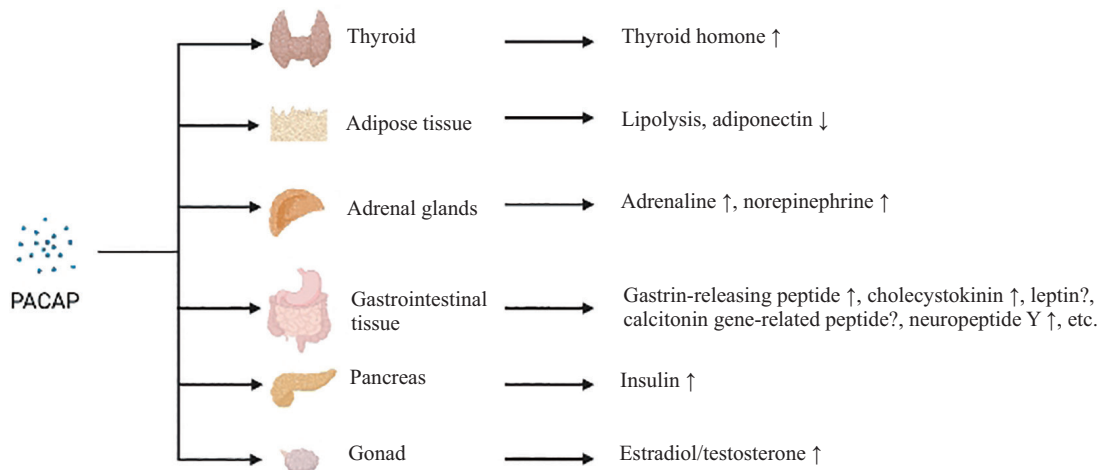


(+): 表示上调激素。

(+): indicates up-regulated hormone.

图1 PACAP对中枢神经内分泌系统的调控

Fig.1 Regulation of PACAP on the central neuroendocrine system



↑: 表示上调激素; ↓: 表示下调激素; ?: 表示未有文献说明激素上调还是下调。

↑: indicates up-regulated hormone; ↓: indicates down-regulated hormone; ?: indicates that there is no literature on whether the hormone is up-regulated or down-regulated.

图2 PACAP对外周神经内分泌系统的调控

Fig.2 Regulation of the peripheral neuroendocrine system by PACAP

素,进而调控垂体-松果体轴、垂体-甲状腺轴、垂体-肾上腺轴、垂体-性腺轴等分泌激素;也作为“平衡器”,通过分布于外周内分泌系统中的受体,发挥对各种激素分泌的精准调控作用;使机体的激素分泌和相互作用达到一种和谐平衡的状态,从而发挥了神经肽PACAP既作为神经递质又作为神经调质、既调控机体响应应激又维护机体代谢平衡的双重生理功能。

本文对PACAP在神经内分泌系统中的作用及其特点进行整理,不仅为PACAP在神经内分泌系统调节功能的相关生物学机制的研究提供理论基础,也将为PACAP及与PACAP相关的活性分子,在神经内分泌系统的相关药用开发方面的提供理论依据。

参考文献 (References)

