

石磊,教授,天津医科大学博士生导师,教育部青年"长江学者"特聘教授。长期 从事基因组稳定性和肿瘤发生发展的表观遗传机制研究。相关研究以独立或共 同通讯作者身份发表在 Genome Biol、J Clin Invest、J Exp Med、Sci Adv、Nat Commun和Proc Natl Acad Sci USA等杂志上。获得国家自然科学基金委优秀青年 基金项目、天津市杰出青年基金项目和霍英东教育基金会高等院校青年教师基 金资助,作为第三完成人于2016年获得国家自然科学二等奖,现为中国抗癌协会 青年理事,天津市抗癌协会理事,中国细胞生物学学会染色质分会委员。

R环形成的调控机制及其生物学意义

李祎 石磊* (天津医科大学基础医学院, 天津 300070)

摘要 R环(R-loop)是由一条DNA:RNA杂交链和一条被置换出的单链DNA组成的三链核酸 结构,通常在转录过程中形成。R环在基因调控、端粒稳定、DNA复制以及组蛋白修饰等方面都 发挥着重要作用。越来越多的研究表明,它们还是复制压力的重要来源,过多的R环累积会造成 DNA损伤以及基因组不稳定。此外,R环与许多人类疾病包括神经紊乱、癌症和自身免疫疾病等 有关。鉴于R环的重要生理功能及其与疾病的潜在关系,该文重点总结了R环的形成机制、生理功 能及R环在基因转录调控和基因组不稳定性中的作用,并讨论了R环调控异常与疾病之间的关系。

关键词 R环; DNA: RNA杂交链; DNA损伤; 基因组不稳定性; 癌症

The Regulation of R-Loop Formation and Its Biological Implications

LI Yi, SHI Lei*

(School of Basic Medical Sciences, Tianjin Medical University, Tianjin 300070, China)

Abstract R-loop is a three-stranded nucleic acid structure consisting of a DNA:RNA hybrid and a displaced single-stranded DNA. It is generally formed during transcription, and plays vital roles in regulating gene expression, DNA replication, DNA damage response, and genome stability. Although R-loop has been implicated in many biological processes, aberrant accumulation of R-loop is one of the major sources of replication stress that threatens genome integrity. Recent evidence suggests that R-loop is involved in many human diseases, including neurological disorders, cancer, and autoimmune diseases. Given the importance of R-loop in physiological and pathological processes, this review summarizes the mechanism of R-loop formation and its biological functions, and also discusses the relationship between R-loop dysregulation and human diseases.

Keywords R-loop; DNA:RNA hybrid; DNA damage; genome instability; cancer

收稿日期: 2022-02-18 接受日期: 2022-03-11

国家自然科学基金(批准号: 81972660)资助的课题

^{*}通讯作者。Tel: 022-83336998, E-mail: shilei@tmu.edu.cn

Received: February 18, 2022 Accepted: March 11, 2022

This work was supported by the National Natural Science Foundation of China (Grant No.81972660)

^{*}Corresponding author. Tel: +86-22-83336998, E-mail: shilei@tmu.edu.cn

R环(R-loop)是一种由转录过程中新生的RNA 链与模板DNA链结合形成的DNA:RNA杂交链,以 及游离的单链 DNA(single-stranded DNA, ssDNA) 构成的三链核酸复合物^[1]。近年来, R环在基因转 录调控和DNA损伤修复等过程中都发挥着重要作 用因而备受关注^[2-3]。一般来说, R环形成于RNA聚 合酶的后侧,其长度可超过1 Kb^[4-5]。根据R环在体 内的功能可分为两类,即"生理性"和"病理性"R环。 "生理性"R环通常需要特定的因素诱导形成,并在 其发挥功能的特定区域进一步增多,主要过程包括 B细胞中的体细胞超突变、免疫球蛋白类转换重组、 DNA复制、CRISPR-Cas9活性、转录起始和终止 的调控、端粒稳态等^[6-7]。然而,过多R环在体内累 积可能会干扰DNA复制、转录和修复,从而破坏基 因组的完整性并与疾病发生发展相关,这称为"病 理性"R环。机体存在着不同的机制来防止或消除 此类R环。

1 R环的形成与调控

R环的形成受基因组DNA序列特征或结构特征 等因素的调节,这些因素包括:有R环倾向的DNA序 列^[8-11],非模板DNA链发生断裂^[12-13],负超螺旋结构^[14], 非典型的DNA结构^[15-17]。它们独立性或协同性地促 进新生RNA和DNA模板链的结合。若RNA的5′端附近 含有四个及以上连续的鸟嘌呤(guanine, G), R环的形成 率会显著提高^[18]。此外,其他因素也可以影响R环的形 成,例如在转录泡后面出现的负超螺旋结构也会增加 新生RNA和模板链结合的概率^[19]。有趣的是,距离转 录起始位点越远, R环出现的概率就越低^[12]。最有可 能的原因是,较长的RNA"尾巴"形成了高级结构或 者被蛋白质复合物所保护,在空间上阻碍RNA侵入 DNA双链。尽管已经有大量的相关研究,但转录过 程中R环形成的实际概率和调控机理还有待深入研究。

R环通常富集于基因启动子区未甲基化修饰的 CpG岛和转录终止区域^[20]。细胞内存在多种蛋白质 机器负责消除异常积累的R环,进而维持转录泡的 完整性和转录的保真性^[21-23]。如:RNA酶RNase H1 和RNase H2利用其核酸内切酶活性将DNA:RNA杂 交链中的RNA水解^[24]。除RNase H1/2外,多种解旋 酶如DHX9^[25]、AQR^[26]、和SETX^[26-27]及染色质重塑 复合物(表1)等,也可以参与DNA:RNA杂交链解旋。 此外,哺乳动物的加帽酶会结合磷酸化的RNAPII, 从而促进R环的形成^[28]。

