

香鱼半乳糖凝集素3基因的克隆、鉴定和功能初探

赵 桐 李长红 陆新江 陈 焰*

(宁波大学生物与海洋科学系, 宁波 315211)

摘要 半乳糖凝集素3(galectin-3, Gal3)是凝集素的一类, 在哺乳动物非特异性免疫中具有重要作用, 但在鱼类中尚未见研究报道。该研究通过香鱼(*Plecoglossus altivelis*)单核/巨噬细胞转录组测序获得*Gal3(PaGal3)*的cDNA序列, 全长2 009个核苷酸, 包含一个完整的开放阅读框, 推测其编码一个由380个氨基酸组成、相对分子质量为38.28 kDa的蛋白, 等电点为4.58, N-末端无信号肽。多重序列比对表明, PaGal3具有Galectin(Gal)家族成员典型的结构特征。系统进化树揭示, 各物种Gal3的进化关系符合目前广泛认可的动物分类关系, PaGal3与虹鳟(*Oncorhynchus mykiss*)Gal3紧密成簇, 氨基酸序列同源性达77%。实时荧光定量PCR结果表明, 鳗弧菌(*Vibrio anguillarum*)感染后, 香鱼肝、肾、脾等组织和单核/巨噬细胞中*PaGal3*的mRNA表达量显著上调。Western blot结果表明, PaGal3存在于香鱼血清中。抗体封闭PaGal3后, 香鱼单核/巨噬细胞吞噬细菌能力和杀菌活性均显著下降, 揭示PaGal3可以增强香鱼单核/巨噬细胞功能。综上, PaGal3与鱼类免疫紧密相关, 可能在香鱼炎症免疫中具有重要作用。

关键词 香鱼; 半乳糖凝集素3; 单核/巨噬细胞; 吞噬; 杀菌

Molecular Cloning, Identification and Preliminary Function Study of Galectin-3 Gene in Ayu (*Plecoglossus altivelis*)

Zhao Tong, Li Changhong, Lu Xinjiang, Chen Jiong*

(Department of Biology and Marine Science, Ningbo University, Ningbo 315211, China)

Abstract Galectin-3 (Gal3), a member of lectins, plays an important role in mammalian innate immunity. However, little is known about fish Gal3. In this paper, the nucleotide sequence of a full-length cDNA clone for ayu (*Plecoglossus altivelis*) *Gal3* (*PaGal3*) gene, 2 009 nucleotides in length, was determined by *de novo* transcriptome sequencing of ayu monocytes/macrophages. It contained a completely open reading frame (ORF) which encoded a polypeptide of 380 amino acids with a calculated molecular weight of 38.28 kDa and a theoretical isoelectric point (pI) of 4.58. Multiple alignment showed that PaGal3 had the typical characters of animal galectin (Gal) family. Phylogenetic tree analysis revealed that PaGal3 was most closely related to that of rainbow trout (*Oncorhynchus mykiss*) with 77% amino acid sequence identity, and the relationships of the different Gal3 coincided well with the widely accepted evolutionary relationships of their organisms. After *Vibrio anguillarum* infection, *PaGal3* mRNA expression of liver, kidney, spleen and monocytes/macrophages of ayu increased significantly, which was deter-

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*通讯作者。Tel: 0574-87609571, E-mail: jchen1975@163.com

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*Corresponding author. Tel: +86-574-87609571, E-mail: jchen1975@163.com

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mined by Real-time quantitative PCR (RT-qPCR) method. When we used antibody to block PaGal3, the phagocytic and bacterial killing activities of ayu monocytes/macrophages both decreased significantly, suggesting that PaGal3 could enhance the functions of monocytes/macrophages. In summary, PaGal3 gene is closely related to fish immune response, and may play an important role in the inflammatory immune response of ayu.

Keywords *Plecoglossus altivelis*; galectin-3; monocytes/macrophages; phagocytosis; bacterial killing

半乳糖凝集素(galectin, Gal)是一种在动物免疫组织中普遍表达的模式识别受体(pattern recognition receptor, PRR), 能调控诱发先天免疫反应^[1]。根据糖基识别功能域(carbohydrate-recognition domain, CRD)的数量及结构特征, Gal可分为三种: (1)由约130个氨基酸组成的单CRD型; (2)由两个同源CRD构成的双CRD型, 两个CRD被一个有70个氨基酸的连接体分隔; (3)嵌合型, 含有一个富于脯氨酸和甘氨酸重复序列的氨基端以及一个带有CRD的球形羧基端^[2]。所有的Gal均可形成低聚物, 并有效识别病原体, 参与先天和适应性免疫应答^[3]。此外, 一些Gal还参与调控胰岛素抗性、成脂^[4]及刺激抗肿瘤免疫反应^[5]。近年来, 一些鱼类Gal家族成员相继被报道。例如, Nakamura等^[6]从日本鳗鲡(*Anguilla japonica*)消化道分离的Gal样蛋白能凝集兔红细胞, 还能与肠道细菌结合, 并且其还原型还能诱导宿主细胞黏附到线虫表面, 参与细胞对寄生虫的防御调控^[7]。鳑鲏(*Rhodeus uyekii*)的*Gal9*基因在各组织均表达, 脂多糖(lipopolysaccharide, LPS)处理后, 脾中*Gal9*基因mRNA表达量增加^[8]。大西洋鳕(*Gadus morhua*)的*Gal1*存在于胚胎发育的各个阶段, 能广泛结合凝集革兰氏阴性菌和阳性菌, 并且在细菌感染后分泌增多, 揭示它可能参与大西洋鳕的发育和抗细菌感染免疫防御反应^[9]。