- [1] MIYATA A, ARIMURA A, DAHL R R, et al. Isolation of a novel 38 residue-hypothalamic polypeptide which stimulates adenylate cyclase in pituitary cells [J]. *Biochem Biophys Res Commun*, 1989, 164(1): 567-74.
- [2] HARMAR A J, ARIMURA A, GOZES I, et al. International Union of Pharmacology. XVIII. Nomenclature of receptors for vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide [J]. *Pharmacol Rev*, 1998, 50(2): 265-70.
- [3] VAUDRY D, FALLUEL-MOREL A, BOURGAULT S, et al. Pituitary adenylate cyclase-activating polypeptide and its receptors: 20 years after the discovery [J]. *Pharmacol Rev*, 2009, 61(3): 283-357.
- [4] GOURLET P, WOUSSEN-COLLE M C, ROBBERECHT P, et al. Structural requirements for the binding of the pituitary adenylate-cyclase-activating peptide to receptors and adenylate-cyclase activation in pancreatic and neuronal membranes [J]. *J Biochem*, 1991, 195(2): 535-41.
- [5] ROBBERECHT P, GOURLET P, DE NEEF P, et al. Structural requirements for the occupancy of pituitary adenylate-cyclase-activating-peptide (PACAP) receptors and adenylate cyclase activation in human neuroblastoma NB-OK-1 cell membranes. Discovery of PACAP (6-38) as a potent antagonist [J]. *Eur J Biochem*, 1992, 207(1): 239-46.
- [6] POUJOL DE MOLLIENS M, LÉTOURNEAU M, DEVOST D, et al. New insights about the peculiar role of the 28-38 C-terminal segment and some selected residues in PACAP for signaling and neuroprotection [J]. *Biochem Pharmacol*, 2018, doi: 10.1016/j.bcp.2018.04.024.
- [7] MURAKAMI Y, KOSHIMURA K, YAMAUCHI K, et al. Pituitary adenylate cyclase activating polypeptide (PACAP) stimulates growth hormone release from GH3 cells through type II PACAP receptor [J]. *Regul Pept*, 1995, 56(1): 35-40.
- [8] KOSHIMURA K, MURAKAMI Y, MITSUSHIMA M, et al. Activation of Na⁺ channels in GH3 cells and human pituitary adenoma cells by PACAP [J]. *Peptides*, 1997, 18(6): 877-83.
- [9] VERTONGEN P, VELKENIERS B, HOOGHE-PETERS E, et al. Differential alternative splicing of PACAP receptor in pituitary cell subpopulations [J]. *Mol Cell Endocrinol*, 1995, 113(2): 131-5.
- [10] COLEMAN D T, CHEN X, SASSAROLI M, et al. Pituitary adenylate cyclase-activating polypeptide regulates prolactin promoter activity via a protein kinase A-mediated pathway that is independent of the transcriptional pathway employed by thyrotropin-releasing hormone [J]. *Endocrinology*, 1996, 137(4): 1276-85.
- [11] SAWANGJAROEN K, ANDERSON S T, CURLEWIS J D. Effects of pituitary adenylate cyclase-activating polypeptide (PACAP) and vasoactive intestinal polypeptide (VIP) on hormone secretion from sheep pituitary cells *in vitro* [J]. *J Neuroendocrinol*