2 R环的生理功能

目前研究揭示, R环可以调控多种生理活动(图 1),包括:免疫球蛋白抗体类型重组(immunoglobulin class switch recombination, CSR)^[10], CRISPR-Cas9 活性^[44],在线粒体DNA、细菌质粒和噬菌体中调控 DNA复制过程^[6]。DDX1解旋酶可以结合在免疫球 蛋白重链(immunoglobulin heavy, IgH)可变(switch, S)区转录产物的G四链体(G-quadruplex, G4)结构 上,促进RNA与DNA的杂交,为胞嘧啶核苷脱氨酶 (activation-induced cytidine deaminase, AID)提供可 靶向的ssDNA底物,进而促进CSR^[39]。

在基因转录调控过程中,长链非编码RNA(long non-coding RNA, lncRNA)可以通过促进R环的形成 来诱导转录^[45]。在芽殖酵母(*S. cerevisiae*)中的研究 发现, *GAL*基因簇相关的 lncRNA(GAL lncRNA)会 在 *GAL*基因簇中形成R环,其中 DEAD-box RNA解 旋酶 Dbp2可以通过调控该 DNA:lncRNA杂交链,提 高基因转录活性。参与调控细胞和组织完整性的 *VIM*(vimentin)基因在多种癌症中表达上调,有研究

Table 1 Factors involved in DNA:RNA hybrid	
相关因子	功能
Factors	Function
SETX ^[26] , THO complex ^[5] , capping enzyme ^[28] , WDR33 ^[29] , XRN2 ^[30]	Transcription and mRNA processing
Fanconi anemia pathway (FANCM, FANCD2) ^[17] , BRCA1 ^[31] , BRCA2 ^[31] , XPG ^[32] , XPF ^[32] , CtIP ^[33]	DNA repair
FACT complex ^[34] , SIN3A ^[35] , SNF2 ^[36]	Chromatin remodelers
RNase H1 ^[24] , RNase H2 ^[24]	Ribonucleases
DHX9 ^[25] , SETX ^[26] , AQR ^[26] , DDX23 ^[37] , DDX19 ^[38] , DDX1 ^[39] , DDX21 ^[40] , BLM ^[41] , RECQL5 ^[42]	Helicases
TOP1 ^[43] , TOP2 ^[43]	Topoisomerases

表1 DNA:RNA杂交链调控因子 Table 1 Factors involved in DNA:RNA hybrid



RNAP: RNA聚合酶。 RNAP: RNA polymerase.

图1 R环的生理作用(根据参考文献[22]修改) Fig.1 Physiological roles of R-loops (modified from reference [22])

表明*VIM*的表达受制于反义lncRNA VIM-AS1(VIM antisense RNA 1)在*VIM*启动子区域和转录起始位 点(transcription start site, TSS)形成的R环^[46]。此外, 反义lncRNA TARID(TCF21 antisense RNA inducing demethylation)与抑癌基因*TCF21*的启动子区DNA形 成R环,应激反应蛋白GADD45A与之结合后招募甲 基胞嘧啶双加氧酶TET1,诱导局部DNA去甲基化进 而活化*TCF21*的表达^[47]。

R环不仅可以在基因启动子区调控转录活化,还可以于G-rich区域富集终止RNA聚合酶II(RNA polymerase II, RNAPII)的延伸。在该过程中,解旋酶SETX作用于poly(A)下游的R环^[48]并与Tudor结构域蛋白SMN相互作用,允许Xrn2核酸外切酶和终止因子的进入,从而去除这些R环^[49]。SMN通过识别RNAPII的C末端结构域(carboxy-terminal domain,CTD)的精氨酸二甲基化修饰进而募集SETX^[50]。然而,这种机制的普适性如何,尚不清楚。G-rich终止区上的R环也会诱导反义转录,形成双链RNA并募集核酸酶DICER、AGO1和AGO2等RNA干扰(RNAi)因子,导致转录终止^[51]。

3 R环与基因组稳定性

3.1 R环阻碍复制叉进程

由于转录和复制共享相同的DNA模板,当复制复合体遇到转录机器时,会导致转录-复制冲撞(transcription-replication collisions, TRCs)。当复制和转录对向进行时,产生的对向转录-复制冲撞(head-

on transcription-replication collisions, HO TRCs)可 以诱导R环生成,阻碍复制叉前进,导致复制压力和 DNA损伤^[52-53]。过表达RNase H可以显著减少TRCs 引起的DNA损伤,并恢复复制叉进程^[54-55],这表明R环 干扰了复制过程。范可尼贫血信号通路中的FANCA 和FANCD2通常作用于复制叉来调控链内交联,可 以抑制R环的形成进而减轻TRCs造成的损伤。此外, FANCM可以利用其转位酶活性直接消除R环^[56-57]。 相反,当复制和转录同向进行时,共向的转录--复制 冲撞(co-directional transcription-replication collisions, CD TRCs)则会减少R环的累积,基因组对这种冲撞 具有更大的承受性^[58-59],表明机体对不同方向TRCs 的调控机制存在差异。