Galetin-3(Gal3)为Gal家族成员, 在动物体内表达量高, 已被发现广泛分布于若干组织和发育阶段。在哺乳动物中, Gal3已被证实具有多种生物学功能, 包括介导宿主和病原体相互作用、调节免疫、胚胎发生、血管生成、细胞迁移、伤口愈合和细胞凋亡等^[10]。研究表明, Gal3由巨噬细胞大量表达并分泌^[11], 影响巨噬细胞表达促炎性细胞因子^[12], 并介导细菌对巨噬细胞的感染^[13]。在急性免疫反应中, Gal3参与诸多进程, 包括中性粒细胞的活化与黏着、巨噬细胞的趋化以及肥大细胞的激活等, 缺乏Gal3的细胞则表现出吞噬作用和选择性激活的效率降低^[14]。然而在鱼类中, Gal3的功能及作用机制尚未见研究报道。

香鱼(*Plecoglossus altivelis*)是中国、日本和朝鲜等东亚地区国家特有的一种降海洄游鱼类, 因其肉质鲜美, 人工养殖不断扩大。高密度人工养殖易导致病害频发, 主要为细菌性病害, 其中鳗弧菌(*Vibrio anguillarum*)是常见的病原之一^[15]。人工养殖香鱼要求无污染和绿色健康, 抗生素和农药的使用受到诸多限制。因此, 有必要深入研究香鱼免疫机制, 为指导病害防治和抗病遗传育种奠定理论基础^[16]。鉴于Gal家族成员在动物抗菌免疫反应中的重要作用, 我们拟对香鱼Gal3(PaGal3)进行研究, 测定其基因的cDNA全序列, 分析其结构特征、系统进化关系及mRNA在免疫器官和细胞中的表达特征, 并研究鳗弧菌感染后香鱼免疫组织及免疫细胞中PaGal3 mRNA的表达变化; 原核表达PaGal3重组蛋白并制备抗体, 抗体封闭单核/巨噬细胞上清中的PaGal3, 检测细胞吞噬能力和杀菌活性的变化, 为进一步深入研究Gal3在鱼类炎症反应中的作用奠定基础。

1 材料与方法

1.1 材料与试剂

健康香鱼(体重20~25 g/条)购自宁波水产大世界。ICR小鼠购自浙江省医学科学院动物中心。大肠杆菌(*Escherichia coli*) TG1和BL21 plys E菌株、载体pET-28a和鳗弧菌香鱼分离株ayu-H080701等由本实验室保存。Trizol试剂、Oligotex-dT30 mRNA Purification Kit、AMV逆转录酶、Ex Taq DNA聚合酶、T4 DNA连接酶、SYBR Premix Ex Taq试剂盒购自TaKaRa公司(大连); DNA分子量标准GeneRuler™ 1 Kb DNA Marker、卡那霉素(kanamycin)和异丙基-β-D-硫代半乳糖苷(isopropyl-β-D-thiogalactoside, IPTG)等购自生工生物工程股份有限公司(上海); QIAquick Gel Extraction购自Qiagen公司(德国); Ficoll购自Invitrogen公司(上海); RPMI1640培养基、胎牛血清购自Gibco公司(美国); 引物合成及序列测定由Invitrogen公司(上海)完成; 二抗(辣根酶标记山羊抗小鼠IgG)购自中杉金桥生物技术有限公司(北京); ECL化学发

光试剂盒、显影定影试剂盒、柯达X-OMAT BT胶片和压片暗盒等购自碧云天生物技术研究所(北京)。

1.2 *PaGal3*基因cDNA序列测定及分析

从香鱼头肾来源的单核/巨噬细胞的转录组数据库中筛选Gal3相关Unigenes, 用BLAST(<http://blast.ncbi.nlm.nih.gov/blast.cgi>)分析进行功能注释, 随后用特异引物PCR扩增cDNA和序列测定的方法确认。蛋白分子量大小及等电点测定采用Compute pI/Mw程序(http://web.expasy.org/compute_pi/), *PaGal3*功能位点分析采用ScanProsite软件(<http://prosite.expasy.org/scanprosite/>), 多重序列比对采用Clustalw程序(<http://clustalw.ddbj.nig.ac.jp/>), 进化树构建采用MEGA version 5程序^[17]。多重序列比对及进化树构建涉及序列详见表1。