- nol, 1997, 9(4): 279-86.
- [12] HASHIZUME T, SOLIMAN E B, KANEMATSU S. Effects of pituitary adenylate cyclase-activating polypeptide (PACAP), prostaglandin E2 (PGE2) and growth hormone releasing factor (GRF) on the release of growth hormone from cultured bovine anterior pituitary cells *in vitro* [J]. *Domest Anim Endocrinol*, 1994, 11(4): 331-7.
- [13] YONEHARA T, KANASAKI H, YAMAMOTO H, et al. Involvement of mitogen-activated protein kinase in cyclic adenosine 3', 5'-monophosphate-induced hormone gene expression in rat pituitary GH(3) cells [J]. *Endocrinology*, 2001, 142(7): 2811-9.
- [14] MIJIDDORJ T, KANASAKI H, PURWANA I N, et al. Stimulatory effect of pituitary adenylate-cyclase activating polypeptide (PACAP) and its PACAP type I receptor (PAC1R) on prolactin synthesis in rat pituitary somatotroph GH3 cells [J]. *Mol Cell Endocrinol*, 2011, 339(1/2): 172-9.
- [15] KANASAKI H, MIJIDDORJ T, SUKHBAATAR U, et al. Pituitary adenylate cyclase-activating polypeptide (PACAP) increases expression of the gonadotropin-releasing hormone (GnRH) receptor in GnRH-producing GT1-7 cells overexpressing PACAP type I receptor [J]. *Gen Comp Endocrinol*, 2013, 193: 95-102.
- [16] HALVORSON L M. PACAP modulates GnRH signaling in gonadotropes [J]. *Mol Cell Endocrinol*, 2014, 385(1/2): 45-55.
- [17] FUJII Y, OKADA Y, MOORE, JR J P, et al. Evidence that PACAP and GnRH down-regulate follicle-stimulating hormone-beta mRNA levels by stimulating follistatin gene expression: effects on folliculostellate cells, gonadotropes and LbetaT2 gonadotroph cells [J]. *Mol Cell Endocrinol*, 2002, 192(1/2): 55-64.
- [18] MOORE, JR J P, WILSON L, DALGIN A C, et al. Differential expression of the pituitary gonadotropin subunit genes during male rat sexual maturation: reciprocal relationship between hypothalamic pituitary adenylate cyclase-activating polypeptide and follicle-stimulating hormone beta expression [J]. *Biol Reprod*, 2003, 69(1): 234-41.
- [19] YEH D M, COSS D. PACAP induces FSH β gene expression via EPAC [J]. *Mol Cell Endocrinol*, 2019, doi: 10.1016/j.mce.2019.04.018.
- [20] PETERSEN B, BUCHFELDER M, FAHLBUSCH R, et al. Pituitary adenylate cyclase-activating polypeptide directly stimulates LH and FSH secretion by human pituitary gonadotrophinomas [J]. *Exp Clin Endocrinol Diabetes*, 1996, 104(3): 250-5.
- [21] KANASAKI H, PURWANA I N, MIJIDDORJ T, et al. Possible involvement of PACAP and PACAP type I receptor in GnRH-induced FSH β -subunit gene expression [J]. *Regul Pept*, 2011, 167(2/3): 227-32.
- [22] OSUGA Y, MITSUHASHI N, MIZUNO M. *In vivo* effect of pituitary adenylate cyclase activating polypeptide 38 (PACAP 38) on the secretion of luteinizing hormone (LH) in male rats [J]. *Endocrinol Jpn*, 1992, 39(1): 153-6.
- [23] KANASAKI H, ORIDE A, HARA T, et al. Interactions between two different G protein-coupled receptors in reproductive hormone-producing cells: the role of PACAP and its receptor PAC1R [J]. *Int J Mol Sci*, 2016, 17(10): 1635.
- [24] SZABÓ E, NEMESKÉRI A, ARIMURA A, et al. Effect of PACAP on LH release studied by cell immunoblot assay depends on the gender, on the time of day and in female rats on the day of the estrous cycle [J]. *Regul Pept*, 2004, 123(1/2/3): 139-45.
- [25] KÖVES K, SZABÓ E, KÁNTOR O, et al. Current state of understanding of the role of pacap in the hypothalamo-hypophyseal gonadotropin functions of mammals [J]. *Front Endocrinol*, 2020, doi: 10.3389/fendo.2020.00088.
- [26] AGARWAL A, HALVORSON L M, LEGRADI G. Pituitary adenylate cyclase-activating polypeptide (PACAP) mimics neuroendocrine and behavioral manifestations of stress: evidence for PKA-mediated expression of the corticotropin-releasing hormone (CRH) gene [J]. *Brain Res Mol Brain Res*, 2005, 138(1): 45-57.
- [27] KAGEYAMA K, SUDA T. Regulatory mechanisms underlying corticotropin-releasing factor gene expression in the hypothalamus [J]. *Endocr J*, 2009, 56(3): 335-44.
- [28] GASPERINI L, PIUBELLI C, CARBONI L. Proteomics of rat hypothalamus, hippocampus and pre-frontal/frontal cortex after central administration of the neuropeptide PACAP [J]. *Mol Biol Rep*, 2012, 39(3): 2921-35.
- [29] PROPATO-MUSSAFIRI R, KANSE S M, GHATEI M A, et al. Pituitary adenylate cyclase-activating polypeptide releases 7B2, adrenocorticotrophin, growth hormone and prolactin from the mouse and rat clonal pituitary cell lines AtT-20 and GH3 [J]. *J Endocrinol*, 1992, 132(1): 107-13.
- [30] OKADA R, YAMAMOTO K, ITO Y, et al. VIP and PACAP stimulate TSH release from the bullfrog pituitary [J]. *Peptides*, 2007, 28(9): 1784-9.
- [31] HART G R, GOWING H, BURRIN J M. Effects of a novel hypothalamic peptide, pituitary adenylate cyclase-activating polypeptide, on pituitary hormone release in rats [J]. *J Endocrinol*, 1992, 134(1): 33-41.
- [32] CHIODERA P, VOLPI R, CAPRETTI L, et al. Effects of intravenously infused pituitary adenylate cyclase-activating polypeptide on adeno-hypophyseal hormone secretion in normal men [J]. *Neuroendocrinology*, 1996, 64(3): 242-6.
- [33] VIGH S, ARIMURA A, GOTTSCHALL P E, et al. Cytochemical characterization of anterior pituitary target cells for the neuropeptide, pituitary adenylate cyclase activating polypeptide (PACAP), using biotinylated ligands [J]. *Peptides*, 1993, 14(1): 59-65.
- [34] CULLER M D, PASCHALL C S. Pituitary adenylate cyclase-activating polypeptide (PACAP) potentiates the gonadotropin-releasing activity of luteinizing hormone-releasing hormone [J]. *Endocrinology*, 1991, 129(4): 2260-2.
- [35] KIDANE A H, ROUBOS E W, JENKS B G. Pituitary adenylate cyclase-activating polypeptide regulates brain-derived neurotrophic factor exon IV expression through the VPAC1 receptor in the amphibian melanotrope cell [J]. *Endocrinology*, 2008, 149(8): 4177-82.
- [36] KOCH B, LUTZ-BUCHER B. Pituitary adenylate cyclase-activating polypeptide (PACAP) stimulates cyclic AMP formation as well as peptide output of cultured pituitary melanotrophs and AtT-20 corticotrophs [J]. *Regul Pept*, 1992, 38(1): 45-53.
- [37] RENÉ F, MONNIER D, GAIDDON C, et al. Pituitary adenylate cyclase-activating polypeptide transduces through cAMP/PKA and PKC pathways and stimulates proopiomelanocortin gene transcription in mouse melanotopes [J]. *Neuroendocrinology*,