3.2 R环与转录调控

基因启动子区域的R环可以促进部分基因转录, 但RNA聚合酶的过度停滞或回溯引发的R环,可能 延缓转录从而引起细胞损伤^[60]。这种转录应激不良 状态伴生的R环,在细胞中存在时间较短,没有持续 足够长的时间来阻碍RNA聚合酶。但是,它们可能 会建立瞬时招募平台,通过下游效应因子引发转录 应激^[7]。在不同染色质环境下,R环差异性调节转录 活性的分子机制还有待进一步探究。

目前有研究发现,同源重组修复因子BRCA1 和BRCA2也参与调控转录过程中生成的R环^[61-63]。 BRCA1通过将解旋酶SETX招募到终止位点来消除 R环,从而防止DNA损伤和突变。反之,R环的形成 也会使RNAPII停滞从而阻隔BRCA1和BRCA2,导 致R环进一步积累造成DNA损伤^[64]。这提示, BRCA 参与转录相关R环的识别和消除可能存在精密的负 反馈调控机制。

转录阻断诱发的DNA损伤,可以激活转录偶联 核苷酸切除修复(transcription-coupled nucleotide excision repair, TC-NER)途径。但由于R环比典型的阻断 损伤更大, TC-NER中的核酸酶XPG和XPF切除阻碍 转录的R环,留下一个ssDNA缺口,该缺口可进一步发 展为DNA双链断裂(double strand break, DSB)^[22,26,32,63]。 近期研究发现, XPG和XPF可以通过这种机制消除 酵母细胞和哺乳动物中的R环^[32,65]。虽然该切除方 式可能会导致DNA损伤,但它仍是调控R环和修复 DNA损伤的有效方法。

3.3 R环与DNA损伤应答

虽然R环具有多种生理作用,但R环也是造成 DNA损伤的一个主要的来源,大多数损伤来自于体 内异常积累的R环^[66]。例如,在酵母中HO TRCs是一 种导致DNA损伤的主要R环形式^[67]。在人类细胞中, HO TRCs也已被证明会促进R环进一步累积,进而破 坏复制进程,严重威胁基因组稳定性^[58]。细胞自身 接受R环的量可能存在一个临界阈值,当细胞内R环 的量达到该阈值时,体内消除R环的途径会达到饱 和状态^[7]。这可能会导致R环在体内异常持续增加 并影响复制或转录过程,进而影响基因组稳定性。

ATR和ATM蛋白激酶作为DNA损伤应答(DNA damage response, DDR)和维持基因组稳定性的关键 激酶,分别参与调控复制叉停滞产生的复制压力和 DSB^[68-69]。复制叉停滞可以激活ATR信号通路,复制 叉崩溃产生的DSB则进一步活化ATM信号通路^[22,69]。 然而,并不是所有的R环诱导的复制压力都可以同 时激活ATM和ATR。例如,由剪接因子突变引起的R 环累积造成的复制压力仅激活ATR信号通路^[70]。此 外,最新研究表明,HOTRCs可以特异性激活ATR(图 2A), 而CD TRCs则特异性激活ATM^[58](图2B)。不同 情况下,R环选择性激活ATR或ATM信号通路的机制 有待进一步研究。ATM的激活可能发生在R环异常 积累产生DSB时,或者是在通过移位的ssDNA缺口 进行复制时。在细菌中,同向碰撞时RNA聚合酶的 回溯会导致R环介导的DSB^[71], 真核生物中也可能存 在类似的机制^[58]。R环上停滞的复制叉可以通过将 复制蛋白A(replication protein A, RPA)招募到复制叉 处暴露的ssDNA上从而激活ATR^[7]。然而,其他ATR 激活途径也可能存在。例如,R环中游离的ssDNA, 也可以被RPA包被^[72]。近期研究发现,在有丝分裂 过程中,ATR由R环激活,而不受着丝粒DNA损伤的影 响,从而促进染色体分离^[73]。此外,ATR和ATM的激 活可以促进解旋酶SETX向TRCs的募集^[74];ATR的激 活会使解旋酶DDX19入核,在核内解开DNA:RNA 杂交链,以减轻TRCs^[38]。

3.4 R环与DNA双链断裂

虽然R环是造成DNA损伤的一个潜在原因,但 也有研究表明,DNA:RNA杂交链可以在DNA损伤 后形成。这些DNA:RNA杂交链既存在于转录起始 位点^[75],也可以由DNA末端剪切后的ssDNA产物和 新生RNA杂交形成^[76]。DNA:RNA杂交链能够以多 种方式影响DSB修复,例如该结构可能阻碍损伤修 复因子在DSB位点的募集,或影响DSB附近染色质 结构从而抑制修复^[24,77]。相反,R环也可能促进DSB 修复^[75],R环的过度去除会降低DSB修复的两条主要 途径即同源重组(homologous recombination,HR)和 非同源末端连接(non-homologous end joining, NHEJ) 的修复效率^[1,78-79]。

R环调控DSB修复的方式之一是影响DNA末 端切除效率。在酵母中, R环的形成防止了 DSB 末端过度切除, R环的去除需要DSB应答过程中 ssDNA有效地与RPA结合^[75]。此外,酵母中末端 剪切因子SAE2及其同源基因CtIP被证明可以促 进R环的解开^[33]。相反,人类细胞中的DNA:RNA 杂交链可以增强切除能力^[78]。有趣的是, R环会促 进一种特殊形式的HR:转录相关的同源重组修复 (transcription-associated homologous recombination repair, TA-HRR)。在TA-HRR过程中, DNA:RNA 杂交链可以在DSB位点招募Rad52,促进XPG介导 的R环消除进而启动后续的HR修复过程^[32];若TA-HRR活性降低, DSB修复则倾向于选择 NHEJ途 径,导致基因组稳定性降低。一种更为普遍的调节 机制是, DSB附近的转录产物与HR过程中的重要 三链DNA结构D-loop(displacement loop)形成由重 组因子RAD51相互作用蛋白1(RAD51 associated protein 1, RAD51AP1)所驱动的DR-loop, 该结构可 以促进RAD51的重组活性,提高HR修复效率^[79]。 DNA:RNA杂交链也可能有助于另一种以RNA为替 代模板形式的HR过程。在此过程, 同源RNA分子 被用来代替DNA作为DSB修复的模板^[80-81]。以上研