1.3 香鱼头肾来源的单核/巨噬细胞分离培养

麻醉香鱼, 无菌条件下快速取出头肾, 用剪刀将头肾剪碎置筛网内, 加入适量含2%胎牛血清的RPMI1640培养基, 用无菌玻璃注射器活塞轻柔研磨头肾组织, 获得分散的单细胞。采用Ficoll密度梯度离心法离心细胞悬液, 用含2%胎牛血清的RPMI1640培养基将所得细胞沉淀洗涤2次, 重悬于含2%胎牛血清的RPMI1640培养基中。24 h过夜培养后, PBS洗去非黏附细胞, 黏附细胞在含10%胎牛血清的RPMI1640培养基中培养。经姬姆萨染色后镜检, 确定95%的黏附细胞是单核/巨噬细胞^[18]。

1.4 鳗弧菌感染的香鱼单核/巨噬细胞及组织样品制备

过夜培养的鳗弧菌用PBS稀释后, 按照感染复

数(multiplicity of infection, MOI)为20:1的比例接种至香鱼单核/巨噬细胞中, 于4, 8, 12, 24 hpi(hours post infection)时去除培养基, PBS洗涤3次, 然后加入Trizol试剂裂解细胞, 5 min后收集裂解液作为香鱼单核/巨噬细胞样品, -70 °C超低温冰箱保存备用。

鳗弧菌感染香鱼的实验过程及感染剂量参考杨旦阳等^[16]的方法, 具体步骤如下: 感染组香鱼用1.0×10⁴ CFU/尾的浓度腹腔注射鳗弧菌悬液, 对照组香鱼注射等剂量的无菌生理盐水, 于4, 8, 12, 24 hpi时取肝、脾和头肾等组织立即投入液氮中, 随后转于-70 °C超低温冰箱保存。

1.5 实时荧光定量PCR(RT-qPCR)

提取香鱼肝、脾、头肾、单核/巨噬细胞总RNA, 并用DNase I(RNase-free)进行处理^[16]。取1 μg总RNA为模板, 以oligo(dT)₃₀为引物, 用AMV逆转录酶在42 °C作用1.5 h, 合成第一链cDNA。根据*PaGal3*序列设计特异性扩增引物: *PaGal3F*: 5'-ACC TCC ACA CCA TCA ACC AT-3', *PaGal3R*: 5'-TCC ATA GTT CCA GAG CCA CA-3'。香鱼看家基因*β-actin*作为内标(扩增引物对为: *pActinF*: 5'-TCG TGC GTG ACA TCA AGG AG-3', *pActinR*: 5'-CGC ACT TCA TGA TGC TGT TG-3')。RT-qPCR检测*PaGal3*基因mRNA在各组织器官中的表达变化。25 μL RT-qPCR反应体系包括cDNA模板0.5 μL、SYBR Premix Ex Taq(2×)缓冲液12.5 μL、上下游引物各1 μL, 蒸馏水补足体积至25 μL。扩增反应在ABI StepOne荧光定量PCR仪(ABI公司)上进行, 扩增程序为: 94 °C变性3 min; 94 °C 30 s, 58 °C 30 s, 72 °C 30 s, 36个循

表1 构建系统发育进化树采用序列

Table 1 Sequences used for the phylogenetic tree construction

登录号 Accession number	物种 Species		
	拉丁名 Latin name	中文名 Chinese name	英文名 English name
NP_002297	<i>Homo sapiens</i>	人	Human
XM_005746494	<i>Oreochromis niloticus</i>	尼罗罗非鱼	Nile tilapia
XP_704272	<i>Danio rerio</i>	斑马鱼	Zebrafish
KP100522	<i>Plecoglossus altivelis</i>	香鱼	Ayu
XM_004082078	<i>Oryzias latipes</i>	青鳉	Japanese ricefish
XM_008285540	<i>Stegastes partitus</i>	深裂眶锯雀鲷	Bicolor damselfish
XM_006798573	<i>Neolamprologus brichardi</i>	女王燕尾	Burundi fairy cichlid
XM_005746493	<i>Pundamilia nyererei</i>	慈鲷	Lake Victoria cichlid
XM_004539924	<i>Maylandia zebra</i>	斑马拟丽鱼	Zebra mbuna
CDQ93743	<i>Oncorhynchus mykiss</i>	虹鳟	Rainbow trout

环。最后对扩增产物进行熔解曲线分析(94 °C 30 s, 72 °C 60 s, 95 °C 30 s)。每个样品重复3次实验。使用程序MxPro 3.2读取荧光定量结果, 相对定量结果分析采用 $2^{-\Delta\Delta Ct}$ 法^[19]。