- 1996, 64(1): 2-13.
- [38] TANAKA K, SHIBUYA I, HARAYAMA N, et al. Pituitary adenylate cyclase-activating polypeptide potentiation of Ca^{2+} entry via protein kinase C and A pathways in melanotrophs of the pituitary pars intermedia of rats [J]. *Endocrinology*, 1997, 138(10): 4086-95.
- [39] FUKUHARA C, INOUE S I, MATSUMOTO Y, et al. Pituitary adenylate cyclase-activating polypeptide rhythm in the rat pineal gland [J]. *Neurosci Lett*, 1998, 241(2/3): 115-8.
- [40] MASUO Y, OHTAKI T, MASUDA Y, et al. Binding sites for pituitary adenylate cyclase activating polypeptide (PACAP): comparison with vasoactive intestinal polypeptide (VIP) binding site localization in rat brain sections [J]. *Brain Res*, 1992, 575(1): 113-23.
- [41] CHIK C L, HO A K. Pituitary adenylate cyclase-activating polypeptide: control of rat pineal cyclic AMP and melatonin but not cyclic GMP [J]. *J Neurochem*, 1995, 64(5): 2111-7.
- [42] FALUHELYI N, REGLODI D, LENGVÁRI I, et al. Development of the circadian melatonin rhythm and the effect of PACAP on melatonin release in the embryonic chicken pineal gland. An *in vitro* study [J]. *Regul Pept*, 2004, 123(1/2/3): 23-8.
- [43] NAKAHARA K, ABE Y, MURAKAMI T, et al. Pituitary adenylate cyclase-activating polypeptide (PACAP) is involved in melatonin release via the specific receptor PACAP-r1, but not in the circadian oscillator, in chick pineal cells [J]. *Brain Res*, 2002, 939(1/2): 19-25.
- [44] CSERNUS V, JÓZSA R, REGLODI D, et al. The effect of PACAP on rhythmic melatonin release of avian pineals [J]. *Gen Comp Endocrinol*, 2004, 135(1): 62-9.
- [45] FAHRENKRUG J, HANNIBAL J. Localisation of the neuropeptide PACAP and its receptors in the rat parathyroid and thyroid glands [J]. *Gen Comp Endocrinol*, 2011, 171(1): 105-13.
- [46] HARMAR A J, SHEWARD W J, MORRISON C F, et al. Distribution of the VPAC2 receptor in peripheral tissues of the mouse [J]. *Endocrinology*, 2004, 145(3): 1203-10.
- [47] BARDOSI S, BARDOSI A, NAGY Z, et al. Expression of PACAP and PAC1 receptor in normal human thyroid gland and in thyroid papillary carcinoma [J]. *J Mol Neurosci*, 2016, 60(2): 171-8.
- [48] CHEN W, INUI T, HACHIYA T, et al. Stimulatory action of pituitary adenylate cyclase-activating polypeptide (PACAP) on thyroid gland [J]. *Biochem Biophys Res Commun*, 1993, 194(2): 923-9.
- [49] EGRI P, FEKETE C, DÉNES Á, et al. Pituitary adenylate cyclase-activating polypeptide (pacap) regulates the hypothalamo-pituitary-thyroid (hpt) axis via type 2 deiodinase in male mice [J]. *Endocrinology*, 2016, 157(6): 2356-66.
- [50] WATANABE T, MASUO Y, MATSUMOTO H, et al. Pituitary adenylate cyclase activating polypeptide provokes cultured rat chromaffin cells to secrete adrenaline [J]. *Biochem Biophys Res Commun*, 1992, 182(1): 403-11.
- [51] FRÖDIN M, HANNIBAL J, WULFF B S, et al. Neuronal localization of pituitary adenylate cyclase-activating polypeptide 38 in the adrenal medulla and growth-inhibitory effect on chromaffin cells [J]. *Neuroscience*, 1995, 65(2): 599-608.
- [52] LAMOUCHE S, YAMAGUCHI N. Role of PAC(1) receptor in adrenal catecholamine secretion induced by PACAP and VIP *in vivo* [J]. *Am J Physiol Regul Integr Comp Physiol*, 2001, 280(2): R510-8.