A: 当复制与转录方向相向时,会发生对向转录--复制冲撞(HO TRCs),R环积累产生复制压力并激活ATR-Chk1信号通路。B: 当复制与转录方向相同时,会发生共向转录--复制冲撞(CD TRCs),R环水平降低并激活ATM-Chk2信号通路。向上的红色箭头:增加;向下的红色箭头:减少。
A: the ATR-Chk1 DNA damage signaling pathway is activated by HO TRCs (head-on transcription-replication collisions). B: co-directional collisions trigger the ATM-Chk2 DNA damage checkpoint. Upward red arrow: increase; downward red arrow: reduce.
图2 对向和共向转录-复制冲撞调控R-环的模型(根据参考文献[58]修改)

Fig.2 Model for head-on and co-directional transcription-replication conflicts regulating R-loop (modified from reference [58])

究提示, R环可以通过不同的机制调控HR修复, 但 调控方式的选择性和特异性还需深入探索。

4 R环与疾病

4.1 癌症

癌症是一种复杂且多样化的疾病,许多癌症表现出高水平的DNA突变和DNA损伤^[82]。R环在转录过程中形成,在调控异常时会导致DNA损伤,影响基因组稳定性,这提示了癌症与R环之间的潜在联系。在高水平雌激素刺激的乳腺癌细胞中,R环积累并驱动了DNA损伤的产生^[55]。RAS原癌基因的突变同样会导致R环的积累,并造成DNA损伤和复制压

力^[54]。

同源重组修复因子BRCA1和BRCA2驱动的信号通路有缺陷时也会诱导R环介导的DNA损伤,这些R环的积累被发现与小鼠乳腺肿瘤的发生有关^[83]。一些癌症诱发因素也会导致BRCA功能不全,从而导致R环调控异常,且R环自身也会干扰BRCA的功能。在尤文氏肉瘤患者细胞中,EWSFLI融合蛋白诱导形成的R环阻断了BRCA1的功能,使这些癌细胞中BRCA1单倍剂量不足,DNA修复无法正常进行。

R环可以通过诱导DNA损伤的方式对癌细胞施 加选择压力。在对白血病骨髓增生异常综合征的研 究中发现,许多剪接因子的突变都可以诱导R环的 形成,这些R环会激活ATR并影响细胞增殖,在这种 压力下仍能增殖的细胞最终可能会发生癌变^[70]。针 对癌症中R环的调控异常进行特定治疗,可能是治 疗某些难治性肿瘤的一种有效方法。例如,在滑膜 肉瘤细胞中,抑制ATR可使肿瘤细胞中R环积累,增 加其对化疗的敏感性^[84]。

最近有研究表明,基因组不稳定性和DNA损伤 也会触发先天免疫和炎症反应通路,特别是通过激 活cGAS-STING途径^[85]。由致癌刺激导致的R环是 诱发DNA损伤的主要来源之一,癌细胞中R环的形 成可能直接激活固有免疫应答^[85]。cGAS-STING的 激活对抗肿瘤免疫至关重要。然而,cGAS-STING 介导的炎症反应也可能会促进某些肿瘤的生长和转 移^[85]。因此,R环是如何影响cGAS-STING和其他先 天免疫信号通路的,同样需要进一步探究。

4.2 神经紊乱性疾病

R环还与一些神经系统疾病相关,在扩展的三 核苷酸DNA重复序列上形成的R环与某些神经疾 病相关基因的转录抑制有关,包括弗里德赖希共济 失调(Friedreich's ataxia)和脆性X综合征(fragile X syndrome)^[86-87]。在这种类型的疾病中, R环的形成 仅限于扩展的三核苷酸,并主要影响含重复序列的 基因^[88]。解旋酶SETX的突变存在于多种神经系统 疾病中,包括II型精神性视觉失明共济失调(ataxiaocular apraxia type 2, AOA2)和IV型肌萎缩性(脊髓) 侧索硬化(amyotrophic lateral sclerosis type 4, ALS4)。 AOA2患者细胞中, R环水平升高^[89], 但这与SETX的 功能是否相关还有待研究。虽然R环水平的升高通 常与疾病相关,但R环水平降低也可能是病理性的。 在ALS4患者细胞中发现, SETX解旋酶功能的增强 使负调控TGF-β的基因启动子处R环水平降低、甲 基化增加,进而导致了ALS4患者细胞TGF-β信号异 常,最终诱发神经元功能障碍和死亡[90]。

4.3 自体免疫性疾病

R环失调在自身免疫性疾病中也发挥着重要 作用。Aicardi Goutieres综合征(Aicardi Goutieres syndrome, AGS)是一种罕见的炎症性疾病, 通常由核酸酶 TREX1、SAMHD1或RNase H2突变引起。这些核酸 酶的突变, 可导致核酸在细胞质内积累、激活cGAS-STING和干扰素应答通路, 引发机体发生炎性反应^[91]。 一方面, AGS细胞中的R环水平升高, 可能改变了基因 表达或重新激活了逆转录因子^[92]; 另一方面, AGS细胞 中R环诱导的停滞复制叉处产生的DNA片段可能被 释放到细胞质中。两者共同参与了cGAS-STING和干 扰素应答通路的激活^[93-94],最终导致AGS的发生。