1.6 原核表达、抗血清制备及Western blot

根据 *PaGal3*基因的糖基识别功能域和原核表达载体pET-28a酶切位点, 设计原核表达引物为: PaGal3pF: 5'-CCA TAT GAT GTA TAA CCT CCC TCT CCA-3', PaGal3pR: 5'-GGA ATT CTC ACA CGT CGA TCA TGC TGG G-3'(下划线分别为添加的限制性内切酶 *Nde* I和*Eco* R I的识别序列)。以合成的第一链cDNA为模板, PCR扩增目标序列, 扩增产物用2%(w/v)的琼脂糖凝胶电泳分离。符合预期大小的目的片段经切胶后用QIAquick Gel Extraction Kit纯化, 经*Nde* I和*Eco* R I双酶切后, 插入到同样双酶切的原核表达载体中, 获得重组质粒pET28a-PaGal3。重组质粒随后转化大肠杆菌BL21 pLys E, IPTG诱导表达后, SDS-PAGE电泳分离, 考马斯亮蓝G-250染色检测目的蛋白的表达情况。重组蛋白样品准备以及小鼠抗血清制备参考文献[16]。

Western blot检测方法详见文献[16]。香鱼单核/巨噬细胞培养上清和重组蛋白样本经SDS-PAGE电泳分离后湿转法转移至硝酸纤维素膜(nitrocellulose membrane, NC膜)上, 将NC膜浸入含5%脱脂奶粉的PBS-T中, 4 °C封闭过夜; 加一抗, 37 °C摇床孵育2 h, PBS-T摇洗5次; 加入辣根过氧化物酶标记的山羊抗小鼠IgG, 37 °C摇床孵育1 h, PBS-T摇洗5次; ECL发光法进行显影, 胶片扫描后根据Quantity One软件进行灰度值计算, 分析实验结果。

1.7 PaGal3抗体封闭对香鱼单核/巨噬细胞吞噬能力的影响

采用流式细胞术检测PaGal3抗体封闭对香鱼单核/巨噬细胞吞噬能力的影响。24 °C条件下, 在香鱼单核/巨噬细胞中加入PaGal3抗体(Anti-PaGal3)培养30 min, 对照组加入等体积的非免疫小鼠血清(IsoIgG)或PBS, 然后加入FITC标记的大肠杆菌DH5α(简称为*E. coli*-FITC)共培养1 h, 去除上清, 用PBS洗去未吞噬细菌, 台盼蓝淬灭细胞外荧光。用流式细胞仪(Beckman Coulter, Fullerton, CA, USA)检测吞噬的荧光信号。

1.8 PaGal3抗体封闭对香鱼单核/巨噬细胞杀菌活性的影响

采用TCBS平板计数法检测PaGal3抗体封闭对

香鱼单核/巨噬细胞杀菌活性的影响。PaGal3抗体封闭PaGal3的方法见本文1.7。抗体封闭结束后, 加入含10%胎牛血清的RPMI1640培养基, 按MOI为20:1的比例将鳗弧菌加入细胞中。培养30 min后, 一组细胞(uptake组)去除上清, 用PBS洗涤去未吞噬细菌, 细胞刮刀刮下贴壁细胞, 加入1% Triton X-100溶液裂解细胞, 将细胞裂解液梯度稀释后涂布于TCBS平板; 另一组细胞(kill组)去除上清, PBS洗涤去未吞噬细菌, 加入新的培养基, 继续培养1.5 h, 刮下贴壁细胞, 裂解后梯度稀释涂布TCBS平板。TCBS平板均置于28 °C培养20 h, 计算细菌菌落形成单位(colony forming unit, CFU)。将kill组的鳗弧菌CFU除以uptake组的鳗弧菌CFU, 计算鳗弧菌存活率。

1.9 统计分析

实验结果表示为平均值±标准误(mean±S.D.), 采用SPSS 13.0软件中的单因素方差分析(One-way ANOVA)进行统计, $P<0.05$ 为差异具有显著性。

2 结果

2.1 PaGal3基因cDNA序列分析

转录组测序和随后的特异引物PCR扩增测序揭示, *PaGal3*基因cDNA序列长2 009个核苷酸, 包含一个1 143个核苷酸长的大开放阅读框架, 推测其编码一个由380个氨基酸组成、相对分子质量为38.28 kDa的蛋白, 等电点为4.58, N-末端无信号肽序列。多重氨基酸序列比对显示, PaGal3具有典型的Gal结构特征, 含有一个C-端CRD(图1), 鱼类Gal3序列高度保守, 但与人Gal3差异较大。另外, 人Gal3中已知存在一个与其成熟肽活性有关的基质金属蛋白酶剪切位点(Ala⁶²-Tyr⁶³), 但这一位点在鱼类Gal3中并不存在(图1)。

序列分析表明, PaGal3与虹鳟Gal3氨基酸序列同一性最高(达77%), 与其他鱼类Gal3的氨基酸序列同一性为68%~75%, 而与哺乳动物Gal3的氨基酸序列同一性仅为48%。系统进化树分析也揭示, PaGal3与虹鳟的Gal3进化关系最近(图2)。

2.2 感染鳗弧菌对免疫组织和细胞 *PaGal3*基因 mRNA的表达影响

香鱼鳗弧菌感染4~8 h开始出现游动异常、保持平衡困难等症状, 12 h出现肠道、鳃盖和鱼鳍基部充血发红等症状, 24 h出现病鱼死亡, 而注射生理盐水的对照组健康香鱼表现正常。因此, 在4, 8, 12, 24 hpi, 我们收集鳗弧菌侵染香鱼肝、肾和脾组织,