- [53] FUKUSHIMA Y, HIKICHI H, MIZUKAMI K, et al. Role of endogenous PACAP in catecholamine secretion from the rat adrenal gland [J]. *Am J Physiol Regul Integr Comp Physiol*, 2001, 281(5): R1562-7.
- [54] HAMELINK C, TJURMINA O, DAMADZIC R, et al. Pituitary adenylate cyclase-activating polypeptide is a sympathoadrenal neurotransmitter involved in catecholamine regulation and glucose homeostasis [J]. *Proc Natl Acad Sci USA*, 2002, 99(1): 461-6.
- [55] LAMOUCHE S, MARTINEAU D, YAMAGUCHI N. Modulation of adrenal catecholamine release by PACAP *in vivo* [J]. *Am J Physiol*, 1999, 276(1): R162-70.
- [56] MAZZOCCHI G, MALENDOWICZ L K, NERI G, et al. Pituitary adenylate cyclase-activating polypeptide and PACAP receptor expression and function in the rat adrenal gland [J]. *Int J Mol Med*, 2002, 9(3): 233-43.
- [57] YAMAGUCHI N, FUKUSHIMA Y. PACAP enhances stimulation-induced norepinephrine release in canine pancreas *in vivo* [J]. *Can J Physiol Pharmacol*, 1998, 76(7/8): 788-97.
- [58] JUNGLING A, REGLODI D, MAASZ G, et al. Alterations of nigral dopamine levels in parkinson's disease after environmental enrichment and PACAP treatment in aging rats [J]. *Life*, 2021, 11(1): 35.
- [59] TAKEI N, SKOGLÖSA Y, LINDHOLM D. Neurotrophic and neuroprotective effects of pituitary adenylate cyclase-activating polypeptide (PACAP) on mesencephalic dopaminergic neurons [J]. *J Neurosci Res*, 1998, 54(5): 698-706.
- [60] MAASZ G, ZRINYI Z, REGLODI D, et al. Pituitary adenylate cyclase-activating polypeptide (PACAP) has a neuroprotective function in dopamine-based neurodegeneration in rat and snail parkinsonian models [J]. *Dis Model Mech*, 2017, 10(2): 127-39.
- [61] BROWN D, TAMAS A, REGLÖDI D, et al. PACAP protects against salsolinol-induced toxicity in dopaminergic SH-SY5Y cells: implication for Parkinson's disease [J]. *J Mol Neurosci*, 2013, 50(3): 600-7.
- [62] FILIPSSON K, SUNDLER F, HANNIBAL J, et al. PACAP and PACAP receptors in insulin producing tissues: localization and effects [J]. *Regul Pept*, 1998, 74(2/3): 167-75.
- [63] AHRÉN B. Role of pituitary adenylate cyclase-activating polypeptide in the pancreatic endocrine system [J]. *Ann N Y Acad Sci*, 2008, 1144: 28-35.
- [64] LIU M, YANG X, BAI T, et al. PACAP stimulates insulin secretion by PAC(1) receptor and ion channels in β -cells [J]. *Cell Signal*, 2019, doi: 10.1016/j.cellsig.2019.05.006.
- [65] YAMAGUCHI N. Pituitary adenylate cyclase activating polypeptide enhances glucose-evoked insulin secretion in the canine pancreas *in vivo* [J]. *Jop*, 2001, 2(5): 306-16.
- [66] GRÄS S, HANNIBAL J, GEORG B, et al. Transient periovulatory expression of pituitary adenylate cyclase activating peptide in rat ovarian cells [J]. *Endocrinology*, 1996, 137(11): 4779-85.
- [67] LATINI S, CHIARPOTTO M, MUCIACCIA B, et al. Inhibitory effect of pituitary adenylate cyclase activating polypeptide on the initial stages of rat follicle development [J]. *Mol Cell Endocrinol*, 2010, 320(1/2): 34-44.
- [68] CECCONI S, ROSSI G, BARBERI M, et al. Effect of pituitary adenylate cyclase-activating polypeptide and vasoactive intesti-