5 总结与展望

大量实验证据表明, R环的生成和消除依赖于 蛋白质机器的协同性和精密性调节。RNase H作 为核酸内切酶广泛参与了R环的消除, 我们非常好 奇细胞内是否存在核酸外切酶可以降解R环中的 RNA。R环在许多基于染色质的生理活动中发挥着 重要的功能,同时它们也是DNA损伤和基因组不稳 定性的来源。当前, R环形成的分子机制及对基因组 稳定性的影响已引起广泛关注。尽管关于R环与复 制压力应答的关系已经相对清楚,但R环是否可以 直接影响ATR激活或在空间上与复制压力应答机器 产生联系,还有待深入研究。围绕这些科学问题的 研究可以更好地理解R环的生理和病理功能, 加深 我们对相关疾病机理的认识。

参考文献 (References)

- THOMAS M, WHITE R L, DAVIS R W. Hybridization of RNA to double-stranded DNA: formation of R-loops [J]. Proc Natl Acad Sci USA, 1976, 73(7): 2294-8.
- [2] SANTOS-PEREIRA J M, AGUILERA A. R loops: new modulators of genome dynamics and function [J]. Nat Rev Genet, 2015, 16(10): 583-97.
- [3] GARCIA-MUSE T, AGUILERA A. R loops: from physiological to pathological roles [J]. Cell, 2019, 179(3): 604-18.
- [4] GARCIA-PICHARDO D, CANAS J C, GARCIA-RUBIO M L, et al. Histone mutants separate R loop formation from genome instability induction [J]. Mol Cell, 2017, 66(5): 597-609,e5.
- [5] HUERTAS P, AGUILERA A. Cotranscriptionally formed DNA:RNA hybrids mediate transcription elongation impairment and transcription-associated recombination [J]. Mol Cell, 2003, 12(3): 711-21.
- [6] AGUILERA A, GARCIA-MUSE T. R loops: from transcription byproducts to threats to genome stability [J]. Mol Cell, 2012, 46(2): 115-24.
- [7] CROSSLEY M P, BOCEK M, CIMPRICH K A. R-loops as cellular regulators and genomic threats [J]. Mol Cell, 2019, 73(3): 398-411.
- [8] ROY D, YU K, LIEBER M R. Mechanism of R-loop formation at immunoglobulin class switch sequences [J]. Mol Cell Biol, 2008, 28(1): 50-60.
- [9] DANIELS G A, LIEBER M R. RNA:DNA complex formation upon transcription of immunoglobulin switch regions: implications for the mechanism and regulation of class switch recombination [J]. Nucleic Acids Res, 1995, 23(24): 5006-11.
- [10] YU K, CHEDIN F, HSIEH C L, et al. R-loops at immunoglobulin class switch regions in the chromosomes of stimulated B cells

- [11] BELOTSERKOVSKII B P, LIU R, TORNALETTI S, et al. Mechanisms and implications of transcription blockage by guanine-rich DNA sequences [J]. Proc Natl Acad Sci USA, 2010, 107(29): 12816-21.
- [12] ROY D, ZHANG Z, LU Z, et al. Competition between the RNA transcript and the nontemplate DNA strand during R-loop formation *in vitro*: a nick can serve as a strong R-loop initiation site [J]. Mol Cell Biol, 2010, 30(1): 146-59.
- [13] BELOTSERKOVSKII B P, NEIL A J, SALEH S S, et al. Transcription blockage by homopurine DNA sequences: role of sequence composition and single-strand breaks [J]. Nucleic Acids Res, 2013, 41(3): 1817-28.
- [14] MASSE E, DROLET M. Escherichia coli DNA topoisomerase I inhibits R-loop formation by relaxing transcription-induced negative supercoiling [J]. J Biol Chem, 1999, 274(23): 16659-64.
- [15] GRABCZYK E, MANCUSO M, SAMMARCO M C. A persistent RNA.DNA hybrid formed by transcription of the Friedreich ataxia triplet repeat in live bacteria, and by T7 RNAP *in vitro* [J]. Nucleic Acids Res, 2007, 35(16): 5351-9.
- [16] DUQUETTE M L, HANDA P, VINCENT J A, et al. Intracellular transcription of G-rich DNAs induces formation of G-loops, novel structures containing G4 DNA [J]. Genes Dev, 2004, 18(13): 1618-29.
- [17] NEIL A J, LIANG M U, KHRISTICH A N, et al. RNA-DNA hybrids promote the expansion of Friedreich's ataxia (GAA)n repeats via break-induced replication [J]. Nucleic Acids Res, 2018, 46(7): 3487-97.
- [18] ROY D, LIEBER M R. G clustering is important for the initiation of transcription-induced R-loops *in vitro*, whereas high G density without clustering is sufficient thereafter [J]. Mol Cell Biol, 2009, 29(11): 3124-33.
- [19] KUZMINOV A. When DNA topology turns deadly-RNA Polymerases dig in their R-loops to stand their ground: new positive and negative (super)twists in the replication-transcription conflict [J]. Trends Genet, 2018, 34(2): 111-20.
- [20] GINNO P A, LOTT P L, CHRISTENSEN H C, et al. R-loop formation is a distinctive characteristic of unmethylated human CpG island promoters [J]. Mol Cell, 2012, 45(6): 814-25.
- [21] FREUDENREICH C H. R-loops: targets for nuclease cleavage and repeat instability [J]. Curr Genet, 2018, 64(4): 789-94.
- [22] SOLLIER J, CIMPRICH K A. Breaking bad: R-loops and genome integrity [J]. Trends Cell Biol, 2015, 25(9): 514-22.
- [23] GROH M, GROMAK N. Out of balance: R-loops in human disease [J]. PLoS Genet, 2014, 10(9): e1004630.
- [24] AMON J D, KOSHLAND D. RNase H enables efficient repair of R-loop induced DNA damage [J]. eLife, 2016, 5: e20533.
- [25] CHAKRABORTY P, HUANG J T J, HIOM K. DHX9 helicase promotes R-loop formation in cells with impaired RNA splicing [J]. Nat Commun, 2018, 9(1): 4346.
- [26] SOLLIER J, STORK C T, GARCIA-RUBIO M L, et al. Transcription-coupled nucleotide excision repair factors promote Rloop-induced genome instability [J]. Mol Cell, 2014, 56(6): 777-85.
- [27] CHAKRABORTY P, GROSSE F. Human DHX9 helicase preferentially unwinds RNA-containing displacement loops (R-loops) and G-quadruplexes [J]. DNA Repair, 2011, 10(6): 654-65.