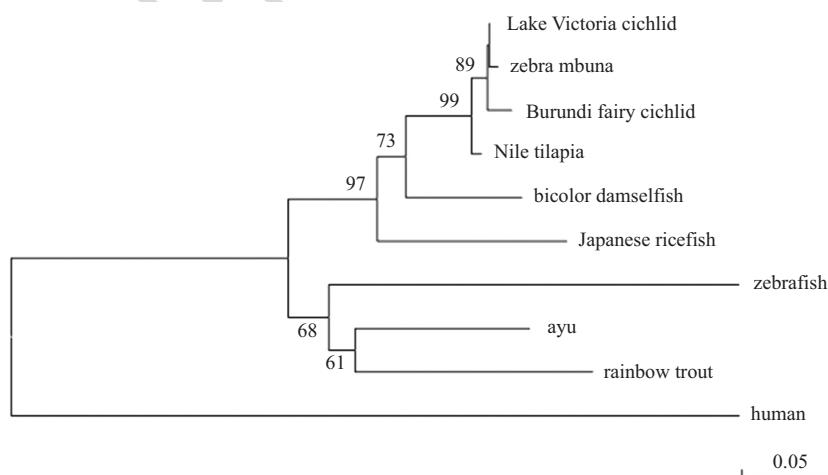
ayu	-TAA-TSLADAISDNDPQDQPRNGNINPSAIGNIGAPTAINPGPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	87
rainbow trout	-MD-HSLADAIDDPVPSQAKRASMNPTAATP-PDPINPGPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	81
zebrafish	-MD-HSLADAIDDPVPSQAKRASMNPTAATP-PDPINPGPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	81
Lake Victoria cichlid	MIDDD-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	82
zebra mbuna	MIDDD-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	82
Burundi fairy cichlid	MIDDD-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	80
Nile tilapia	MIDDD-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	82
bicolor damselfish	-MSP-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	80
Japanese ricefish	-MAD-HSLADALGDDITTSQAKKIGNNPNSAPTSNIAAPSNICPSSAPGAPTQPSAAGGFLGCPMPSGPGAPGQWGSQGPFPSPGPAP	78
human	-MADNEHSUHDAISGS-GNPNP-QCPW-GAW-NQPA-GAGGYG-PAS-YIGAMF-GQAPP	51
ayu	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	158
rainbow trout	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	171
zebrafish	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	148
Lake Victoria cichlid	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	152
zebra mbuna	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	152
Burundi fairy cichlid	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	150
Nile tilapia	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	152
bicolor damselfish	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	149
Japanese ricefish	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	148
human	GQYHAPSAESFGPPGPGIPGQYIPSPGPGAPGGFPSPGPAGPGQHFG-QYPPG-QDPPGEPGPMPYPSGVVIGPEP-S	82
ayu	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	240
rainbow trout	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	258
zebrafish	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	231
Lake Victoria cichlid	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	240
zebra mbuna	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	240
Burundi fairy cichlid	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	238
Nile tilapia	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	240
bicolor damselfish	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	238
Japanese ricefish	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	235
human	SPGAPPGPG-DP-QPGG-CMYGPGGPGQVEPPAAGCFCIFPAFFPGNAPPIPPGSWGLGGCQGFPQFCGIPSPGPMPGPGYGGAA	109
ayu	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	321
rainbow trout	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	345
zebrafish	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	311
Lake Victoria cichlid	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	321
zebra mbuna	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	321
Burundi fairy cichlid	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	319
Nile tilapia	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	319
bicolor damselfish	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	319
Japanese ricefish	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	316
human	PGGMUWVFYLPLHAGIMPNLIMTVVGEPIEGADRIIHDFTIKGD-VVHFHNPRFQEQT-VVRNSLQCVNGPEERDGFPV	193
ayu	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	380
rainbow trout	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	402
zebrafish	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	368
Lake Victoria cichlid	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	378
zebra mbuna	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	378
Burundi fairy cichlid	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	376
Nile tilapia	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	378
bicolor damselfish	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	376
Japanese ricefish	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	373
human	PGRRFELKILVEEDFKVAVDGHILLEYEHRVGGDFDVLILLRWGDNLYSAPSMLDV	250

图中的方框代表糖基识别功能域。**标识处为人Gal3的基质金属蛋白酶特异性剪切位点。

The box showed the CRD domain. ** showed the human matrix metalloproteinase-specific cleavage site.