- nal polypeptide on mouse preantral follicle development *in vitro* [J]. *Endocrinology*, 2004, 145(4): 2071-9.
- [69] GRÅS S, HØST E, FAHRENKRUG J. Role of pituitary adenylate cyclase-activating peptide (PACAP) in the cyclic recruitment of immature follicles in the rat ovary [J]. *Regul Pept*, 2005, 128(1): 69-74.
- [70] KOTANI E, USUKI S, KUBO T. Effect of pituitary adenylate cyclase-activating polypeptide (PACAP) on progesterin biosynthesis in cultured granulosa cells from rat ovary and expression of mRNA encoding PACAP type IA receptor [J]. *J Reprod Fertil*, 1998, 112(1): 107-14.
- [71] KOH P O, KWAK S D, KANG S S, et al. Expression of pituitary adenylate cyclase activating polypeptide (PACAP) and PACAP type I A receptor mRNAs in granulosa cells of preovulatory follicles of the rat ovary [J]. *Mol Reprod Dev*, 2000, 55(4): 379-86.
- [72] TENG C H, KE F C, LEE M T, et al. Pituitary adenylate cyclase-activating polypeptide acts synergistically with relaxin in modulating ovarian cell function in rats [J]. *J Endocrinol*, 2000, 167(1): 61-9.
- [73] ZHONG Y, KASSON B G. Pituitary adenylate cyclase-activating polypeptide stimulates steroidogenesis and adenosine 3', 5'-monophosphate accumulation in cultured rat granulosa cells [J]. *Endocrinology*, 1994, 135(1): 207-13.
- [74] HEINDEL J J, SNEEDEN J, POWELL C J, et al. A novel hypothalamic peptide, pituitary adenylate cyclase-activating peptide, regulates the function of rat granulosa cells *in vitro* [J]. *Biol Reprod*, 1996, 54(3): 523-30.
- [75] VACCARI S, LATINI S, BARBERI M, et al. Characterization and expression of different pituitary adenylate cyclase-activating polypeptide/vasoactive intestinal polypeptide receptors in rat ovarian follicles [J]. *J Endocrinol*, 2006, 191(1): 287-99.
- [76] USUKI S, KOTANI E. Effect of pituitary adenylate cyclase-activating polypeptide (PACAP) on progesterin biosynthesis in cultured luteal cells from rat ovary [J]. *Gynecol Endocrinol*, 2001, 15(3): 184-91.
- [77] LV C M, CHENG D L, ZHAO W, et al. Pituitary adenylate cyclase-activating polypeptide mRNA expression in rat testis and epididymis during postnatal development and experimental cryptorchidism [J]. *Mol Med Rep*, 2011, 4(5): 793-8.
- [78] SHUTO Y, SOMOGYVÁRI-VIGH A, SHIODA S, et al. Effect of hypophysectomy on pituitary adenylate cyclase-activating polypeptide gene expression in the rat testis [J]. *Peptides*, 1995, 16(6): 1039-44.
- [79] EL-GEHANI F, TENA-SEMPERE M, HUHTANIEMI I. Evidence that pituitary adenylate cyclase-activating polypeptide is a potent regulator of fetal rat testicular steroidogenesis [J]. *Biol Reprod*, 2000, 63(5): 1482-9.
- [80] TANG Z, YUAN X, BAI Y, et al. Seasonal changes in the expression of PACAP, VPAC1, VPAC2, PAC1 and testicular activity in the testis of the muskrat (*Ondatra zibethicus*). *Eur J Histochem*, 2022, 66(2): 339.
- [81] REGLODI D, CSEH S, SOMOSKOI B, et al. Disturbed spermatogenic signaling in pituitary adenylate cyclase activating polypeptide-deficient mice [J]. *Reproduction*, 2018, 155(2): 129-39.
- [82] KREMPELS K, USDIN T B, HARTA G, et al. PACAP acts through VIP type 2 receptors in the rat testis [J]. *Neuropeptides*, 1995, 29(6): 315-20.