- [28] KANEKO S, CHU C, SHATKIN A J, et al. Human capping enzyme promotes formation of transcriptional R loops *in vitro* [J]. Proc Natl Acad Sci USA, 2007, 104(45): 17620-5.
- [29] XU C, WU Z, DUAN H C, et al. R-loop resolution promotes co-transcriptional chromatin silencing [J]. Nat Commun, 2021, 12(1): 1790.
- [30] SKOURTI-STATHAKI K, PROUDFOOT N J, GROMAK N. Human senataxin resolves RNA/DNA hybrids formed at transcriptional pause sites to promote Xrn2-dependent termination [J]. Mol Cell, 2011, 42(6): 794-805.
- [31] TENG Y, YADAV T, DUAN M, et al. ROS-induced R loops trigger a transcription-coupled but BRCA1/2-independent homologous recombination pathway through CSB [J]. Nat Commun, 2018, 9(1): 4115.
- [32] YASUHARA T, KATO R, HAGIWARA Y, et al. Human Rad52 promotes XPG-mediated R-loop processing to initiate transcription-associated homologous recombination repair [J]. Cell, 2018, 175(2): 558-70,e11.
- [33] MAKHARASHVILI N, ARORA S, YIN Y, et al. Sae2/CtIP prevents R-loop accumulation in eukaryotic cells [J]. eLife, 2018, 7.
- [34] HERRERA-MOYANO E, MERGUI X, GARCIA-RUBIO M L, et al. The yeast and human FACT chromatin-reorganizing complexes solve R-loop-mediated transcription-replication conflicts [J]. Genes Dev, 2014, 28(7): 735-48.
- [35] SALAS-ARMENTEROS I, PEREZ-CALERO C, BAYONA-FELIU A, et al. Human THO-Sin3A interaction reveals new mechanisms to prevent R-loops that cause genome instability [J]. EMBO J, 2017, 36(23): 3532-47.
- [36] TANEJA N, ZOFALL M, BALACHANDRAN V, et al. SNF2 family protein Fft3 suppresses nucleosome turnover to promote epigenetic inheritance and proper replication [J]. Mol Cell, 2017, 66(1): 50-62,e6.
- [37] SRIDHARA S C, CARVALHO S, GROSSO A R, et al. Transcription dynamics prevent RNA-mediated genomic instability through SRPK2-dependent DDX23 phosphorylation [J]. Cell Rep, 2017, 18(2): 334-43.
- [38] HODROJ D, RECOLIN B, SERHAL K, et al. An ATR-dependent function for the Ddx19 RNA helicase in nuclear R-loop metabolism [J]. EMBO J, 2017, 36(9): 1182-98.
- [39] RIBEIRO DE ALMEIDA C, DHIR S, DHIR A, et al. RNA helicase DDX1 converts RNA G-quadruplex structures into R-Loops to promote IgH class switch recombination [J]. Mol Cell, 2018, 70(4): 650-62,e8.
- [40] SONG C, HOTZ-WAGENBLATT A, VOIT R, et al. SIRT7 and the DEAD-box helicase DDX21 cooperate to resolve genomic R loops and safeguard genome stability [J]. Genes Dev, 2017, 31(13): 1370-81.
- [41] CHANG E Y, NOVOA C A, ARISTIZABAL M J, et al. RECQlike helicases Sgs1 and BLM regulate R-loop-associated genome instability [J]. J Cell Biol, 2017, 216(12): 3991-4005.
- [42] BARROSO S, HERRERA-MOYANO E, MUNOZ S, et al. The DNA damage response acts as a safeguard against harmful DNA-RNA hybrids of different origins [J]. EMBO Rep, 2019, 20(9): e47250.
- [43] TUDURI S, CRABBE L, CONTI C, et al. Topoisomerase I suppresses genomic instability by preventing interference between replication and transcription [J]. Nat Cell Biol, 2009, 11(11):

1315-24.