图1 香鱼与其他物种Gal3全长氨基酸序列多重比对结果

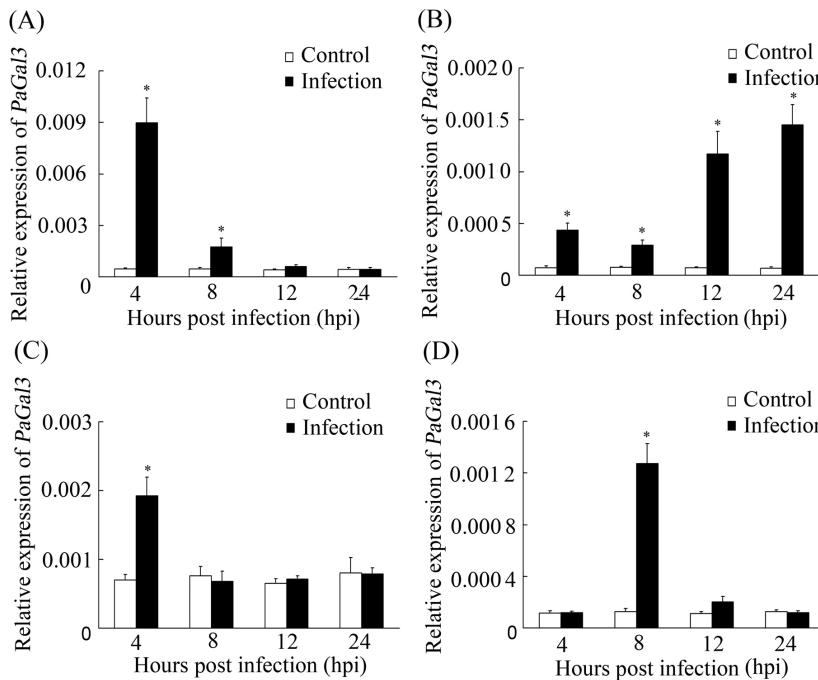
Fig.1 Multiple alignment of ayu Gal3 amino acid sequence with those of other species



人Gal3序列作为外群,分叉上的数值表示1 000次成簇中该进化簇的出现次数,仅置信度>60%的显示。标尺长度表示每一位点发生0.05次置换。Human Gal3 sequence is used as the outgroup. The values at the forks indicated the percentage of trees in which this grouping occurred after bootstrapping (1 000 replicates; shown only when confidence level>60%). The scale bar showed the number of substitutions per base.

图2 基于NJ法构建的香鱼及其他物种Gal3全长氨基酸序列的系统进化树

Fig.2 Phylogenetic tree analysis of Gal3 amino acid sequences of ayu and other species using Neighbor-Joining method



A: 肝; B: 头肾; C: 脾; D: 单核/巨噬细胞。以 *PaGal3* 与香鱼 β -actin mRNA 的比值作为 *PaGal3* mRNA 的相对表达量, * $P<0.05$, 与对照组相比。n=4。
A: liver; B: head kidney; C: spleen; D: monocytes/macrophages. Relative expression of *PaGal3* mRNA is normalized against β -actin mRNA. * $P<0.05$ compared with control group. n=4.

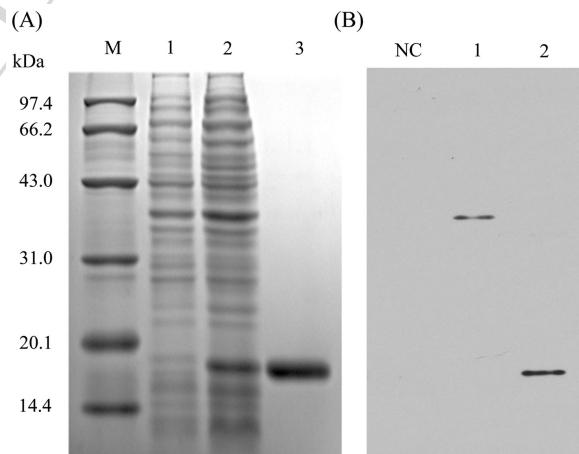
图3 实时荧光定量PCR分析 *PaGal3* 基因 mRNA 在感染鳗弧菌的香鱼免疫组织和单核/巨噬细胞中的表达变化

Fig.3 *PaGal3* mRNA expression changes in immune tissues and monocytes/macrophages in ayu upon *V. anguillarum* infection by RT-qPCR analysis

以及鳗弧菌感染的体外培养的头肾来源单核/巨噬细胞样品, RT-qPCR 分析其 *PaGal3* 基因 mRNA 的表达变化。结果表明, 4 hpi 时, 香鱼肝组织中 *PaGal3* 基因 mRNA 表达急剧上调, 随后降低, 12 hpi 时恢复正常对照值; 头肾组织各时间点样品中 *PaGal3* 基因 mRNA 表达均表现为显著上调, 24 hpi 达到峰值; 脾组织 4 hpi 时 *PaGal3* 基因 mRNA 表达显著上调, 随后回落到正常对照值(图3); 鳗弧菌感染的香鱼头肾来源单核/巨噬细胞在 8 hpi 时 *PaGal3* 基因 mRNA 表达显著上调, 其他时间点变化不明显(图3)。

2.3 *PaGal3* 原核表达、抗血清制备及 Western blot 验证

转化有 pET28a-PaGal3 表达质粒的大肠杆菌 BL21 pLys E 经 IPTG 诱导表达、SDS-PAGE 分离菌体蛋白及考马斯亮蓝 G-250 染色后, 胶上出现一条分子量约为 17 kDa 的蛋白诱导表达条带, 与预期大小相吻合(17.5 kDa)(图4A)。蛋白条带切胶纯化后免疫注射小鼠, 制备抗血清。Western blot 分析表明, 制备的香鱼 Gal3 抗血清能与重组蛋白特异性杂交, 而与大肠杆菌自身蛋白无作用; 另外, 在香鱼原代培养单核/巨噬细胞上清中检测到分子量为 38 kDa 左右的特异性