- [83] AKESSON L, AHRÉN B, EDGREN G, et al. VPAC2-R mediates the lipolytic effects of pituitary adenylate cyclase-activating polypeptide/vasoactive intestinal polypeptide in primary rat adipocytes [J]. *Endocrinology*, 2005, 146(2): 744-50.
- [84] LIJNEN H R, FRESON K, HOYLAERTS M F. Effect of VPAC1 blockade on adipose tissue formation and composition in mouse models of nutritionally induced obesity [J]. *J Obes*, 2010, doi: 10.1155/2010/359527.
- [85] BIK W, WOLINSKA-WITORT E, BARANOWSKA-BIK A, et al. PACAP 38 inhibits adiponectin release [J]. *Neuro Endocrinol Lett*, 2007, 28(2): 166-9.
- [86] YU R J, ZHANG L, YI T H, et al. *In vivo* anti-obesity effect of the agonist for receptor VPAC1 [J]. *Sheng Li Xue Bao*, 2008, 60(6): 751-8.
- [87] HANNIBAL J, EKBLAD E, MULDER H, et al. Pituitary adenylate cyclase activating polypeptide (PACAP) in the gastrointestinal tract of the rat: distribution and effects of capsaicin or denervation [J]. *Cell Tissue Res*, 1998, 291(1): 65-79.
- [88] AZUMA Y T, HAGI K, SHINTANI N, et al. PACAP provides colonic protection against dextran sodium sulfate induced colitis [J]. *J Cell Physiol*, 2008, 216(1): 111-9.
- [89] MUNGAN Z, ARIMURA A, ERTAN A, et al. Pituitary adenylate cyclase-activating polypeptide relaxes rat gastrointestinal smooth muscle [J]. *Scand J Gastroenterol*, 1992, 27(5): 375-80.
- [90] MUNGAN Z, OZMEN V, ERTAN A, et al. Pituitary adenylate cyclase activating polypeptide-27 (PACAP-27) inhibits pentagastrin-stimulated gastric acid secretion in conscious rats [J]. *Regul Pept*, 1992, 38(3): 199-206.
- [91] LI P, CHANG T M, COY D, et al. Inhibition of gastric acid secretion in rat stomach by PACAP is mediated by secretin, somatostatin, and PGE(2) [J]. *Am J Physiol Gastrointest Liver Physiol*, 2000, 278(1): G121-7.
- [92] SANDVIK A K, CUI G, BAKKE I, et al. PACAP stimulates gastric acid secretion in the rat by inducing histamine release [J]. *Am J Physiol Gastrointest Liver Physiol*, 2001, 281(4): G997-G1003.
- [93] TAKEUCHI K, YAGI K, KITAMURA M, et al. Stimulatory effect of PACAP on gastroduodenal bicarbonate secretion in rats [J]. *J Physiol Paris*, 1997, 91(3-5): 229-34.
- [94] TORNØE K, HANNIBAL J, GEORG B, et al. PACAP 1-38 as neurotransmitter in the porcine antrum [J]. *Regul Pept*, 2001, 101(1/2/3): 109-21.
- [95] ZHANG Z H, QIN C K, WU S D, et al. Roles of sphincter of Oddi motility and serum vasoactive intestinal peptide, gastrin and cholecystokinin octapeptide [J]. *World J Gastroenterol*, 2014, 20(16): 4730-6.
- [96] CHANG C H, CHEY W Y, BRAGGINS L, et al. Pituitary adenylate cyclase-activating polypeptide stimulates cholecystokinin secretion in STC-1 cells [J]. *Am J Physiol*, 1996, 271(3Pt1): G516-23.
- [97] DEAVALL D G, RAYCHOWDHURY R, DOCKRAY G J, et al. Control of CCK gene transcription by PACAP in STC-1 cells [J]. *Am J Physiol Gastrointest Liver Physiol*, 2000, 279(3): G605-12.
- [98] KO J K, CHO C H. Adaptive cytoprotection and the brain-gut axis [J]. *Digestion*, 2011, 83 (Suppl 1): 19-24.
- [99] JANSEN-OLESEN I, BAUN M, AMRUTKAR D V, et al. PA-