- [44] JINEK M, CHYLINSKI K, FONFARA I, et al. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity [J]. Science, 2012, 337(6096): 816-21.
- [45] CLOUTIER S C, WANG S W, MA W K, et al. Regulated formation of lncRNA-DNA hybrids enables faster transcriptional induction and environmental adaptation [J]. Mol Cell, 2016, 61(3): 393-404.
- [46] BOQUE-SASTRE R, SOLER M, OLIVEIRA-MATEOS C, et al. Head-to-head antisense transcription and R-loop formation promotes transcriptional activation [J]. Proc Natl Acad Sci USA, 2015, 112(18): 5785-90.
- [47] ARAB K, KARAULANOV E, MUSHEEV M, et al. GADD45A binds R-loops and recruits TET1 to CpG island promoters [J]. Nat Genet, 2019, 51(2): 217-23.
- [48] SKOURTI-STATHAKI K, PROUDFOOT N J, GROMAK N. Human senataxin resolves RNA/DNA hybrids formed at transcriptional pause sites to promote Xrn2-dependent termination [J]. Mol Cell, 2011, 42(6): 794-805.
- [49] MORALES J C, RICHARD P, PATIDAR P L, et al. XRN2 links transcription termination to DNA damage andreplication stress [J]. PLoS Genet, 2016, 12(7): e1006107.
- [50] ZHAO D Y, GISH G, BRAUNSCHWEIG U, et al. SMN and symmetric arginine dimethylation of RNA polymerase II C-terminal domain control termination [J]. Nature, 2016, 529(7584): 48-53.
- [51] SKOURTI-STATHAKI K, KAMIENIARZ-GDULA K, PROUD-FOOT N J. R-loops induce repressive chromatin marks over mammalian gene terminators [J]. Nature, 2014, 516(7531): 436-9.
- [52] GAN W, GUAN Z, LIU J, et al. R-loop-mediated genomic instability is caused by impairment of replication fork progression [J]. Genes Dev, 2011, 25(19): 2041-56.
- [53] WELLINGER R E, PRADO F, AGUILERA A. Replication fork progression is impaired by transcription in hyperrecombinant yeast cells lacking a functional THO complex [J]. Mol Cell Biol, 2006, 26(8): 3327-34.
- [54] KOTSANTIS P, SILVA L M, IRMSCHER S, et al. Increased global transcription activity as a mechanism of replication stress in cancer [J]. Nat Commun, 2016, 7: 13087.
- [55] STORK C T, BOCEK M, CROSSLEY M P, et al. Co-transcriptional R-loops are the main cause of estrogen-induced DNA damage [J]. eLife, 2016, 5: e17548..
- [56] GARCIA-RUBIO M L, PEREZ-CALERO C, BARROSO S I, et al. The Fanconi Anemia pathway protects genome integrity from R-loops [J]. PLoS Genet, 2015, 11(11): e1005674.
- [57] SCHWAB R A, NIEMINUSZCZY J, SHAH F, et al. The Fanconi Anemia pathway maintains genome stability by coordinating replication and transcription [J]. Mol Cell, 2015, 60(3): 351-61.
- [58] HAMPERL S, BOCEK M J, SALDIVAR J C, et al. Transcription-replication conflict orientation modulates R-loop levels and activates distinct DNA damage responses [J]. Cell, 2017, 170(4): 774-86,e19.
- [59] LANG K S, HALL A N, MERRIKH C N, et al. Replicationtranscription conflicts generate R-loops that orchestrate bacterial stress survival and pathogenesis [J]. Cell, 2017, 170(4): 787-99,e18.
- [60] SAPONARO M, KANTIDAKIS T, MITTER R, et al. RECQL5

controls transcript elongation and suppresses genome instability associated with transcription stress [J]. Cell, 2014, 157(5): 1037-49.