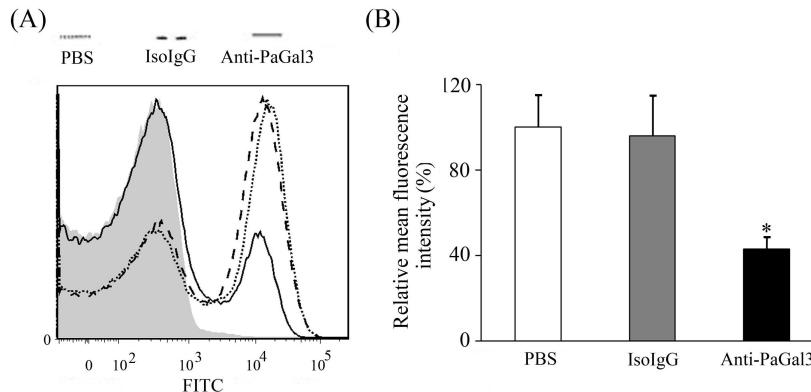


A: *PaGal3* 基因原核表达的 SDS-PAGE。M: 蛋白分子量标准(kDa); 1: BL21 菌体蛋白, 经 IPTG 诱导; 2: pET28a-PaGal3/BL21, 经 IPTG 诱导; 3: 切胶纯化的诱导蛋白; B: PaGal3 抗体 Western blot 验证。NC: 阴性对照(BL21 菌体蛋白); 1: 香鱼单核/巨噬细胞培养上清; 2: 阳性对照(pET28a-PaGal3/BL21, 经 IPTG 诱导)。

A: SDS-PAGE analysis of prokaryotic expression of *PaGal3* gene. M: protein molecular weight standards (kDa); 1: BL21 with IPTG induction; 2: pET28a-PaGal3/BL21 after IPTG induction; 3: purified PaGal3; B: Western blot analysis of PaGal3 antibody. NC: negative control (BL21); 1: the culture supernatant of ayu monocytes/macrophages; 2: positive control (pET28a-PaGal3/BL21 after IPTG induction).

图4 *PaGal3* 基因的原核表达和制备的 PaGal3 抗血清的 Western blot 验证

Fig.4 Prokaryotic expression of *PaGal3* gene and confirmation of prepared PaGal3 antiserum by Western blot

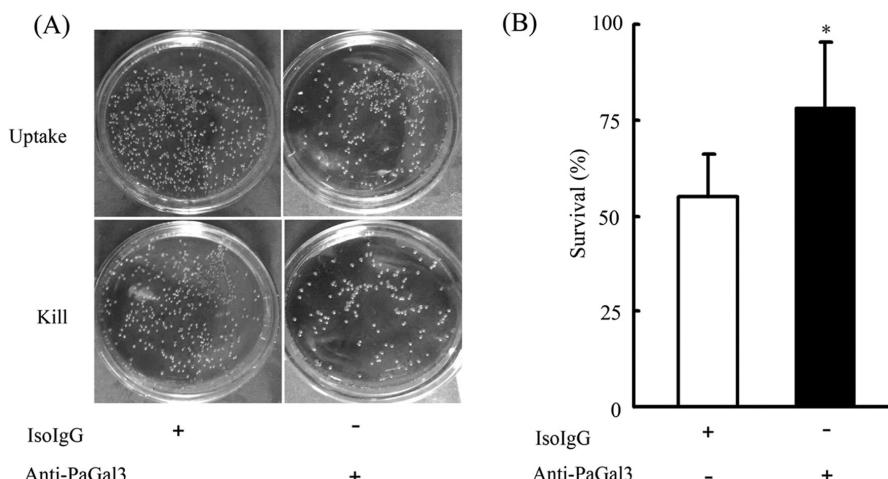


A: 吞噬的流式柱状图; B: 相对荧光密度。PBS对照组的相对荧光密度设为100%。*P<0.05, 与对照组(PBS和IsoIgG)相比。n=3。

A: flow cytometry histogram represents the phagocytosis; B: relative fluorescence intensity. The relative fluorescence intensity of the PBS control group is set to be 100%. *P<0.05 compared with PBS group and IsoIgG group. n=3.

图5 PaGal3抗体封闭对香鱼单核/巨噬细胞吞噬能力的影响

Fig.5 Effect of PaGal3 antibody blockage on the phagocytic activity of ayu monocytes/macrophages



A: TCBS平板上的鳗弧菌; B: 鳗弧菌存活率。*P<0.05, 与对照组(IsoIgG)相比。n=3。

A: *V. anguillarum* on the TCBS plates; B: survival rates of *V. anguillarum*. *P<0.05 compared with IsoIgG group. n=3.