- CAP-38 but not VIP induces release of CGRP from trigeminal nucleus caudalis via a receptor distinct from the PAC1 receptor [J]. *Neuropeptides*, 2014, 48(2): 53-64.
- [100] LIUDYNO M, SKOGLÖSA Y, TAKEI N, et al. Pituitary adenylate cyclase-activating polypeptide (PACAP) protects dorsal root ganglion neurons from death and induces calcitonin gene-related peptide (CGRP) immunoreactivity *in vitro* [J]. *J Neurosci Res*, 1998, 51(2): 243-56.
- [101] VU J P, GOYAL D, LUONG L, et al. PACAP intraperitoneal treatment suppresses appetite and food intake via PAC1 receptor in mice by inhibiting ghrelin and increasing GLP-1 and leptin [J]. *Am J Physiol Gastrointest Liver Physiol*, 2015, 309(10): G816-25.
- [102] TANIDA M, H AYATA A, SHINTANI N, et al. Central PACAP mediates the sympathetic effects of leptin in a tissue-specific manner [J]. *Neuroscience*, 2013, doi: 10.1016/j.neuroscience.2013.02.016.
- [103] BRAAS K M, MAY V. Pituitary adenylate cyclase-activating polypeptides, PACAP-38 and PACAP-27, regulation of sympathetic neuron catecholamine, and neuropeptide Y expression through activation of type I PACAP/VIP receptor isoforms [J]. *Ann N Y Acad Sci*, 1996, doi: 10.1111/j.1749-6632.1996.tb17484.x.
- [104] MAY V, BRAAS K M. Pituitary adenylate cyclase-activating polypeptide (PACAP) regulation of sympathetic neuron neuropeptide Y and catecholamine expression [J]. *J Neurochem*, 1995, 65(3): 978-87.
- [105] MOUNIEN L, BIZET P, BOUTELET I, et al. Expression of PACAP receptor mRNAs by neuropeptide Y neurons in the rat arcuate nucleus [J]. *Ann N Y Acad Sci*, 2006, 1070(1): 457-61.
- [106] GUO Y, CHENG Y, AN J, et al. Neuropeptide changes in an improved migraine model with repeat stimulations [J]. *Transl Neurosci*, 2021, 12(1): 523-32.
- [107] JUST L, MÖRL F, BÄRMANN C, et al. Evidence for cell specific regulation by PACAP38 of the proenkephalin gene expression in neocortical cells [J]. *Glia*, 2000, 30(3): 242-52.
- [108] HAHM S H, HSU C M, EIDEN L E. PACAP activates calcium influx-dependent and -independent pathways to couple met-enkephalin secretion and biosynthesis in chromaffin cells [J]. *J Mol Neurosci*, 1998, 11(1): 43-56.