- [61] BHATIA V, BARROSO S I, GARCIA-RUBIO M L, et al. BRCA2 prevents R-loop accumulation and associates with TREX-2 mRNA export factor PCID2 [J]. Nature, 2014, 511(7509): 362-5.
- [62] HATCHI E, SKOURTI-STATHAKI K, VENTZ S, et al. BRCA1 recruitment to transcriptional pause sites is required for R-loopdriven DNA damage repair [J]. Mol Cell, 2015, 57(4): 636-47.
- [63] SHIVJI M K K, RENAUDIN X, WILLIAMS C H, et al. BRCA2 regulates transcription elongation by RNA polymerase II to prevent R-loop accumulation [J]. Cell Rep, 2018, 22(4): 1031-9.
- [64] GORTHI A, ROMERO J C, LORANC E, et al. EWS-FLI1 increases transcription to cause R-loops and block BRCA1 repair in Ewing sarcoma [J]. Nature, 2018, 555(7696): 387-91.
- [65] SU X A, FREUDENREICH C H. Cytosine deamination and base excision repair cause R-loop-induced CAG repeat fragility and instability in *Saccharomyces cerevisiae* [J]. Proc Natl Acad Sci USA, 2017, 114(40): E8392-401.
- [66] COSTANTINO L, KOSHLAND D. The Yin and Yang of R-loop biology [J]. Curr Opin Cell Biol, 2015, 34: 39-45.
- [67] COSTANTINO L, KOSHLAND D. Genome-wide map of Rloop-induced damage reveals how a subset of R-loops contributes to genomic instability [J]. Mol Cell, 2018, 71(4): 487-97,e3.
- [68] BLACKFORD A N, JACKSON S P. ATM, ATR, and DNA-PK: the trinity at the heart of the DNA damage response [J]. Mol Cell, 2017, 66(6): 801-17.
- [69] SALDIVAR J C, CORTEZ D, CIMPRICH K A. The essential kinase ATR: ensuring faithful duplication of a challenging genome [J]. Nat Rev Mol Cell Biol, 2017, 18(10): 622-36.
- [70] CHEN L, CHEN J Y, HUANG Y J, et al. The augmented R-loop ss a unifying mechanism for myelodysplastic syndromes induced by high-risk splicing factor mutations [J]. Mol Cell, 2018, 69(3): 412-25,e6.
- [71] DUTTA D, SHATALIN K, EPSHTEIN V, et al. Linking RNA polymerase backtracking to genome instability in *E. coli* [J]. Cell, 2011, 146(4): 533-43.
- [72] NGUYEN H D, YADAV T, GIRI S, et al. Functions of replication protein A as a sensor of R loops and a regulator of RNaseH1[J]. Mol Cell, 2017, 65(5): 832-47,e4.
- [73] KABECHE L, NGUYEN H D, BUISSON R, et al. A mitosisspecific and R loop-driven ATR pathway promotes faithful chromosome segregation [J]. Science, 2018, 359(6371): 108-14.
- [74] YUCE O, WEST S C. Senataxin, defective in the neurodegenerative disorder ataxia with oculomotor apraxia 2, lies at the interface of transcription and the DNA damage response [J]. Mol Cell Biol, 2013, 33(2): 406-17.
- [75] OHLE C, TESORERO R, SCHERMANN G, et al. Transient RNA-DNA hybrids are required for efficient double-strand break repair [J]. Cell, 2016, 167(4): 1001-13,e7.
- [76] AGUILERA A, GOMEZ-GONZALEZ B. DNA-RNA hybrids: the risks of DNA breakage during transcription [J]. Nat Struct Mol Biol, 2017, 24(5): 439-43.
- [77] COHEN S, PUGET N, LIN Y L, et al. Senataxin resolves RNA:DNA hybrids forming at DNA double-strand breaks to prevent translocations [J]. Nat Commun, 2018, 9(1): 533.

- [78] LU W T, HAWLEY B R, SKALKA G L, et al. Drosha drives the formation of DNA:RNA hybrids around DNA break sites to facilitate DNA repair [J]. Nat Commun, 2018, 9(1): 532.
- [79] OUYANG J, YADAV T, ZHANG J M, et al. RNA transcripts stimulate homologous recombination by forming DR-loops [J]. Nature, 2021, 594(7862): 283-8.
- [80] KESKIN H, SHEN Y, HUANG F, et al. Transcript-RNA-templated DNA recombination and repair [J]. Nature, 2014, 515(7527): 436-9.
- [81] MAZINA O M, KESKIN H, HANAMSHET K, et al. Rad52 Inverse strand exchange drives RNA-templated DNA double-strand break repair [J]. Mol Cell, 2017, 67(1): 19-29,e3.
- [82] HANAHAN D, WEINBERG R A. Hallmarks of cancer: the next generation [J]. Cell, 2011, 144(5): 646-74.
- [83] TAN S L W, CHADHA S, LIU Y, et al. A class of environmental and endogenous toxins induces BRCA2 haploinsufficiency and genome instability [J]. Cell, 2017, 169(6): 1105-18,e15.
- [84] JONES S E, FLEUREN E D G, FRANKUM J, et al. ATR is a therapeutic target in synovial sarcoma [J]. Cancer Res, 2017, 77(24): 7014-26.
- [85] NG K W, MARSHALL E A, BELL J C, et al. cGAS-STING and Cancer: dichotomous roles in tumor immunity and development [J]. Trends Immunol, 2018, 39(1): 44-54.
- [86] COLAK D, ZANINOVIC N, COHEN M S, et al. Promoterbound trinucleotide repeat mRNA drives epigenetic silencing in fragile X syndrome [J]. Science, 2014, 343(6174): 1002-5.

- [87] GROH M, LUFINO M M, WADE-MARTINS R, et al. Rloops associated with triplet repeat expansions promote gene silencing in Friedreich ataxia and fragile X syndrome [J]. PLoS Genet, 2014, 10(5): e1004318.
- [88] LOOMIS E W, SANZ L A, CHEDIN F, et al. Transcriptionassociated R-loop formation across the human FMR1 CGGrepeat region [J]. PLoS Genet, 2014, 10(4): e1004294.
- [89] BECHEREL O J, SUN J, YEO A J, et al. A new model to study neurodegeneration in ataxia oculomotor apraxia type 2 [J]. Hum Mol Genet, 2015, 24(20): 5759-74.
- [90] GRUNSEICH C, WANG I X, WATTS J A, et al. Senataxin mutation reveals how R-Loops promote transcription by blocking DNA methylation at gene promoters [J]. Mol Cell, 2018, 69(3): 426-37,e7.
- [91] CROW Y J, MANEL N. Aicardi-Goutieres syndrome and the type I interferonopathies [J]. Nat Rev Immunol, 2015, 15(7): 429-40.
- [92] LIM Y W, SANZ L A, XU X, et al. Genome-wide DNA hypomethylation and RNA:DNA hybrid accumulation in Aicardi-Goutieres syndrome [J]. eLife, 2015, 4.
- [93] COQUEL F, SILVA M J, TECHER H, et al. SAMHD1 acts at stalled replication forks to prevent interferon induction [J]. Nature, 2018, 557(7703): 57-61.
- [94] MACKENZIE K J, CARROLL P, MARTIN C A, et al. cGAS surveillance of micronuclei links genome instability to innate immunity [J]. Nature, 2017, 548(7668): 461-5.