图6 PaGal3抗体封闭对香鱼单核/巨噬细胞杀菌活性的影响

Fig.6 Effect of PaGal3 antibody blockage on the bacterial killing activity of ayu monocytes/macrophages

条带, 与预测的PaGal3蛋白分子量大小一致(图4B)。

2.4 PaGal3抗体封闭对香鱼单核/巨噬细胞吞噬能力的影响

采用流式细胞术检测PaGal3抗体封闭对香鱼单核/巨噬细胞吞噬能力的影响。结果表明, 与对照组(PBS和IsoIgG)相比, PaGal3被抗体封闭后, 吞噬DH5 α 的单核/巨噬细胞数量明显减少(图5A), 且单核/巨噬细胞吞噬E. coli-FITC的平均荧光强度也明显下降(图5B)。

2.5 PaGal3抗体封闭对香鱼单核/巨噬细胞杀菌活性的影响

采用TCBS平板计数法检测PaGal3抗体封闭对香鱼单核/巨噬细胞杀菌活性的影响。结果表明, 与对照组(IsoIgG)相比, PaGal3被抗体封闭后, 香鱼单

核/巨噬细胞杀菌活性显著降低(图6)。

3 讨论

*Gal3*基因广泛存在于脊椎动物中, 部分鱼类中*Gal3*基因序列也已登录数据库, 但尚未见有相关研究报道。本文从香鱼单核/巨噬细胞转录组测序结果中获得*Gal3*基因的cDNA序列, 序列分析表明, 它与虹鳟*Gal3*最相近, 具有动物*Gal*的典型结构特征, C-末端具有一个CRD。人*Gal3*具有一个MMP剪切位点($\text{Ala}^{62}\text{-Tyr}^{63}$), 经此剪切能增强其与糖链分子结合的能力^[20], 但鱼类*Gal3*均无该位点序列。鱼类和人*Gal3*此剪切位点序列的差异表明, 鱼类*Gal3*的功能调控可能与哺乳动物存在差异。

目前, 多种鱼类的*Gal*家族其他成员已被克隆^[21],

例如, Kong等^[8]从鱂皱脾、肝和肌肉混合组织cDNA文库中获得Gal9基因,发现其在睾丸、脾、肠、胃和肝等组织高度表达,并且沙门氏菌(*Salmonella minnesota*)LPS刺激能诱导其mRNA在脾中的表达上调。Yang等^[21]从松江鲈鱼(*Trachidermus fasciatus*)肝组织cDNA文库中获得Gal样蛋白基因,发现其在卵巢、心脏、胃和皮肤高表达,并且在大肠杆菌LPS刺激下, Gal样蛋白基因mRNA在血细胞和皮肤中的表达显著上调。这些Gal家族成员在致病菌感染后的mRNA表达变化表明,其可能在鱼类免疫反应中起重要作用。但目前尚未有文献明确报道鱼类Gal3基因的表达特征。本研究中,鳗弧菌感染后,香鱼肝和脾组织中Gal3基因的mRNA表达量在感染4 h时显著上调,头肾组织中在感染各时间点均显著上调,单核/巨噬细胞中在感染8 h时显著上调,与上述研究结果较为一致,揭示PaGal3基因表达与香鱼炎症免疫反应紧密相关。

真核生物编码的分泌蛋白一般具有信号肽,信号肽可以引导新生的多肽通过内质网,最终被分泌到胞外^[22],然而哺乳动物Gal3无信号肽,但均可分泌至胞外。本研究揭示,PaGal3也没有N-末端信号肽序列,但它存在于香鱼单核/巨噬细胞上清中,揭示其可能与哺乳动物Gal3有相似的出胞机理^[23]。在哺乳动物中,胞外分泌的Gal3可以在由肝星状细胞(hepatocyte stellate cell, HSC)介导的吞噬、激活及随后的自分泌和旁分泌中起重要作用,从而介导肝纤维化^[24]。新近研究表明,哺乳动物Gal3能增强炎性细胞功能,如增强吞噬、分泌细胞因子及延长中性粒细胞寿命等^[25-27],与M1型巨噬细胞表型的维持相关^[12],M1型巨噬细胞表型由哺乳动物巨噬细胞极化而成,能够分泌多种促炎症因子且有杀菌功能^[28]。此外,人Gal3还可以通过激活中性粒细胞产生超氧化物,促进巨噬细胞的吞噬功能,并充当单核/巨噬细胞的趋化因子^[12]。本研究发现,用抗体封闭香鱼单核/巨噬细胞培养上清中的PaGal3后,香鱼单核/巨噬细胞的吞噬能力和杀菌活性均显著下调,揭示鱼类Gal3能增强单核/巨噬细胞的功能,可能调控M1型巨噬细胞表型分化,但具体机制如何,尚有待进一步的研究。

综上,本研究解析了PaGal3基因和编码蛋白的氨基酸序列特征,明确了其基因表达特征,证实了PaGal3具有增强单核/巨噬细胞吞噬和杀菌的能力,为后续深入研究PaGal3的功能奠定了基础。

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